

UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL
FACULDADE DE FARMÁCIA
TRABALHO DE CONCLUSÃO DE CURSO DE FARMÁCIA

**TEN YEARS OF UNPREDICTABLE CHRONIC STRESS RESEARCH IN
ZEBRAFISH: A SYSTEMATIC REVIEW AND META-ANALYSIS**

MATHEUS GALLAS LOPES

PORTO ALEGRE, 2022

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Trabalho de Conclusão de Curso
apresentado ao Curso de Farmácia da
Universidade Federal do Rio Grande do Sul
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Orientadora: Prof^ª. Dr^ª. Ana Paula Herrmann

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APRESENTAÇÃO

Esse Trabalho de Conclusão de Curso foi redigido sob a forma de artigo científico, o qual foi elaborado segundo as normas da revista *Biological Reviews*, apresentadas em anexo.

1 **Ten years of unpredictable chronic stress research in zebrafish: a systematic**
2 **review and meta-analysis**

3

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ABSTRACT

1
2 The zebrafish (*Danio rerio*) is a model animal that is being increasingly used in
3 neuroscience research. A decade ago, the first study on chronic unpredictable stress
4 (UCS) in zebrafish was published, inspired by protocols established for rodents. Since
5 then, several studies have been published by different groups, in some cases with
6 conflicting results. We conducted a systematic review to identify studies evaluating the
7 effects of UCS in zebrafish and meta-analytically synthesised the data of
8 neurobehavioral outcomes and relevant biomarkers. Literature searches were
9 performed in three databases (PubMed, Scopus and Web of Science) and a two-step
10 screening process based on inclusion/exclusion criteria. The included studies
11 underwent extraction of qualitative and quantitative data, as well as risk of bias
12 assessment. Outcomes of included studies (n = 38) were grouped into anxiety/fear-
13 related behaviour, locomotor function, social behaviour, cortisol levels, *bdnf*, or *crf*
14 expression domains. UCS increased anxiety/fear-related behaviour and cortisol levels
15 while decreased locomotor function, but no effects were found for social behaviour
16 and expression of *bdnf* and *crf*. Despite including a significant number of studies, the
17 high heterogeneity and the methodological and reporting problems evidenced in the
18 risk of bias analysis make it difficult to assess the internal validity of most studies and
19 the overall validity of the model. Our review thus evidences the need to conduct well-
20 designed experiments to better evaluate the effects of UCS on the behaviour of
21 zebrafish.

22
23 **Keywords:** Unpredictable chronic stress, *Danio rerio*, animal model, anxiety,
24 locomotor function, social behaviour, cortisol, systematic review, meta-analysis,
25 depression

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1 I. INTRODUCTION

2 The origins of the unpredictable chronic stress (UCS) protocol go back to the early
3 1980s, when researchers proposed the chronic administration of a variety of stressors
4 to rodents as a way to induce behavioural alterations relevant to the study of
5 depression (Katz & Hersh, 1981; Katz, Roth & Carroll, 1981; Katz, 1982; Willner *et al.*,
6 1987). Construct, face, and predictive validities of this model are supported by many
7 studies that show that rodents exposed to the UCS protocol develop anhedonia-like
8 behaviour, cognitive deficits, hormonal and neurochemical imbalances, weight loss,
9 and many other changes that can be reversed by using antidepressant treatments
10 (Willner, 1997). Given its translational potential, there has been an exponential growth
11 in the implementation of this protocol across laboratories as it has become an
12 important tool for the study of the neurobiological basis of depression and
13 antidepressant action (Willner, 2017a; Nollet, 2021).

14 Whereas this intervention became popular, researchers started adapting the
15 UCS protocol and reports of controversial data and reproducibility problems have also
16 increased (Strekalova & Steinbusch, 2009; Willner, 2017b; Antoniuk *et al.*, 2019). The
17 protocol has been largely criticized for its lack of reliability as many known elements
18 such as the training level of experimenters, the duration of the protocol, and animal
19 characteristics (species, strain, sex, and others) can introduce variability to the
20 intervention and influence the results (Willner, 2017b). Apart from that, even with
21 heterogeneous protocols, the UCS was able to replicate behavioural and physiological
22 alterations within and between labs, adding to the internal and external validity of the
23 model.

24 More than a decade ago, researchers made an effort to transpose this
25 intervention for studies using zebrafish (*Danio rerio* Hamilton, 1822), an emerging

1 model animal in the field of neuroscience at the time (Piato *et al.*, 2011). Cross-species
2 approaches are important tools to evaluate the validity of an intervention, and
3 translating the UCS protocol to zebrafish can help reduce species-specific biases
4 originating from studies conducted solely with rodents (Maximino *et al.*, 2015; Weber-
5 Stadlbauer & Meyer, 2019). In zebrafish, this protocol is also able to induce anxiety-
6 like behaviour and alterations in outcomes like locomotion, cognition, sociability,
7 cortisol levels, and in the mechanisms of defence against oxidative damage (Piato *et*
8 *al.*, 2011; Marcon *et al.*, 2016, 2018b; Bertelli *et al.*, 2021). But just as in the
9 experiments carried out with rats and mice, the heterogeneity between protocols
10 established in each laboratory has grown throughout the years as investigators
11 needed to adapt the procedures to different facilities or the outcomes of interest sought
12 in the studies. Such problems culminated in the publication of many discrepant results
13 for key outcomes to understand the impacts of UCS, like social behaviour, which was
14 shown to be altered in opposing directions depending on the duration of the protocol
15 (Piato *et al.*, 2011), or not altered at all (Golla, Østby & Kermen, 2020; Bertelli *et al.*,
16 2021).

17 Aiming to estimate the overall validity and to summarise the evidence regarding
18 the effects of UCS on behavioural and biochemical outcomes relevant to the study of
19 psychiatric disorders, we conducted a systematic review and meta-analysis of the
20 available scientific literature using zebrafish. We analysed the evolution of this
21 intervention in the first ten years of its use, qualitatively describing the published
22 studies, establishing the direction of the effect of chronic stress on neurobehavioural
23 and neurochemical parameters, detecting patterns and effect moderators, and
24 evaluating the impact of bias arising from methodological conduct, reporting quality,
25 and selective publication.

1

2 **II. METHODS**

3 A protocol for conducting this review was registered on Open Science Framework prior
4 to the screening of records and data collection. Preregistration is available at
5 <https://osf.io/9rvyn> (Gallas-Lopes *et al.*, 2021). The reporting of this study complies
6 with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses
7 (PRISMA) guidelines (Page *et al.*, 2021).

8

9 **(1) Search strategy**

10 Searches were conducted in three bibliographic databases: PubMed, Scopus, and
11 Web of Science. The search strategy was designed to include broad terms that
12 describe the intervention (UCS protocol) and the desired population (zebrafish). The
13 complete query for each database can be found at <https://osf.io/9rvyn> (Gallas-Lopes
14 *et al.*, 2021). There were no language or date restrictions. The first search was
15 performed on the 10th of July, 2021, with an update search carried out on the 26th of
16 October, 2021. The bibliographic data acquired were imported to Rayyan software
17 (Ouzzani *et al.*, 2016), where duplicates were detected and removed by one of the
18 investigators (MGL). The reference lists of the included studies were also screened in
19 order to detect additional relevant articles.

20

21 **(2) Eligibility screening**

22 After the removal of duplicates, the selection of eligible studies was conducted using
23 Rayyann software in a two-step process based, initially, on title and abstract, followed
24 by a full-text analysis. The screening of each record was performed by two
25 independent investigators (MGL and LMB or RB) and disagreements were resolved

1 by a third investigator (APH). Peer-reviewed articles were eligible for inclusion if they
2 had an appropriate control group and assessed the effects of unpredictable chronic
3 stress in zebrafish (any strain or developmental stage) on any of the following domains
4 of interest: morphometric measures, locomotor function, sensory function, learning
5 and memory, social behaviour, reproductive behaviour, anxiety/fear-related
6 behaviour, circadian cycle-related behaviour, and neurochemical or peripheral
7 biomarkers (e.g., cortisol, cytokines, and oxidative stress).

8 In the first screening stage (title and abstract), studies were excluded based on
9 the following reasons: (1) design: not an original primary study (e.g., review,
10 commentary, conference proceedings, and corrections); (2) population: studies using
11 other species than zebrafish (*Danio rerio*) or studies that did not use any animal; (3)
12 intervention: non-interventional studies or studies using other interventions than
13 unpredictable chronic stress (e.g., acute stress (stressed only once) and repetitive
14 or predictable stress (chronic stress using only a single stressor multiple times)). In
15 the second stage (full-text screening), the remaining articles were assessed for
16 exclusion based on the same reasons considered in the first stage plus the following
17 additional reasons: (4) comparison: studies without an adequate control group; (5)
18 outcome: studies that did not evaluate any of the target outcomes. All Rayyan files
19 with investigators' decisions are available at the study repository in Open Science
20 Framework (<https://osf.io/j2zva/>), section "Eligibility screening archives".

21

22 **(3) Data extraction**

23 Data extraction from included studies was conducted by two independent investigators
24 (MGL and LMB or RB) and disagreements were resolved by a third investigator (APH).
25 Whenever available, the exact information and values were extracted directly from text

1 or tables. Otherwise, WebPlotDigitizer software (v4.5, Rohatgi, A., Pacifica, CA, USA,
2 <https://automeris.io/WebPlotDigitizer>) was used to manually estimate numbers from
3 the graphs. In cases of lacking or dubious information, investigators attempted to
4 contact via e-mail the corresponding author of the study in two separate attempts, at
5 least two weeks apart.

6 The following characteristics were extracted: (1) study characteristics: study
7 title, digital object identifier (DOI), first and last authors, last author's institutional
8 affiliation, and year of publication; (2) animal model characteristics: strain, sex, animal
9 source (supplier of the animals used to develop the experiments), the total number of
10 animals used, and the developmental stages during stress induction and outcome
11 assessment; (3) UCS protocol characteristics: the number of different stressors, stress
12 sessions per day, stress sessions in total, the duration of the stress protocol in days,
13 and the time in days between the end of UCS protocol and outcome assessment; (4)
14 test characteristics: experiment identification (to annotate whether the tests conducted
15 within the same study used different sets of animals), the type of the test, test duration,
16 habituation phase (whether the animals were subjected to an habituation phase in the
17 experimental apparatus prior to the test), the category of measured variable, and the
18 measured variable.

19 Outcome data were extracted for each of the variables within the domains of
20 interest. The measure of central tendency and the number of animals (n) were
21 extracted for the control and UCS groups along with the standard deviation (SD) or
22 standard error (SEM) when the mean value was expressed, or the interquartile range
23 (IQR) when data were expressed as the median value. Whenever sample size was
24 reported as a range instead of the exact number of animals in each group, the lowest

1 value was extracted. If the study reported the SEM, SD was calculated by multiplying
2 SEM by the square root of the sample size ($SD = SEM * \sqrt{n}$).

3

4 **(4) Bias assessment**

5 In order to evaluate the quality of included studies, the risk of bias assessment was
6 conducted by two independent investigators (MGL and LMB or RB) for each paper,
7 and disagreements were resolved by a third investigator (APH). The analysis was
8 conducted based on the SYRCLE's risk of bias tool for animal studies (Hooijmans *et*
9 *al.*, 2014) with adaptations to better suit the model animal and the intervention of
10 interest. The following items were evaluated for methodological quality: (1) description
11 of random allocation of animals; (2) description of baseline characteristics; (3)
12 description of random housing conditions during the experiments; (4) description of
13 random selection for outcome assessment; (5) description of blinding methods for
14 outcome assessment; (6) incomplete outcome data; (7) selective outcome reporting.
15 Additionally, four other items were evaluated by the investigators to assess the overall
16 reporting quality of the studies based on a set of reporting standards for rigorous study
17 design (Landis *et al.*, 2012): (8.1) mention of any randomization process; (8.2) sample
18 size estimation; (8.3) mention of inclusion/exclusion criteria; (8.4) mention of any
19 process to ensure blinding during the experiments. For methodological quality, each
20 item was scored with a "Yes" for low risk of bias, "No" for a high risk of bias or "Unclear"
21 when it was not possible to estimate the risk of bias based on the information provided.
22 Items regarding reporting quality were scored with only "Yes" or "No", meaning high
23 or low risk of bias, respectively. A complete guide for assessing the risk of bias
24 associated with each of the items in this review is available at <https://osf.io/sdpwb>.
25 Risk of bias plots were created using *robvis* (McGuinness & Higgins, 2021).

1 Publication bias was investigated by generating funnel plots and performing Egger's
2 regression test (Egger *et al.*, 1997). Analyses were only conducted when at least five
3 studies were available within a given domain for funnel plots and at least ten studies
4 for the regression test. A p -value < 0.1 was considered significant for the regression
5 test.

6

7 **(5) Meta-analysis**

8 Studies were grouped based on the domains of interest (anxiety/fear-related
9 behaviour, locomotor function, social behaviour, cortisol levels, *bdnf* expression, or *crf*
10 expression), and a meta-analysis was performed for each group. When a study
11 reported multiple outcomes for the same domain, only one outcome of interest was
12 chosen for the meta-analysis based on a rank of frequency developed by one of the
13 investigators (MGL). Tests and variables within each test were ranked prior to data
14 extraction, and the most frequent in the rank was included in the meta-analysis. The
15 ranking is available at <https://osf.io/rvn8b>. A minimum of five studies were required for
16 each domain in order to conduct a meta-analysis, as established a priori in our protocol
17 (Gallas-Lopes *et al.*, 2021).

18 The sample size of the control group was divided by the number of comparisons
19 and rounded down whenever two or more experimental groups shared the same
20 control (Vesterinen *et al.*, 2014). When outcomes were analysed across time, the last
21 point was selected for analysis. When animals were subjected to experiments at
22 different time points following the end of the UCS protocol, the outcomes assessed
23 closest to the end of the protocol were chosen. Effect sizes were "flipped" (multiplied
24 by minus one) when needed to adjust the direction of the effect for specific behavioural
25 traits in order to properly interpret the effects of UCS. Studies that only reported

1 outcomes as the median value and interquartile range were excluded from the
2 analyses along with studies with incomplete data (e.g., lacking sample sizes, SD, and
3 SEM) when contact with the authors was unsuccessful.

4 Effects sizes were determined with standardised mean differences (SMD) using
5 Hedge's G method. Analyses were conducted using JASP software version 0.16.3
6 (<https://jasp-stats.org>) with packages *metafor* (Viechtbauer, 2010) ([https://cran.r-](https://cran.r-project.org/package=metafor)
7 [project.org/package=metafor](https://cran.r-project.org/package=metafor)) and *ggplot2* (Wilkinson, 2011) following Hedge's
8 random effects model given the anticipated heterogeneity between studies. Values for
9 SMD were reported with 95% confidence intervals. Heterogeneity between studies
10 was estimated using both the I^2 and Chi^2 tests. Values of 25%, 50%, and 75% were
11 considered as representing low, moderate, and high heterogeneity, respectively for
12 the I^2 , and a p -value ≤ 0.1 was considered significant for the Chi^2 (Higgins &
13 Thompson, 2002). Furthermore, a subgroup meta-analysis was performed to evaluate
14 if the duration of the UCS protocol was a potential source of heterogeneity. Studies
15 were grouped into two categories: those with up to 7 days of UCS protocol and those
16 with more than 7 days. Subgroup analysis was only performed when there were at
17 least five unique studies for each subgroup.

18

19 **(6) Sensitivity analysis**

20 A sensitivity analysis was conducted in order to assess if any experimental or
21 methodological difference between studies was distorting the main effect found in the
22 meta-analysis. Analyses were conducted by excluding studies presenting a significant
23 risk of bias, defined as either a high risk of bias in one of the main items evaluating
24 methodological quality (items 1 to 7), or an unclear risk of bias in five or more of the

1 same items. A minimum of three comparisons were required for each domain in order
2 to conduct a sensitivity analysis.

3

4 **III. RESULTS**

5 **(1) Search results**

6 From the search in the selected databases, 420 records were retrieved altogether.
7 Following the removal of duplicates, 206 records were screened for eligibility based
8 on title and abstract. After the first screening phase, 58 reports remained to be
9 assessed based on full text, and 38 met the criteria and were included in the review
10 (Fig. 1). Out of the reports included in the review, 34 were collected from the first
11 database search on the 10th of July, 2021, and four additional reports were identified
12 in the second search on the 26th of October, 2021. No extra studies were identified by
13 reference list screening. Most of the records sought for inclusion in either stage of
14 screening were excluded because they did not meet the criteria set for the intervention
15 ($n = 89$), followed by the population of interest ($n = 42$), and the design of the study (n
16 $= 37$). Two studies were excluded from the quantitative analyses because the
17 minimum number of studies to perform a meta-analysis was not reached for the
18 outcome reported (Zimmermann *et al.*, 2016; Marcon *et al.*, 2018b), and four studies
19 were excluded because of missing information (Huang, Butler & Lubin, 2019; Zhang
20 *et al.*, 2021; Kirsten *et al.*, 2021; Demin *et al.*, 2021).

21

22 **(2) Study characteristics**

23 As expected, the protocols implemented by each research group varied significantly.
24 The duration of the stress protocol ranged between 3 and 77 days, with 15 studies
25 (39.5%) implementing UCS for up to 7 days, and 27 (71%) for more than a week.

1 Protocols using 7 ($n = 13$, 34.2%) or 14 days ($n = 12$, 31.6%) of UCS were the most
2 common. It is important to mention that some studies ($n = 5$, 13.2%) used UCS
3 protocols of more than 15 days to explore the more severe or long-term impacts of
4 UCS in zebrafish. The protocols were conducted using frequently a group of up to 10
5 different stressors to account for unpredictability. Outcome assessment usually took
6 place within the 24 hours following the last stress session ($n = 31$, 81.6%), with only a
7 few studies evaluating the effects of UCS after a longer washout period ($n = 10$,
8 26.3%). The tests were mostly scheduled to occur at least a day from the last stressor
9 to avoid the acute interference from the last stress session but also not too far off the
10 end of the protocol to avoid losing the effects of UCS.

11 The majority of studies were conducted by exposing adult zebrafish to the
12 protocol ($n = 34$, 89.4%), followed by fish in the larval ($n = 3$, 7.9%), and juvenile life
13 stages ($n = 1$, 2.6%). Of the publications implementing the UCS protocol in early
14 developmental stages, one of them evaluated behavioural data of the exposed
15 animals when animals were still larvae. The remaining were designed to assess the
16 long-lasting effects of the stress and, in this case, animals were tested more than 75
17 days after the protocol ended, when they were considered adults. Experiments were
18 conducted generally with a pool of both male and female zebrafish ($n = 21$, 55.2%). In
19 only two studies both male and female zebrafish were used and sex was analysed as
20 a biological variable, whereas in four papers animals of only one sex were selected (n
21 = 2 for male and $n = 2$ for female fish). The sex of the animals was not specified in 11
22 studies (28.9%). A description of the studies included in the review can be found in
23 Table 1, and the detailed extracted information is available at <https://osf.io/2jzw9>.

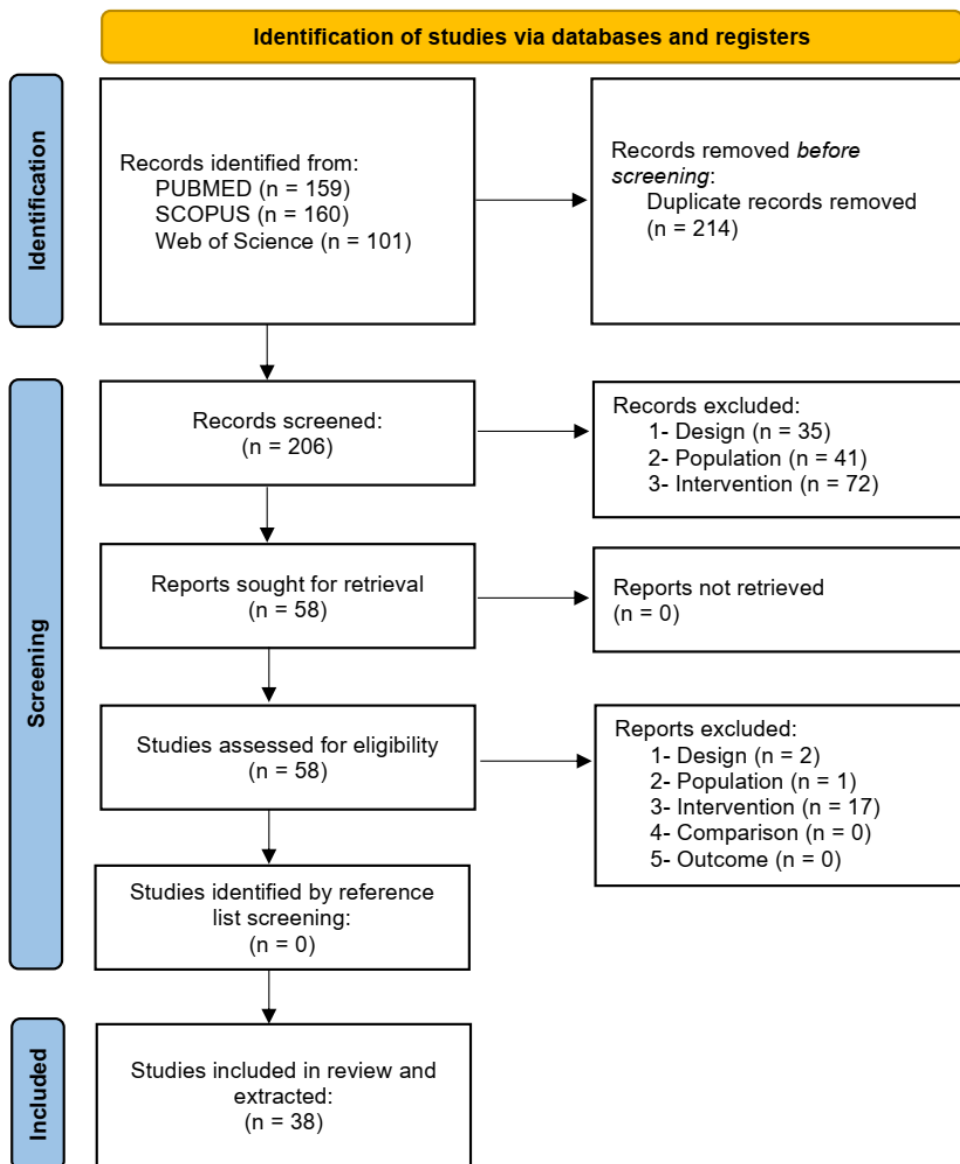


Fig 1. Flowchart diagram of the collection of studies and selection process.

1

2 **(3) Bias assessment**

3 The overall risk of bias associated with the items evaluated for methodological quality
 4 was considered unclear (Fig. 2). In more than 89% of the studies included, the
 5 information given was insufficient to rule out biases arising from the allocation of
 6 animals to the experimental groups or baseline characteristics. Although being an
 7 important methodological conduct, random housing allocation was not reported in any

1 publication. Bias related to blind assessment of outcomes was considered unclear in
 2 14 studies (36.8%) and one study was deemed as having a high risk of bias for this
 3 item. Outcome data was incomplete in two studies (5.3%), and it was unclear whether
 4 data was complete in 63.2% of the assessed papers. For six studies (15.8%), cross-
 5 checking the information for outcomes measured between the methodology and the
 6 results was not possible and selective reporting was considered unclear.

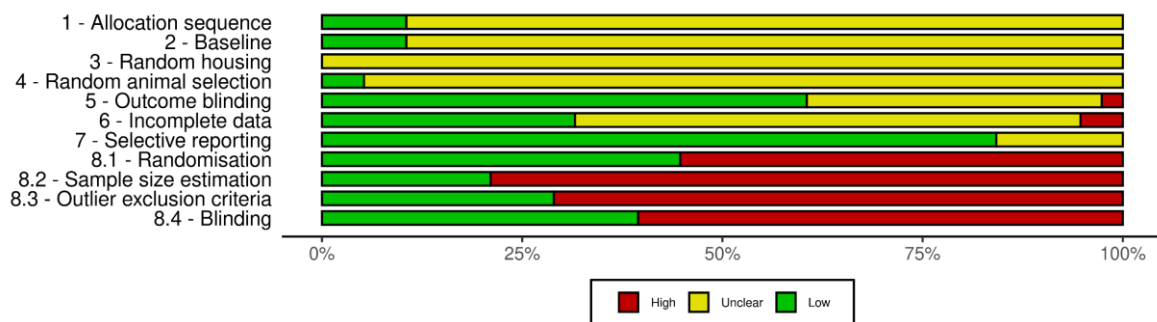


Fig. 2. Risk of bias assessment of included studies. The risk of bias assessment was performed by two independent investigators based on the SYRCLE’s risk of bias assessment tool. Items 1 to 7 account for methodological quality and were scored as presenting a high, unclear or low risk of bias. Items 8.1 to 8.4 evaluate the reporting quality of the studies and were scored as presenting a high or low risk of bias. Classification is given as the percentage of assessed studies ($n = 38$) presenting each score.

7

8 As for the reporting quality, more than 50% of the studies failed to report any
 9 information on the items assessed. Researchers failed to describe if any
 10 randomization method was used in 21 studies (55.3%). Sample size estimation
 11 procedures were not informed in 30 papers (78.9%). Reporting quality was also
 12 considered unsatisfactory when evaluating the report of inclusion/exclusion criteria
 13 and blinding, since there were no reports of these items in 27 (71.1%) and 23 (60.5%)
 14 of the studies, respectively. Out of 418 scores given in the risk of bias assessment,
 15 there were 51 (12.2%) inconsistencies between investigators. Individualised scores
 16 for each study included are available at <https://osf.io/zw6qg>.

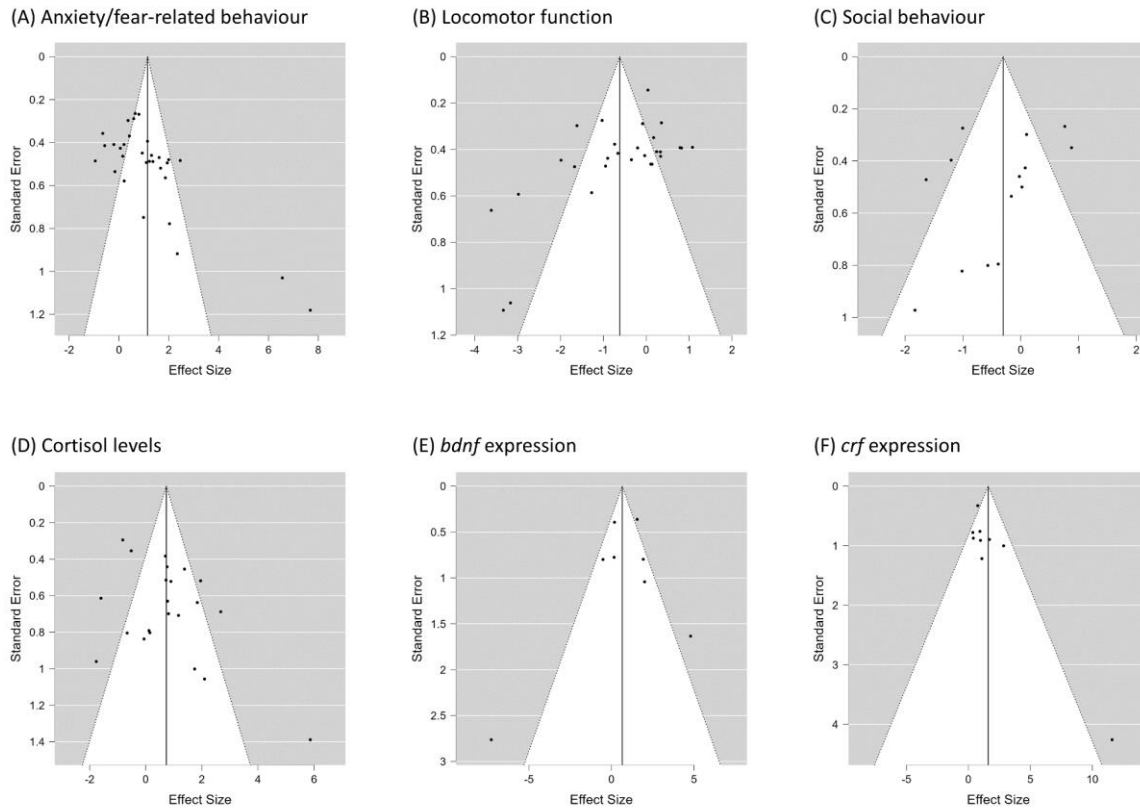


Fig 3. Funnel plots including studies analysed within each domain of interest: (A) anxiety/fear-related behaviour, (B) locomotor function, (C) social behaviour, (D) cortisol levels, (E) *bdnf* expression, and (F) *crf* expression. Each point represents a single comparison. The vertical line represents the overall effect size and the triangular region represents the 95% confidence interval.

1

2 Visual inspection of funnel plots demonstrated a substantial asymmetrical

3 distribution of the studies within some domains of interest (Fig. 3). The scattered plot

4 does not show the expected funnel-shaped distribution of experiments for anxiety/fear-

5 related behaviour (Fig. 3A), locomotor function (Fig. 3B), and social behaviour (Fig.

6 3C). This could be attributed to sample heterogeneity, as the protocols, tests, and

7 measured variables differ significantly among selected studies. On the other hand,

8 funnel plots for cortisol levels (Fig. 3D), *bdnf* expression (Fig. 3E), and *crf* expression

9 (Fig. 3F) show a relatively symmetrical distribution, with the limitation that the latter

10 two are based on a small number of studies.

1 Egger's regression test indicated publication bias for all domains tested (Table
 2 2): anxiety/fear-related behaviour ($p < 0.001$), locomotor function ($p < 0.001$), social
 3 behaviour ($p = 0.077$), and cortisol levels ($p = 0.086$). Both tests suggest a possible
 4 overestimation of the effects of UCS based on published data. Unfortunately, as
 5 mentioned above, studies reporting *bdnf* and *crf* expression were only a few, which
 6 hindered the inference of publication bias based on the regression test, as its statistical
 7 power depends on the number of experiments included in the analysis.

Table 2. Regression test for Funnel plot asymmetry ("Egger's test"). A p -value < 0.1 was considered significant for publication bias.

Domain	z	p-value
Anxiety/fear-related behaviour	5.440	< 0.001
Locomotor function	-4.036	< 0.001
Social Behaviour	-1.771	0.077
Cortisol levels	1.717	0.086

8

9 **(4) Anxiety/fear-related behaviour**

10 The meta-analysis comprised 31 comparisons out of 23 independent studies. A total
 11 of 377 animals were used as controls and 439 composed the stressed groups. The
 12 most frequently used test to assess anxiety/fear-related behaviour in the included
 13 studies was the novel tank (31), followed by the open field (3), light/dark (1), and
 14 stress-induced analgesia tests (1).

15 The overall analysis revealed that stressed animals have higher levels of
 16 anxiety/fear-related behaviour when compared to control animals (SMD 1.15 [0.52,
 17 1.78], $p < 0.001$, Fig. 4A). The estimated heterogeneity was high, with an $I^2 = 93.91\%$
 18 and a $\text{Chi}^2 = 169.092$ ($df = 30$, $p < 0.001$). Subgroup analysis revealed that for

1 experiments with stress duration of up to 7 days there was no statistically significant
 2 effect on anxiety/fear-related behaviour (SMD 0.37 [-0.22, 0.97], $p = 0.218$, Fig. 4B).
 3 The heterogeneity was also high for this subgroup, with an $I^2 = 82.69\%$, and a $\text{Chi}^2 =$
 4 51.242 ($df = 10$, $p < 0.001$). For experiments with a UCS regimen of more than 7 days,
 5 it is possible to observe a significant effect of the stress on increasing anxiety-like
 6 behaviour (SMD 1.61 [0.75, 2.48], $p < 0.001$, Fig. 4B). The heterogeneity remained
 7 high when analysing this subgroup, resulting in an $I^2 = 94.49\%$, and a $\text{Chi}^2 = 100.689$
 8 ($df = 19$, $p < 0.001$).

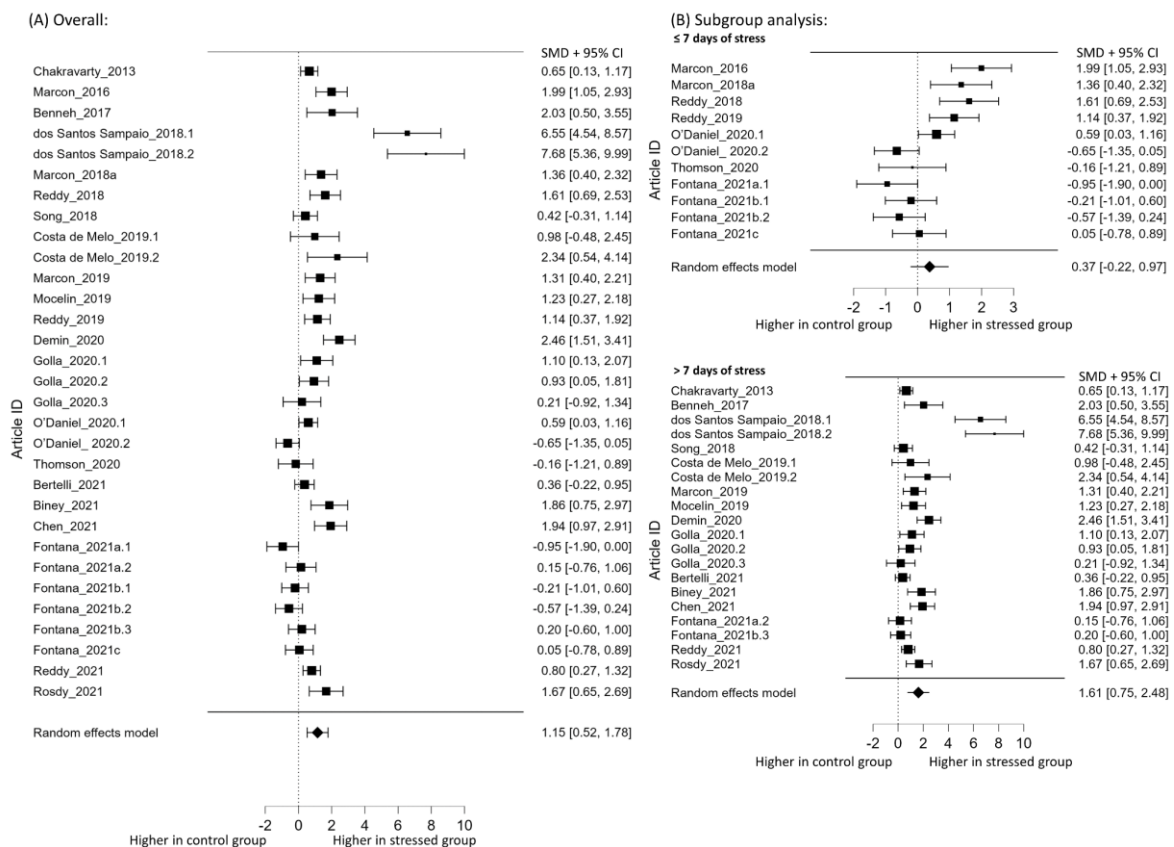


Fig 4. The effect of unpredictable chronic stress UCS protocol on anxiety/fear-related behaviour of zebrafish. (A) Overall effects of UCS on anxiety/fear-related behaviour in included studies. (B) Subgroup analyses based on the duration of the stress protocol (either ≤ 7 days or > 7 days of stress). Data are presented as Hedges' G standardised mean differences and 95% confidence intervals.

1 **(5) Locomotor function**

2 The meta-analysis comprised 28 comparisons out of 21 independent studies. A total
 3 of 454 animals were used as controls and 510 composed the stressed groups. The
 4 most frequently used test to assess locomotor function in the included studies was the
 5 novel tank test (21), followed by the open field (4), mirror-induced aggression (2), and
 6 stress-induced analgesia tests (1).

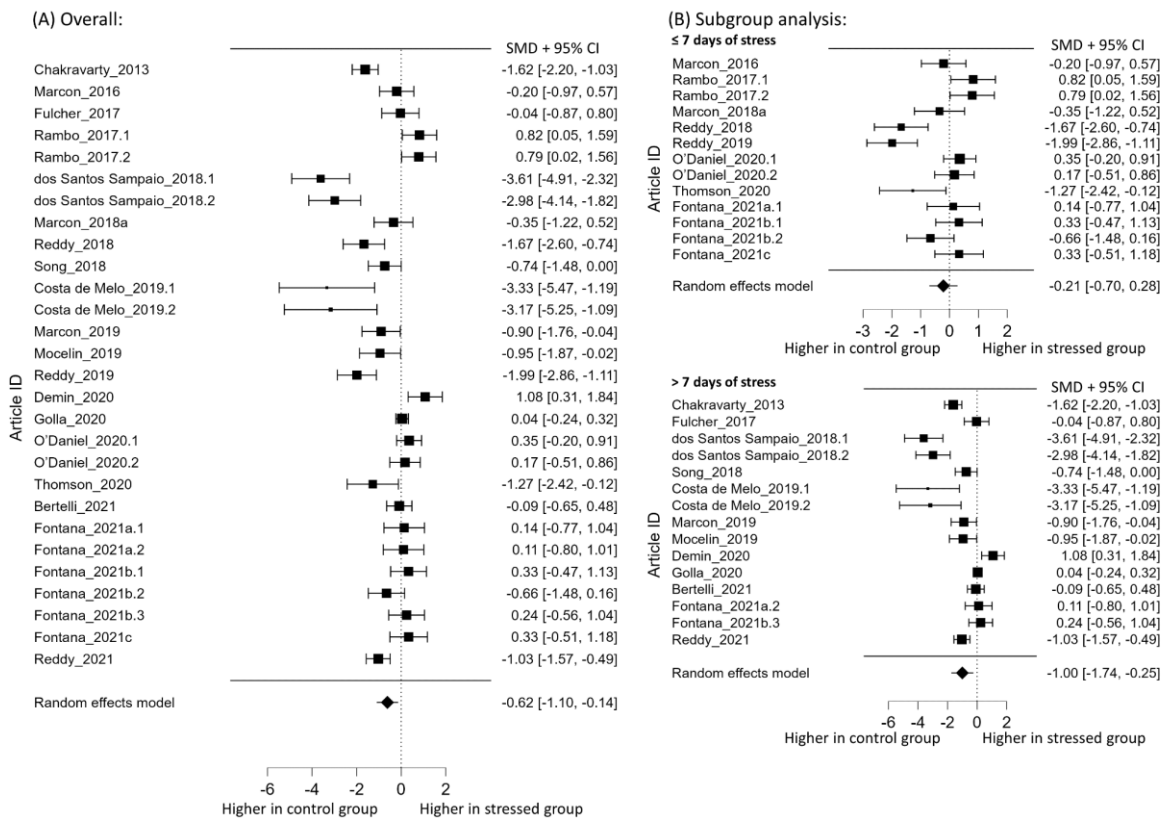


Fig 5. The effect of unpredictable chronic stress UCS protocol on the locomotor function of zebrafish. (A) Overall effects of UCS on the locomotor function in included studies. (B) Subgroup analyses based on the duration of the stress protocol (either ≤ 7 days or > 7 days of stress). Data are presented as Hedges' G standardised mean differences and 95% confidence intervals.

7 The overall analysis showed that stressed animals show lower levels of mobility
 8 when compared to control animals (SMD -0.62 [-1.10, -0.14], $p = 0.012$, Fig. 5A). The
 9 estimated heterogeneity was considered high, with an $I^2 = 91.30\%$ and a $\text{Chi}^2 =$
 10 168.198 ($df = 27$, $p < 0.001$). When analysing separately experiments conducted with

1 a UCS protocol of up to 7 days, there was no statistically significant effect of the stress
2 on locomotor function (SMD -0.21 [-0.70, 0.28], $p = 0.4$, Fig. 5B). The heterogeneity
3 was also high for this subgroup, with an $I^2 = 79.4\%$, and a $\text{Chi}^2 = 51.207$ ($df = 12$, $p <$
4 0.001). As for experiments conducted with a UCS regimen of more than 7 days, it is
5 possible to observe a significant difference in locomotor function between stressed
6 and unstressed groups, evidencing higher mobility for unstressed animals (SMD -1.00
7 [-1.74, -0.25], $p = 0.009$, Fig. 5B). The heterogeneity remained high when analysing
8 this subgroup, resulting in an $I^2 = 93.63\%$, and a $\text{Chi}^2 = 110.784$ ($df = 14$, $p < 0.001$).

9

10 **(6) Social behaviour**

11 The meta-analysis comprised 14 comparisons out of 11 independent studies. A total
12 of 172 animals were used as controls and 190 composed the stressed groups. The
13 most frequently used test to assess social behaviour in the included studies was the
14 shoaling response test (8), followed by social interaction (4), and novel tank tests (2).

15 The overall analysis showed no significant effects of the UCS protocol on social
16 behaviour (SMD -0.31 [-0.71, 0.10], $p = 0.140$, Fig. 6). The estimated heterogeneity
17 was considered moderate, with an $I^2 = 66.20\%$ and a $\text{Chi}^2 = 52.631$ ($df = 13$, $p < 0.001$).

18 There were no sufficient studies to perform a subgroup analysis.

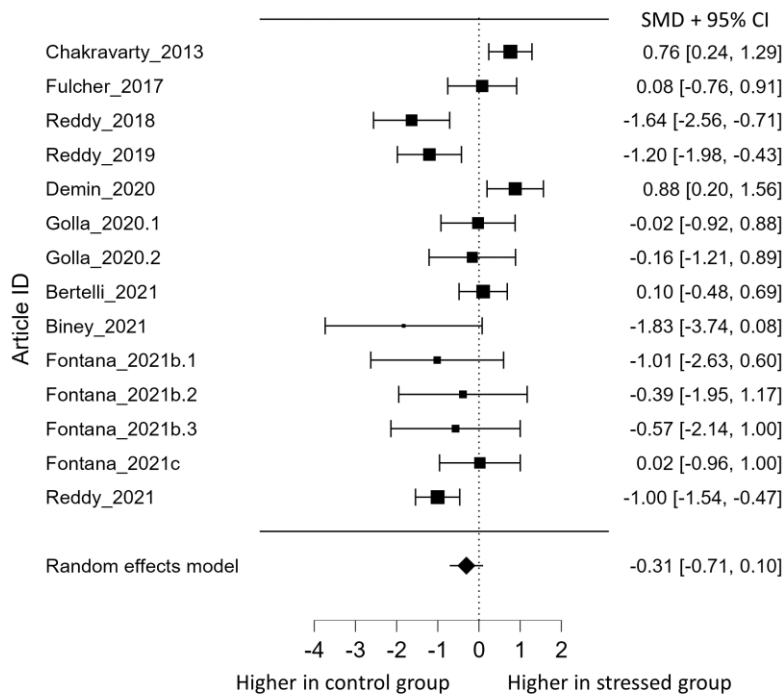


Fig 6. The effect of unpredictable chronic stress UCS protocol on the social behaviour of zebrafish. Data are presented as Hedges' G standardised mean differences and 95% confidence intervals.

1

2 (7) Cortisol levels

3 The meta-analysis comprised 22 comparisons out of 13 independent studies. A total
 4 of 150 animals were used as controls and 223 composed the stressed groups. Whole-
 5 body cortisol levels were measured in most studies (15), followed by trunk (5), and
 6 serum cortisol measurements (2).

7 The overall analysis showed that stressed animals have higher levels of cortisol
 8 when compared to control animals (SMD 0.73 [0.06, 1.40], $p = 0.032$, Fig. 7A). The
 9 estimated heterogeneity was considered high, with an $I^2 = 86.68\%$ and a $\text{Chi}^2 = 95.623$
 10 ($df = 21$, $p < 0.001$). When analysing separately experiments conducted with a UCS
 11 regimen of up to 7 days, there was no statistically significant effect of the stress on
 12 cortisol levels (SMD 0.82 [-0.21, 1.85], $p = 0.120$, Fig. 7B). The heterogeneity was also
 13 high for this subgroup, with an $I^2 = 91.05\%$, and a $\text{Chi}^2 = 87.613$ ($df = 13$, $p < 0.001$).

1 As for experiments conducted with a UCS protocol of more than 7 days, it is possible
 2 to observe a significant effect of the stress on increasing cortisol levels (SMD 0.69
 3 [0.24, 1.13], $p = 0.002$, Fig. 7B). The heterogeneity significantly decreased when
 4 analysing this subgroup, resulting in an $I^2 = 17.88\%$, and a $\text{Chi}^2 = 5.381$ ($df = 7$, $p =$
 5 0.614).

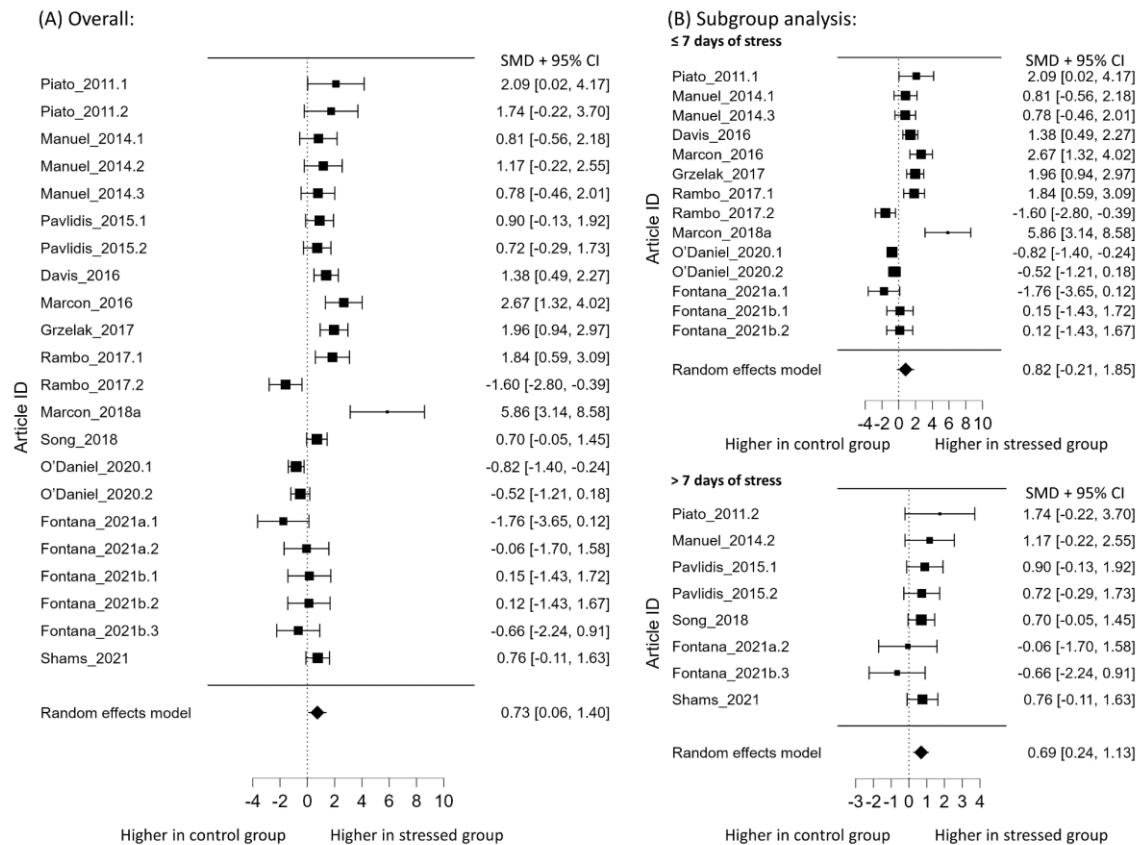


Fig 7. The effect of unpredictable chronic stress UCS protocol on cortisol levels in zebrafish. (A) Overall effects of UCS on the locomotor function in included studies. (B) Subgroup analyses based on the duration of the stress protocol (either ≤ 7 days or > 7 days of stress). Data are presented as Hedges' G standardised mean differences and 95% confidence intervals.

6 (8) *bdnf* and *crf* expression

7 The meta-analysis for *bdnf* expression comprised 8 comparisons out of 5 independent
 8 studies. A total of 45 animals were used as controls and 81 composed the stressed
 9 groups. The overall analysis showed no significant effects of the UCS protocol on the

1 expression of *bdnf* (SMD 0.65 [-1.74, 3.04], $p = 0.592$, Fig. 8A). The estimated
 2 heterogeneity was considered high, with an $I^2 = 95.76\%$ and a $\text{Chi}^2 = 27.967$ ($df = 7$,
 3 $p < 0.001$). There were no sufficient studies to perform a subgroup analysis. For the
 4 *crf* expression, the meta-analysis comprised 9 comparisons out of 5 independent
 5 studies. A total of 36 animals were used as controls and 73 composed the stressed
 6 groups. The overall analysis also showed no significant effects of the UCS protocol on
 7 the expression of *crf* (SMD 1.60 [-0.63, 3.82], $p = 0.159$, Fig. 8B). The estimated
 8 heterogeneity was considered high, with an $I^2 = 94.13\%$. On the other hand,
 9 heterogeneity was found to not be significant with a $\text{Chi}^2 = 11.865$ ($df = 8$, $p = 0.157$).
 10 Again, there were no sufficient studies to perform a subgroup analysis.

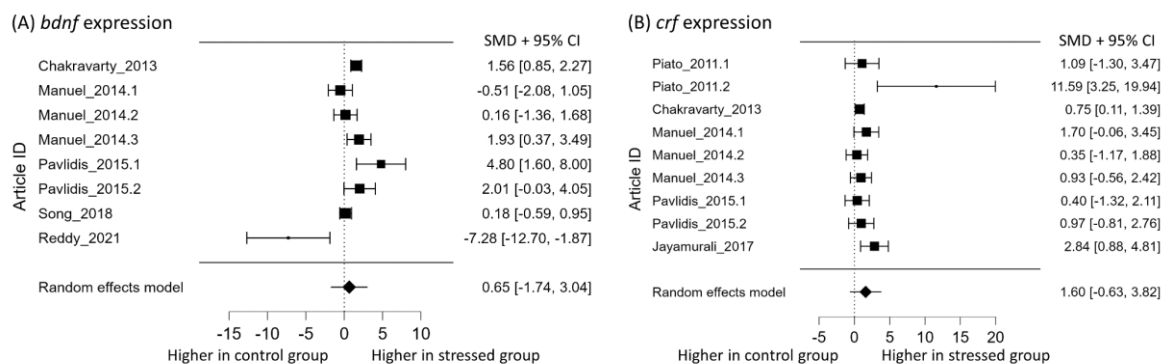


Fig 8. The effect of unpredictable chronic stress UCS protocol on the expression of *bdnf* and *crf* in zebrafish. (A) Meta-analysis of *bdnf* expression. (B) Meta-analysis of *crf* expression. Data are presented as Hedges' G standardised mean differences and 95% confidence intervals.

11

12 (9) Sensitivity analysis

13 The sensitivity analyses for studies presenting a significant risk of bias skewed the
 14 main effect of the domains tested (Fig 9). After excluding studies with a high risk of
 15 bias, no significant effects of UCS on anxiety/fear-related behaviour (SMD 1.07 [-0.13,
 16 2.28], $p = 0.081$, Fig. 9A) and locomotor function (SMD -0.44 [-1.13, 0.25], $p = 0.210$,
 17 Fig. 9B) were observed. For social behaviour, the overall interpretation remained the

1 same, with no significant effects of the intervention on this behaviour (SMD 0.15 [-
 2 0.20, 0.49], $p = 0.410$, Fig. 9C). For cortisol levels, on the other hand, by excluding
 3 studies associated with a high risk of bias the direction of the effect was reversed, as
 4 the meta-analysis evidenced higher levels of cortisol in the control animals when
 5 compared to the stressed groups (SMD -0.61 [-0.99, -0.23], $p = 0.002$, Fig. 9D). As all
 6 studies included in the *bndf* and *crf* expression domains were considered as having a
 7 high risk of bias, conducting this sensitivity analysis was not possible for these
 8 outcomes.

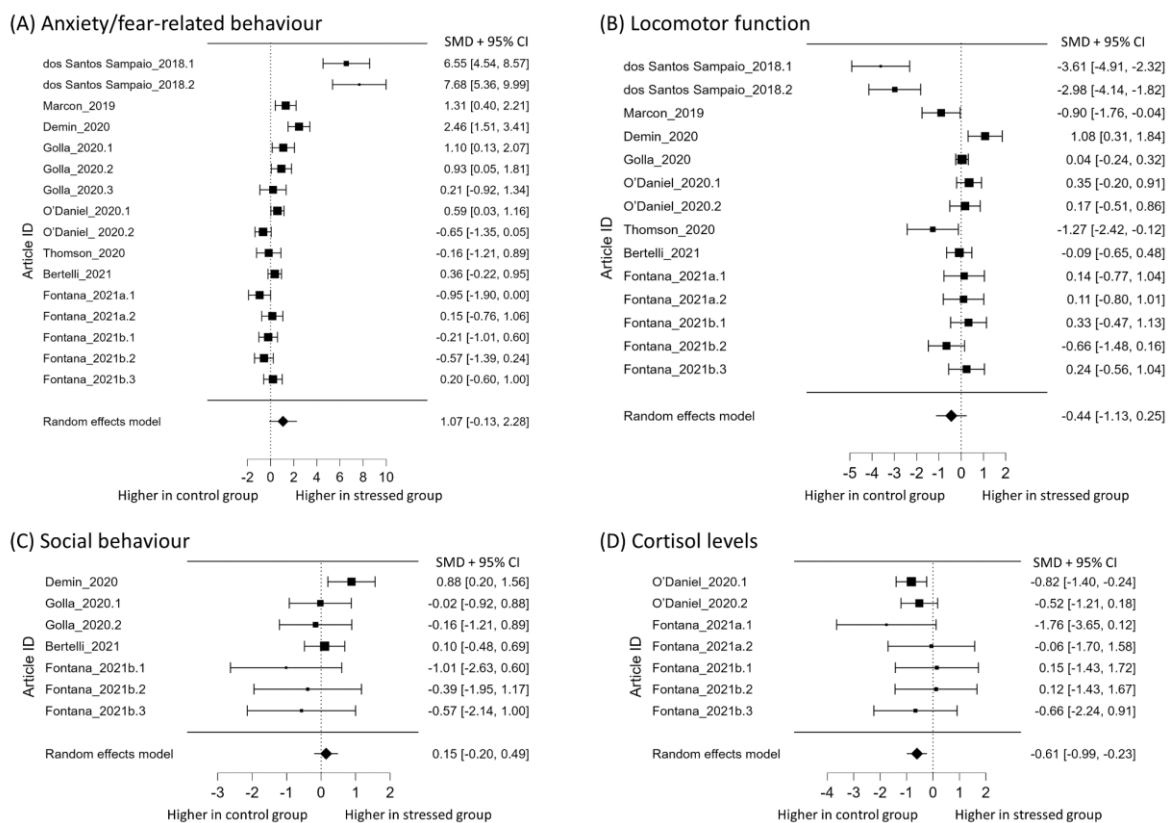


Fig 9. Sensitivity analyses for studies with a high risk of bias. The analyses were conducted by excluding studies presenting a significant risk of bias, defined as either a high risk of bias in one of the main items evaluating methodological quality in the risk of bias assessment (items 1 to 7), or an unclear risk of bias in five or more of the same items. Analyses were conducted for (A) anxiety/fear-related behaviour, (B) locomotor function, (C) social behaviour, and (D) cortisol levels. Data are presented as Hedges' G standardised mean differences and 95% confidence intervals.

Table 1. Qualitative description of studies reporting unpredictable chronic stress (UCS) protocols in research with zebrafish. The sex of the animals used was computed as: M, for male animals; F, for females; M:F, when male and female were included but tested and analyzed as a mixed group; M+F, when male and female fish were discriminated in the experiments; Unclear, for larvae and when the sex of the animals was not reported. Main findings were described as: ↑, higher when compared to the control group; ↓, lower when compared to the control group; =, no difference when compared to the control group.

Reference	Duration of stress protocol (days)	Number of different stressors	Interval between stress protocol and outcome assessment (days)	Developmental stage during stress/outcome assessment	Sex	Main findings
Piato <i>et al.</i> , 2011	7, 14	10	1	Adult	M	<p>Anxiety/fear-related behaviour ↓ Height in the tank</p> <p>Cortisol ↑ Whole-body cortisol</p> <p>Locomotor function ↓ Locomotion (14 days)</p> <p>Neurochemical outcomes ↓ <i>gr</i> expression ↑ <i>crf</i> expression</p> <p>Social behaviour ↑ Shoal cohesion (7 days) ↓ Shoal cohesion (14 days)</p>

Chakravarty <i>et al.</i> , 2013	15	10	1	Adult	M:F	<p>Anxiety/fear-related behaviour</p> <ul style="list-style-type: none"> ↑ Latency to upper zone ↓ Entries in the upper zone ↑ Freezing bouts ↑ Freezing duration ↓ Latency to dark compartment <p>Locomotor function</p> <ul style="list-style-type: none"> ↓ Crossings <p>Neurochemical outcomes</p> <ul style="list-style-type: none"> ↑ <i>crf</i> expression ↑ <i>ppp3r1a</i> expression ↑ <i>bdnf</i> expression <p>Social behaviour</p> <ul style="list-style-type: none"> ↓ Latency to together
Manuel <i>et al.</i> , 2014	7, 14	9	1	Adult	M:F	<p>Cortisol</p> <ul style="list-style-type: none"> ↑ Whole-body cortisol (14 days, 7 nights of UCS) <p>Learning and memory</p> <ul style="list-style-type: none"> ↓ Latency to black compartment day 2 (14 days of UCS) ↓ Latency to black compartment day 3 (14 days, 7 nights of UCS) <p>Neurochemical outcomes</p>

						<p>↑ <i>cart</i> expression (7 days of UCS) ↑ <i>htr1ab</i> expression (7 days of UCS) = <i>crf-bp</i> expression = <i>crf</i> expression ↑ <i>bdnf</i> expression (7 nights of UCS) ↑ <i>grβ</i> expression (7 nights of UCS) = <i>cnr1</i> expression ↑ <i>mr</i> expression (7 nights of UCS) ↑ <i>gra</i> expression (7 nights of UCS) = <i>mr/gra</i> ratio ↑ <i>grβ/gra</i> ratio (7 nights of UCS)</p>
Pavlidis, Theodoridi & Tsalafouta, 2015	11	7-12	1	Adult	M:F	<p>Cortisol ↑ Trunk cortisol concentration (Higher grade stressors)</p> <p>Neurochemical outcomes = <i>crf</i> mRNA relative levels ↑ <i>pomc</i> mRNA relative levels (Higher grade stressors) ↑ <i>gr</i> mRNA relative levels (Higher grade stressors) ↑ <i>mr</i> mRNA relative levels (Higher grade stressors) = <i>mc2r</i> mRNA relative levels ↑ <i>prl</i> mRNA relative levels (Higher grade stressors)</p>

						= <i>avt</i> mRNA relative levels ↑ <i>hypocretin/orexin</i> mRNA relative levels (Higher grade stressors) ↑ <i>bdnf</i> mRNA relative levels ↑ <i>c-FOS</i> mRNA relative levels
Davis <i>et al.</i> , 2016	5	5	Unclear	Adult	Unclear	Cortisol ↑ Serum cortisol Leukogram ↓ Lymphocytes ↑ Monocytes = Neutrophils = Eosinophils
Marcon <i>et al.</i> , 2016	7	7	1	Adult	M:F	Anxiety/fear-related behaviour ↓ Time in the upper zone ↓ Entries in the upper zone Cortisol ↑ Whole-body cortisol Locomotor function = Total distance travelled Neurochemical outcomes ↑ <i>cox-2</i> expression = <i>tnf-α</i> expression ↑ <i>IL-6</i> expression = <i>IL-10</i> expression

Zimmermann <i>et al.</i> , 2016	7	10	1	Adult	M	<p>Neurochemical outcomes</p> <p>↓ Membrane-bound Adenosine Deaminase</p> <p>= Cytosolic Adenosine Deaminase</p> <p>= <i>ada1</i> expression</p> <p>= <i>ada2.1</i> expression</p> <p>= <i>ada2.2</i> expression</p> <p>= <i>adal</i> expression</p> <p>= <i>adaasi</i> expression</p> <p>= ATP hydrolysis</p> <p>= ADP hydrolysis</p> <p>= AMP hydrolysis</p>
Benneh <i>et al.</i> , 2017	14	8	1, 3	Adult	Unclear	<p>Anxiety/fear-related behaviour</p> <p>↓ Time in the upper zone</p> <p>= Entries in the upper zone</p> <p>↑ Latency to upper zone (3 days post UCS)</p> <p>↓ Time spent in light region (1 day post UCS)</p> <p>↓ Entries in the light region (1 day post UCS)</p> <p>Social behaviour</p> <p>↓ Shoal average area (3 days post UCS)</p>
Fulcher <i>et al.</i> , 2017	15	6	1	Adult	M:F	<p>Anxiety/fear-related behaviour</p> <p>↓ Distance to bottom (1-3 minutes of test)</p>

						<p>↓ Freezing duration (1-3 minutes of test)</p> <p>Locomotor function ↑ Distance travelled (1-3, 6-10 minutes of test) ↑ Absolute turn angle (1-3, 11-15 minutes of test)</p> <p>Morphometric measurements ↑ Bodyweight</p> <p>Neurochemical outcomes = Dopamine levels = DOPAC levels = Serotonin levels = 5-HIAA levels</p> <p>Social behaviour = Distance to stimulus ↑ Variance of distance to stimulus (1-3 minutes of test)</p>
Grzelak <i>et al.</i> , 2017	10	5	Unclear	Adult	Unclear	<p>Cortisol ↑ Serum cortisol</p> <p>Leukogram ↓ Lymphocytes differential count ↑ Monocytes differential count</p>

						= Neutrophils differential count = Eosinophils differential count
Jayamurali & Govindarajulu, 2017	15	7	1	Adult	M:F	Neurochemical outcomes ↑ <i>crf</i> expression ↓ <i>gr</i> expression ↑ <i>p53</i> expression ↑ <i>NOXA</i> expression ↓ <i>bcl2</i> expression ↑ <i>casp3</i> expression
Rambo <i>et al.</i> , 2017	7	7	1	Adult	M+F	Aggression ↑ Relative time spent close to the mirror (male) Cortisol ↑ Whole-body cortisol (male) Locomotor function = Total distance travelled = Mean speed = Crossings
dos Santos Sampaio <i>et al.</i> , 2018	15	6	1	Adult	M:F	Anxiety/fear-related behaviour ↓ Time in the upper zone ↑ Latency to upper zone ↑ Freezing duration Locomotor function ↓ Total distance travelled

						↓ Quadrants crossed ↑ Erratic swimming
Marcon <i>et al.</i> , 2018a	7	7	1	Adult	M:F	Anxiety/fear-related behaviour ↓ Time in the upper zone ↓ Entries in the upper zone ↑ Time in the bottom Cortisol ↑ Trunk cortisol Locomotor function = Total distance travelled Neurochemical outcomes ↑ Reactive oxygen species (ROS) levels - DCF fluorescence
Marcon <i>et al.</i> , 2018b	7	6	1	Adult	M:F	Neurochemical outcomes ↑ TBARS levels ↑ Reactive oxygen species (ROS) levels - DCF fluorescence ↓ NPSH levels = SH total levels ↓ SOD activity = CAT activity
Reddy <i>et al.</i> , 2018	7	10	1	Adult	Unclear	Anxiety/fear-related behaviour ↓ Time spent in the upper zone ↑ Latency to upper zone

						<p>↑ Freezing duration</p> <p>Locomotor function ↓ Crossings</p> <p>Social behaviour ↓ Interaction time</p>
Song <i>et al.</i> , 2018	35	>10	1	Adult	M:F	<p>Anxiety/fear-related behaviour ↓ Time in the upper zone ↓ Entries in the upper zone = Freezing bouts</p> <p>Cortisol ↑ Whole-body cortisol</p> <p>Dendritic spines ↑ Average number of spines</p> <p>Locomotor function = Total distance travelled ↓ Mean meander moved = Low mobility duration = Low mobility frequency = Regular mobility duration = Regular mobility frequency = Highly mobility duration = Highly mobility frequency ↓ Mean velocity</p>

						<p>= Mean maximal velocity</p> <p>Neurochemical outcomes = <i>bdnf</i> expression = <i>p75</i> expression = <i>trkB</i> expression = <i>gfap</i> expression</p> <p>Peripheral outcomes ↑ Whole-body <i>IL-1β</i> ↑ Whole-body <i>IL-6</i> ↑ Whole-body <i>IL-10</i> ↑ Whole-body <i>bdnf</i></p>
Costa de Melo <i>et al.</i> , 2019	15	6	1	Adult	F	<p>Anxiety/fear-related behaviour ↓ Time in the upper zone ↑ Latency to upper zone ↑ Freezing duration</p> <p>Locomotor function ↓ Total distance travelled ↓ Quadrants crossed ↑ Erratic swimming</p>
Huang, Butler & Lubin, 2019	14	6	1	Adult	M+F	<p>Anxiety/fear-related behaviour = Percent at bottom</p> <p>Cortisol ↑ Trunk cortisol (15 min after the last stressor)</p>

						<p>Locomotor function = Total distance travelled</p> <p>Neurochemical outcomes ↑ <i>ache</i> expression (female) ↑ <i>nr3c1</i> expression ↑ <i>hsd11b2</i> expression = <i>npv</i> expression</p>
Marcon <i>et al.</i> , 2019	14	6	1	Adult	M:F	<p>Anxiety/fear-related behaviour ↓ Time in the upper zone = Time in the middle zone ↑ Time in the bottom ↓ Entries in the upper zone</p> <p>Locomotor function = Total distance travelled = Crossings</p> <p>Neurochemical outcomes ↑ TBARS levels ↓ NPSH levels = SH total levels ↓ SOD activity = CAT activity</p>
Mocelin <i>et al.</i> , 2019	14	6	1	Adult	M:F	<p>Anxiety/fear-related behaviour ↓ Time in the upper zone ↓ Entries in the upper zone</p>

						<p>↑ Time in the bottom = Entries in the bottom</p> <p>Locomotor function = Total distance travelled = Crossings</p> <p>Neurochemical outcomes ↑ TBARS levels ↑ Reactive oxygen species (ROS) levels - DCF fluorescence ↓ NPSH levels ↓ SOD activity = CAT activity</p>
Reddy <i>et al.</i> , 2019	7	10	1,4	Adult	Unclear	<p>Anxiety/fear-related behaviour ↓ Time spent in the upper zone ↑ Latency to upper zone ↑ Freezing duration (social behaviour test, before drug treatment)</p> <p>Locomotor function ↓ Crossings</p> <p>Social behaviour ↓ Interaction time ↑ Latency to interaction</p>

Demin <i>et al.</i> , 2020	34	>10	7, 14, 21, 28, 35	Adult	M:F	<p>Anxiety/fear-related behaviour</p> <ul style="list-style-type: none"> ↓ Time spent in the upper zone ↓ Time spent in the light zone (1, 2, 3 weeks of UCS) ↓ Distance to the surface (1 week of UCS) ↑ Distance to the surface (2 weeks of UCS) ↓ Time spent active (1 week of UCS) ↑ Time spent active (3 weeks of UCS) <p>Locomotor function</p> <ul style="list-style-type: none"> ↑ Distance travelled (5 weeks of UCS) <p>Neurochemical outcomes</p> <ul style="list-style-type: none"> = Whole-brain serotonin ↑ 5-HIAA levels (2 weeks of UCS) ↓ 5-HIAA levels (4 weeks of UCS) ↑ 5-HIAA/5HT ratio (2 weeks of UCS) ↓ 5-HIAA/5HT ratio (3,4 weeks of UCS) = Norepinephrine = <i>saga</i> expression ↓ <i>isg15</i> expression
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						<p>↓ <i>otx5</i> expression ↑ <i>tpm4b</i> expression</p> <p>Social behaviour ↓ Interfish distance (5 weeks of UCS)</p>
Golla <i>et al.</i> , 2020	8	5	1, 2, 3, 8	Larval	Unclear	<p>Anxiety/fear-related behaviour = Thigmotaxis index = Scototaxis index ↓ Vertical position (1-3 days post UCS) ↑ Ratio of fish in bottom third (1-3 days post UCS)</p> <p>Locomotor function ↑ Total distance travelled (Light-dark test; 2 days post UCS) ↑ Mean velocity (Light-dark test; 2 days post UCS)</p> <p>Morphometric measurements ↓ Size</p> <p>Social behaviour = Nearest neighbour distance = Interfish distance</p>

O'Daniel & Petrunich-Rutherford, 2020	7	7	1, 8	Adult	M:F	<p>Anxiety/fear-related behaviour ↓ Time spent in the upper zone (1 day post UCS) ↑ Entries in the upper zone (7 days post UCS) ↑ Distance travelled in the upper zone (7 days post UCS) = Freezing duration</p> <p>Cortisol ↓ Trunk cortisol (1 day post UCS)</p> <p>Locomotor function = Total distance travelled = Mean ambulatory velocity</p> <p>Morphometric measurements = Trunk weight</p>
Thomson <i>et al.</i> , 2020	7	3	0	Adult	F	<p>Anxiety/fear-related behaviour ↓ Time spent in the bottom</p> <p>Locomotor function ↓ Velocity = Fractal dimension</p>
Bertelli <i>et al.</i> , 2021	14	6	1	Adult	M:F	<p>Anxiety/fear-related behaviour ↓ Time spent in the upper zone = Entries in the upper zone = Time in the centre zone</p>

						<p>↑ Freezing duration</p> <p>Locomotor function ↓ Total distance travelled (open tank test) = Absolute turn angle = Crossings</p> <p>Morphometric measurements ↓ Weight</p> <p>Neurochemical outcomes ↑ TBARS levels ↓ NPSH levels</p> <p>Peripheral outcomes ↑ Blood glucose</p> <p>Social behaviour = Time in the interaction zone = Interaction time = Number of interactions</p>
Biney <i>et al.</i> , 2021	14	8	4	Adult	Unclear	<p>Anxiety/fear-related behaviour ↓ Time spent in the upper zone ↓ Entries in the upper zone ↓ Time spent in the light zone = Entries in the light zone</p>

						Social behaviour = Shoal cohesion
Chen <i>et al.</i> , 2021	35	Unclear	1	Adult	M:F	Anxiety/fear-related behaviour ↓ Time spent in the upper zone ↓ Time spent in the light zone ↑ Latency to the dark zone Cortisol ↑ Peripheral cortisol Neurochemical outcomes ↑ <i>bdnf</i> expression ↑ <i>tnf-α</i> expression ↑ <i>IL-1β</i> expression ↑ <i>IL-10</i> expression Morphometric measurements ↓ Body mass index
Demin <i>et al.</i> , 2021	77	>10	1	Adult	M:F	Anxiety/fear-related behaviour ↓ Time spent in the upper zone Learning and memory ↓ Time spent in the light zone Locomotor function = Mean velocity Neurochemical outcomes

						<p>↑ Norepinephrine levels = Dopamine levels = Serotonin levels = 5HIAA to 5HT ratio</p> <p>Social behaviour ↓ Interfish distance</p>
Fontana <i>et al.</i> , 2021a	7, 14	8	~ 180	Larval / Adult	Unclear	<p>Anxiety/fear-related behaviour ↓ Time spent in the bottom (7 days of UCS protocol)</p> <p>Cortisol = Whole-body cortisol</p> <p>Learning and memory = Time spent close to the object = Entries to the object zone</p> <p>Locomotor function = Total distance travelled</p>
Fontana <i>et al.</i> , 2021b	3, 7, 14	8	1, 120	Larval / Juvenile, Adult	M:F	<p>Anxiety/fear-related behaviour ↑ Time spent in the upper zone (7 days of UCS protocol/ Adult) ↓ Time spent in the dark zone (7 days of UCS protocol/ Adult) ↑ Thigmotaxis (7 days of UCS protocol/ Juvenile) = Preference index</p>

						<p>Cortisol = Whole-body cortisol</p> <p>Learning and memory = Total turns = Alternations = Repetitions</p> <p>Locomotor function = Total distance travelled</p> <p>Social behaviour = Interfish distance = Shoal average area</p>
Fontana <i>et al.</i> , 2021c	3	3	> 75	Juvenile / Adult	Unclear	<p>Anxiety/fear-related behaviour = Time spent in the bottom</p> <p>Learning and memory ↑ Average of turns ↑ Relative alterations ↓ Relative repetitions = Relative right turns = Relative left turns</p> <p>Locomotor function = Total distance travelled</p>

						Social behaviour = Shoal cohesion
Kirsten <i>et al.</i> , 2021	14	9	0.5	Adult	M:F	Neurochemical outcomes = <i>bdnf</i> expression ↑ <i>tnf-α</i> expression ↑ <i>IL-1β</i> expression = <i>IL-4</i> expression = <i>IL-6</i> expression ↑ <i>IL-10</i> expression ↓ <i>c-FOS</i> expression = <i>INF-γ</i> expression
Reddy <i>et al.</i> , 2021	10	10	1, 2	Adult	M:F	Anxiety/fear-related behaviour ↑ Time spent in the bottom ↓ Transitions to upper zone ↑ No movement duration ↑ Latency to feed ↓ Feeding frequency ↓ Latency to freeze ↑ Freezing bouts ↑ Freezing duration ↓ Time spent in the pheromone zone Locomotor function ↓ Total distance travelled ↓ Mean velocity ↓ Movement duration ↓ Highly mobile duration

						<p>↓ Duration of erratic movements</p> <p>Neurochemical outcomes</p> <p>↓ <i>bdnf</i> expression</p> <p>↑ <i>crf</i> expression</p> <p>↑ <i>calcineurin</i> expression</p> <p>↓ <i>B-III tubulin</i> expression</p> <p>= <i>blbp</i> expression</p> <p>↓ <i>pmTOR/mTOR</i> ratio</p> <p>↓ <i>sox2</i> expression</p> <p>↓ <i>sox2</i> positive cells</p> <p>Proliferative index</p> <p>↑ Proliferative index telencephalon (Dm)</p> <p>↓ Proliferative index telencephalon (DId + DIv)</p> <p>Social behaviour</p> <p>↓ Duration of interaction (with target fish in the interaction zone)</p> <p>↓ Interaction frequency (with target fish in the interaction zone)</p>
Rosdy <i>et al.</i> , 2021	14	10	Unclear	Adult	Unclear	<p>Anxiety/fear-related behaviour</p> <p>↓ Time spent in the upper zone</p> <p>↓ Time spent in the light zone</p>
Shams, Khan & Gerlai, 2021	15	6	1	Adult	M:F	<p>Cortisol</p> <p>↑ Whole-body cortisol</p>

Zhang <i>et al.</i> , 2021	28	8	1	Adult	Unclear	<p>Anxiety/fear-related behaviour</p> <ul style="list-style-type: none"> ↓ Time spent in the upper zone ↓ Latency to the upper zone ↓ Freezing bouts ↓ Freezing duration ↑ Immobility time <p>Locomotor function</p> <ul style="list-style-type: none"> ↓ Total distance travelled ↓ Mean velocity ↑ Meandering ↑ Absolut turn angle ↑ Angular velocity
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1 IV. DISCUSSION

2 Ten years after the publication of the first study of UCS conducted using zebrafish as
3 the model animal (Piato *et al.*, 2011), we performed a systematic review and meta-
4 analysis of the literature to evaluate and synthesize the behavioural and neurochemical
5 effects of this protocol. Despite the relatively low number of studies carried out with far
6 fewer animals than the rodent literature, the main findings of our study show that UCS
7 increases anxiety-like behaviour and cortisol levels while decreasing locomotor activity
8 in zebrafish. On the other hand, no effects on social behaviour and other biomarkers
9 (*bdnf* and *crf*) were observed in this species.

10 Such results somewhat correlate with the findings gathered from experiments
11 conducted with rodents. As mentioned before, although the stress regimen is shown
12 to consistently induce anhedonic behaviour in rodents, several variables intrinsic to
13 the organisms such as species, sex, age, and resilience or the protocol itself have a
14 great impact on the outcomes measured, leading to the heterogeneity seen in the
15 literature (Antoniuk *et al.*, 2019). Results for anxiety-like behaviour (Kompagne *et al.*,
16 2008; Cox *et al.*, 2011; Zhu *et al.*, 2014), locomotor function (Kumar, Kuhad & Chopra,
17 2011; Sequeira-Cordero *et al.*, 2019), and social behaviour (Boxelaere *et al.*, 2017)
18 vary considerably depending on the conditions applied in the experiments and are still
19 in need of a thorough systematic review to determine effect direction. The same can
20 be said for the hormonal regulation of the stress response and related neurochemical
21 outcomes. It is also expected to observe an increase in corticosterone and an
22 imbalance of neurochemical markers driven by the UCS in rodents, but many reports
23 reveal behavioural alterations in the absence of detectable modifications in these other
24 parameters as reviewed elsewhere (Willner, 2017a; Lages *et al.*, 2021).

1 Many factors might explain the high heterogeneity revealed between included
2 studies and the behavioural response of fish. The number and classes of stressors
3 used to generate stress differ substantially between studies. This information is crucial
4 since different stressors have been shown to trigger different patterns of behavioural
5 and biochemical responses in rodents (Antoniuk *et al.*, 2019). The majority of
6 experiments have been conducted using mixed samples of both male and female
7 zebrafish without reporting individualised effects of UCS by sex. Unfortunately, it is still
8 difficult to evaluate these differential impacts since more studies are required to
9 conduct analyses grouped by sex; however, a few experiments have already shown
10 that stress can elicit different responses in male and female zebrafish (Rambo *et al.*,
11 2017; Huang, Butler & Lubin, 2019).

12 Subgroup analyses indicate that the duration of the stress protocol might also
13 influence the outcome of the UCS protocol, corroborating what was shown within
14 previous works (Piato *et al.*, 2011; Palucha-Poniewiera *et al.*, 2020; Fontana *et al.*,
15 2021a). When grouping experiments by this variable, no significant effects of the
16 stress are observed in anxiety/fear-related behaviour, locomotor function, and cortisol
17 levels for stress regimens of up to 7 days despite the overall effects of UCS for these
18 domains. Protocols with more than 7 days, on the other hand, show a significant effect
19 of UCS for the same variables, indicating that regimens of more than a week of stress
20 are necessary to reveal the deleterious consequences of stress in zebrafish. It is
21 important to note that most experiments designed to evaluate the long-lasting effects
22 of UCS in zebrafish were included in the group with shorter stress times. In these
23 cases, stress sessions occur in early developmental stages and tests usually take
24 place later in the animal life. This allows for a long washout period between the stress
25 and outcome assessment that might explain the lack of effects of stress when such

1 designs are used. Capturing UCS effects heavily depends on assessment timing
2 (Willner, 2017a; Bosch *et al.*, 2022), and tests should be scheduled to avoid observing
3 acute effects of a single stressor as well as losing the effects of the intervention as a
4 whole since animals are likely to eventually recover, unless the stressors coincide with
5 a window of developmental vulnerability (Jankord *et al.*, 2011).

6 The results of this review should be interpreted with caution considering that
7 the main effects of the analyses were influenced by studies with a high risk of bias.
8 Although many efforts have been made to improve the reporting quality of pre-clinical
9 research (Sert *et al.*, 2020), the publication of studies adhering to measures designed
10 to mitigate the risk of bias associated with methodological conduct is still low (Baker
11 *et al.*, 2014; Macleod *et al.*, 2015). These problems hamper the correct analysis of
12 results and contribute to the reproducibility crisis in the biomedical field (Samsa &
13 Samsa, 2019; Gerlai, 2019), encouraging researchers to question the validity of animal
14 models (Worp *et al.*, 2010). By excluding studies with a high risk of bias in the
15 sensitivity analysis it was possible to visualise the direct impacts of these on distorting
16 the main effects found in the meta-analyses for anxiety/fear-related behaviour,
17 locomotor activity, and especially for cortisol, for which effect direction was inverted in
18 sensitivity analysis. Conclusions should also be conservative for *bdnf* and *crf* since, as
19 mentioned before, all of the studies included presented a high risk of bias, revealing
20 the alarming need to improve internal validity and reporting quality.

21 In the same way, publication bias plays a part in generating misleading
22 assumptions even in meta-analyses based on broad and rigorous systematic reviews
23 (Worp *et al.*, 2010). There is evidence of selective publishing of studies for the domains
24 tested based on funnel plot inspection and Egger's test evaluation, pointing to the need

1 to conduct well-delineated experiments using this model, as these results denote a
2 possible overestimation of the effects of chronic stress in zebrafish.

3

4 **V. CONCLUSIONS**

5 (1) The overall results of our meta-analysis reveal the effects of UCS in increasing
6 anxiety/fear-related behaviour and cortisol levels in stressed animals while
7 decreasing locomotor function.

8 (2) No effects of stress were found on social behaviour and the expression of *bdnf*
9 and *crf*, but the literature reporting these outcomes is limited and with evidence
10 of bias.

11 (3) The risk of bias was considered generally high for the studies included in this
12 review, indicating poor methodological and reporting quality of studies
13 conducted using zebrafish.

14 (4) We found moderate to high heterogeneity in the data, suggesting that several
15 variables could influence the results obtained. Given the small number of
16 studies included, it is difficult to point out the sources of variation other than the
17 duration of the stress protocol.

18 (5) Protocols of more than a week of stress seem to be better suited to induce
19 behavioural and biochemical alterations that are expected to occur with UCS.

20 (6) The analyses conducted stress the need to conduct well-designed experiments
21 using the UCS model to assess its effects on zebrafish behaviour and
22 neurochemical parameters, further exploring the sources of variation that might
23 influence the results, such as the nature of stressors and sex.

24 (7) Overall, this review corroborates the need for improvement in methodological
25 and reporting conduct across preclinical research.

1

2 VI. AUTHOR CONTRIBUTIONS

3 **Matheus Gallas-Lopes:** conceptualization, data curation, formal analysis,
4 investigation, methodology, project administration, visualisation, and writing - original
5 draft; **Leonardo M. Bastos:** conceptualization, investigation, methodology, and
6 writing – review & editing; **Radharani Benvenuti:** conceptualization, investigation,
7 methodology, and writing – review & editing; **Alana C. Panzenhagen:**
8 conceptualization, formal analysis, methodology, visualisation, and writing - review &
9 editing; **Angelo Piato:** conceptualization, investigation, methodology, and writing –
10 review & editing; **Ana P. Herrmann:** conceptualization, data curation, formal analysis,
11 investigation, methodology, project administration, supervision, visualisation, and
12 writing – review & editing;

13

14 VII. CONFLICT OF INTEREST

15 The authors declare no conflicts of interest.

16

17 VIII. DATA AVAILABILITY

18 All data is available in Open Science Framework (<https://osf.io/j2zva/>).

19

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X. REFERENCES

Antoniuk, S., Bijata, M., Ponimaskin, E. & Wlodarczyk, J. (2019) Chronic unpredictable mild stress for modeling depression in rodents: Meta-analysis of model reliability. *Neuroscience & Biobehavioral Reviews* **99**, 101–116.

Baker, D., Lidster, K., Sottomayor, A. & Amor, S. (2014) Two Years Later: Journals Are Not Yet Enforcing the ARRIVE Guidelines on Reporting Standards for Pre-Clinical Animal Studies. *PLOS Biology* **12**, e1001756. Public Library of Science.

Benneh, C.K., Biney, R.P., Mante, P.K., Tandoh, A., Adongo, D.W. & Woode, E. (2017) *Maerua angolensis* stem bark extract reverses anxiety and related behaviours in zebrafish—Involvement of GABAergic and 5-HT systems. *Journal of Ethnopharmacology* **207**, 129–145.

Bertelli, P.R., Mocelin, R., Marcon, M., Sachett, A., Gomez, R., Rosa, A.R., Herrmann, A.P. & Piato, A. (2021) Anti-stress effects of the glucagon-like peptide-1 receptor agonist liraglutide in zebrafish. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* **111**, 110388.

Biney, R.P., Benneh, C.K., Adongo, D.W., Ameyaw, E.O. & Woode, E. (2021) Evidence of an antidepressant-like effect of xylopic acid mediated by serotonergic mechanisms. *Psychopharmacology* **238**, 2105–2120.

Bosch, K., Sbrini, G., Burattini, I., Nieuwenhuis, D., Calabrese, F., Schubert, D., Henckens, M.J.A.G. & Homberg, J.R. (2022) Repeated testing modulates chronic unpredictable mild stress effects in male rats. *Behavioural Brain Research* **432**, 113960.

Boxelaere, M. van, Clements, J., Callaerts, P., D’Hooge, R. & Callaerts-Vegh, Z. (2017) Unpredictable chronic mild stress differentially impairs social and

1 contextual discrimination learning in two inbred mouse strains. *PLOS ONE* **12**,
2 e0188537. Public Library of Science.

3 Chakravarty, S., Reddy, B.R., Sudhakar, S.R., Saxena, S., Das, T., Meghah, V.,
4 Swamy, C.V.B., Kumar, A. & Idris, M.M. (2013) Chronic Unpredictable Stress
5 (CUS)-Induced Anxiety and Related Mood Disorders in a Zebrafish Model:
6 Altered Brain Proteome Profile Implicates Mitochondrial Dysfunction. *PLOS*
7 *ONE* **8**, e63302. Public Library of Science.

8 Chen, B., Peng, Z., Zhang, C., Lin, H., Gao, J., Zheng, H., Cao, W. & Qin, X. (2021)
9 Study on Improving Effect of Oyster Hydrolysate on Depressive Behavior of
10 Zebrafish Under Chronic Unpredictable Mild Stress. *Shipin kexue jishu xuebao*
11 **39**, 55–63. Beijing Technology and Business University, Department of Science
12 and Technology.

13 Costa de Melo, N., Sánchez-Ortiz, B.L., dos Santos Sampaio, T.I., Matias Pereira,
14 A.C., Pinheiro da Silva Neto, F.L., Ribeiro da Silva, H., Alves Soares Cruz, R.,
15 Keita, H., Soares Pereira, A.M. & Tavares Carvalho, J.C. (2019) Anxiolytic and
16 Antidepressant Effects of the Hydroethanolic Extract from the Leaves of *Aloysia*
17 *polystachya* (Griseb.) Moldenke: A Study on Zebrafish (*Danio rerio*).
18 *Pharmaceuticals* **12**, 106. Multidisciplinary Digital Publishing Institute.

19 Cox, B.M., Alsawah, F., McNeill, P.C., Galloway, M.P. & Perrine, S.A. (2011)
20 Neurochemical, hormonal, and behavioral effects of chronic unpredictable
21 stress in the rat. *Behavioural Brain Research* **220**, 106–111.

22 Davis, D.J., Doerr, H.M., Grzelak, A.K., Busi, S.B., Jasarevic, E., Ericsson, A.C. &
23 Bryda, E.C. (2016) *Lactobacillus plantarum* attenuates anxiety-related behavior
24 and protects against stress-induced dysbiosis in adult zebrafish. *Scientific*
25 *Reports* **6**, 33726. Nature Publishing Group.

- 1 Demin, K.A., Kolesnikova, T.O., Galstyan, D.S., Krotova, N.A., Ilyin, N.P., Derzhavina,
2 K.A., Levchenko, N.A., Strekalova, T., de Abreu, M.S., Petersen, E.V.,
3 Seredinskaya, M., Cherneyko, Y.V., Kositsyn, Y.M., Sorokin, D.V., Zabegalov,
4 K.N., et al. (2021) Modulation of behavioral and neurochemical responses of
5 adult zebrafish by fluoxetine, eicosapentaenoic acid and lipopolysaccharide in
6 the prolonged chronic unpredictable stress model. *Scientific Reports* **11**, 14289.
7 Nature Publishing Group.
- 8 Demin, K.A., Lakstygal, A.M., Krotova, N.A., Masharsky, A., Tagawa, N., Chernysh,
9 M.V., Ilyin, N.P., Taranov, A.S., Galstyan, D.S., Derzhavina, K.A., Levchenko,
10 N.A., Kolesnikova, T.O., Mor, M.S., Vasyutina, M.L., Efimova, E.V., et al. (2020)
11 Understanding complex dynamics of behavioral, neurochemical and
12 transcriptomic changes induced by prolonged chronic unpredictable stress in
13 zebrafish. *Scientific Reports* **10**, 19981. Nature Publishing Group.
- 14 Egger, M., Smith, G.D., Schneider, M. & Minder, C. (1997) Bias in meta-analysis
15 detected by a simple, graphical test. *BMJ* **315**, 629–634. British Medical Journal
16 Publishing Group.
- 17 Fontana, B.D., Cleal, M., Norton, W.H.J. & Parker, M.O. (2021a) The impact of chronic
18 unpredictable early-life stress (CUELS) on boldness and stress-reactivity:
19 Differential effects of stress duration and context of testing. *Physiology &*
20 *Behavior* **240**, 113526.
- 21 Fontana, B.D., Gibbon, A.J., Cleal, M., Norton, W.H.J. & Parker, M.O. (2021b) Chronic
22 unpredictable early-life stress (CUELS) protocol: Early-life stress changes
23 anxiety levels of adult zebrafish. *Progress in Neuro-Psychopharmacology and*
24 *Biological Psychiatry* **108**, 110087.
- 25 Fontana, B.D., Gibbon, A.J., Cleal, M., Sudwarts, A., Pritchett, D., Miletto Petrazzini,

- 1 M.E., Brennan, C.H. & Parker, M.O. (2021c) Moderate early life stress improves
2 adult zebrafish (*Danio rerio*) working memory but does not affect social and
3 anxiety-like responses. *Developmental Psychobiology* **63**, 54–64.
- 4 Fulcher, N., Tran, S., Shams, S., Chatterjee, D. & Gerlai, R. (2017) Neurochemical
5 and Behavioral Responses to Unpredictable Chronic Mild Stress Following
6 Developmental Isolation: The Zebrafish as a Model for Major Depression.
7 *Zebrafish* **14**, 23–34. Mary Ann Liebert, Inc., publishers.
- 8 Gallas-Lopes, M., Herrmann, A.P., Benvenuti, R., Piato, A., Panzenhagen, A.C. &
9 Bastos, L.M. (2021) Unpredictable chronic stress in zebrafish: a systematic
10 review. OSF.
- 11 Gerlai, R. (2019) Reproducibility and replicability in zebrafish behavioral neuroscience
12 research. *Pharmacology Biochemistry and Behavior* **178**, 30–38.
- 13 Golla, A., Østby, H. & Kermen, F. (2020) Chronic unpredictable stress induces anxiety-
14 like behaviors in young zebrafish. *Scientific Reports* **10**, 10339. Nature
15 Publishing Group.
- 16 Grzelak, A.K., Davis, D.J., Caraker, S.M., Crim, M.J., Spitsbergen, J.M. & Wiedmeyer,
17 C.E. (2017) Stress Leukogram Induced by Acute and Chronic Stress in
18 Zebrafish (*Danio rerio*). *Comparative Medicine* **67**, 263–269.
- 19 Higgins, J.P.T. & Thompson, S.G. (2002) Quantifying heterogeneity in a meta-
20 analysis. *Statistics in Medicine* **21**, 1539–1558.
- 21 Hooijmans, C.R., Rovers, M.M., de Vries, R.B., Leenaars, M., Ritskes-Hoitinga, M. &
22 Langendam, M.W. (2014) SYRCLE's risk of bias tool for animal studies. *BMC*
23 *Medical Research Methodology* **14**, 43.
- 24 Huang, V., Butler, A.A. & Lubin, F.D. (2019) Telencephalon transcriptome analysis of
25 chronically stressed adult zebrafish. *Scientific Reports* **9**, 1379. Nature

- 1 Publishing Group.
- 2 Jankord, R., Solomon, M.B., Albertz, J., Flak, J.N., Zhang, R. & Herman, J.P. (2011)
- 3 Stress Vulnerability during Adolescent Development in Rats. *Endocrinology*
- 4 **152**, 629–638.
- 5 Jayamurali, D. & Govindarajulu, S.N. (2017) Impact of chronic unpredictable stress on
- 6 the expression of apoptotic genes in zebrafish brain. *International Journal of*
- 7 *Pharmaceutical Sciences and Research* **8**, 4363–4370.
- 8 Katz, R.J. (1982) Animal model of depression: pharmacological sensitivity of a hedonic
- 9 deficit. *Pharmacology, Biochemistry, and Behavior* **16**, 965–968.
- 10 Katz, R.J. & Hersh, S. (1981) Amitriptyline and scopolamine in an animal model of
- 11 depression. *Neuroscience and Biobehavioral Reviews* **5**, 265–271.
- 12 Katz, R.J., Roth, K.A. & Carroll, B.J. (1981) Acute and chronic stress effects on open
- 13 field activity in the rat: implications for a model of depression. *Neuroscience and*
- 14 *Biobehavioral Reviews* **5**, 247–251.
- 15 Kirsten, K., Pompermaier, A., Koakoski, G., Mendonça-Soares, S., da Costa, R.A.,
- 16 Maffi, V.C., Kreutz, L.C. & Barcellos, L.J.G. (2021) Acute and chronic stress
- 17 differently alter the expression of cytokine and neuronal markers genes in
- 18 zebrafish brain. *Stress* **24**, 107–112. Taylor & Francis.
- 19 Kompagne, H., Bárdos, G., Szénási, G., Gacsályi, I., Hársing, L.G. & Lévy, G. (2008)
- 20 Chronic mild stress generates clear depressive but ambiguous anxiety-like
- 21 behaviour in rats. *Behavioural Brain Research* **193**, 311–314.
- 22 Kumar, B., Kuhad, A. & Chopra, K. (2011) Neuropsychopharmacological effect of
- 23 sesamol in unpredictable chronic mild stress model of depression: behavioral
- 24 and biochemical evidences. *Psychopharmacology* **214**, 819–828.
- 25 Lages, Y.V.M., Rossi, A.D., Krahe, T.E. & Landeira-Fernandez, J. (2021) Effect of

1 chronic unpredictable mild stress on the expression profile of serotonin
2 receptors in rats and mice: a meta-analysis. *Neuroscience & Biobehavioral*
3 *Reviews* **124**, 78–88.

4 Landis, S.C., Amara, S.G., Asadullah, K., Austin, C.P., Blumenstein, R., Bradley, E.W.,
5 Crystal, R.G., Darnell, R.B., Ferrante, R.J., Fillit, H., Finkelstein, R., Fisher, M.,
6 Gendelman, H.E., Golub, R.M., Goudreau, J.L., et al. (2012) A call for
7 transparent reporting to optimize the predictive value of preclinical research.
8 *Nature* **490**, 187–191. Nature Publishing Group.

9 Macleod, M.R., McLean, A.L., Kyriakopoulou, A., Serghiou, S., Wilde, A. de, Sherratt,
10 N., Hirst, T., Hemblade, R., Bahor, Z., Nunes-Fonseca, C., Potluru, A.,
11 Thomson, A., Baginskitaie, J., Egan, K., Vesterinen, H., et al. (2015) Risk of
12 Bias in Reports of In Vivo Research: A Focus for Improvement. *PLOS Biology*
13 **13**, e1002273. Public Library of Science.

14 Manuel, R., Gorissen, M., Zethof, J., Ebbesson, L.O.E., van de Vis, H., Flik, G. & van
15 den Bos, R. (2014) Unpredictable chronic stress decreases inhibitory
16 avoidance learning in Tuebingen long-fin zebrafish: stronger effects in the
17 resting phase than in the active phase. *Journal of Experimental Biology* **217**,
18 3919–3928.

19 Marcon, M., Herrmann, A.P., Mocelin, R., Rambo, C.L., Koakoski, G., Abreu, M.S.,
20 Conterato, G.M.M., Kist, L.W., Bogo, M.R., Zanatta, L., Barcellos, L.J.G. &
21 Piato, A.L. (2016) Prevention of unpredictable chronic stress-related
22 phenomena in zebrafish exposed to bromazepam, fluoxetine and nortriptyline.
23 *Psychopharmacology* **233**, 3815–3824.

24 Marcon, M., Mocelin, R., Benvenuti, R., Costa, T., Herrmann, A.P., de Oliveira, D.L.,
25 Koakoski, G., Barcellos, L.J.G. & Piato, A. (2018a) Environmental enrichment

1 modulates the response to chronic stress in zebrafish. *Journal of Experimental*
2 *Biology* **221**, jeb176735.

3 Marcon, M., Mocelin, R., de Oliveira, D.L., da Rosa Araujo, A.S., Herrmann, A.P. &
4 Piato, A. (2019) Acetyl-L-carnitine as a putative candidate for the treatment of
5 stress-related psychiatric disorders: Novel evidence from a zebrafish model.
6 *Neuropharmacology* **150**, 145–152.

7 Marcon, M., Mocelin, R., Sachett, A., Siebel, A.M., Herrmann, A.P. & Piato, A. (2018b)
8 Enriched environment prevents oxidative stress in zebrafish submitted to
9 unpredictable chronic stress. *PeerJ* **6**, e5136. PeerJ Inc.

10 Maximino, C., Silva, R., da Silva, S. de N., Rodrigues, L. do S., Barbosa, H., de
11 Carvalho, T., Leão, L.K., Lima, M., Oliveira, K.R. & Herculano, A. (2015) Non-
12 mammalian models in behavioral neuroscience: consequences for biological
13 psychiatry. *Frontiers in Behavioral Neuroscience* **9**.

14 McGuinness, L.A. & Higgins, J.P.T. (2021) Risk-of-bias VISualization (robvis): An R
15 package and Shiny web app for visualizing risk-of-bias assessments. *Research*
16 *Synthesis Methods* **12**, 55–61.

17 Mocelin, R., Marcon, M., D’ambros, S., Mattos, J., Sachett, A., Siebel, A.M.,
18 Herrmann, A.P. & Piato, A. (2019) N-Acetylcysteine Reverses Anxiety and
19 Oxidative Damage Induced by Unpredictable Chronic Stress in Zebrafish.
20 *Molecular Neurobiology* **56**, 1188–1195.

21 Nollet, M. (2021) Models of Depression: Unpredictable Chronic Mild Stress in Mice.
22 *Current Protocols* **1**, e208.

23 O’Daniel, M.P. & Petrunich-Rutherford, M.L. (2020) Effects of chronic prazosin, an
24 alpha-1 adrenergic antagonist, on anxiety-like behavior and cortisol levels in a
25 chronic unpredictable stress model in zebrafish (*Danio rerio*). *PeerJ* **8**, e8472.

1 PeerJ Inc.

2 Ouzzani, M., Hammady, H., Fedorowicz, Z. & Elmagarmid, A. (2016) Rayyan—a web
3 and mobile app for systematic reviews. *Systematic Reviews* **5**, 210.

4 Page, M.J., McKenzie, J.E., Bossuyt, P.M., Boutron, I., Hoffmann, T.C., Mulrow, C.D.,
5 Shamseer, L., Tetzlaff, J.M., Akl, E.A., Brennan, S.E., Chou, R., Glanville, J.,
6 Grimshaw, J.M., Hróbjartsson, A., Lalu, M.M., et al. (2021) The PRISMA 2020
7 statement: an updated guideline for reporting systematic reviews. *BMJ* **372**,
8 n71. British Medical Journal Publishing Group.

9 Palucha-Poniewiera, A., Podkowa, K., Rafalo-Ulinska, A., Branski, P. & Burnat, G.
10 (2020) The influence of the duration of chronic unpredictable mild stress on the
11 behavioural responses of C57BL/6J mice. *Behavioural Pharmacology* **31**, 574–
12 582.

13 Pavlidis, M., Theodoridi, A. & Tsalafouta, A. (2015) Neuroendocrine regulation of the
14 stress response in adult zebrafish, *Danio rerio*. *Progress in Neuro-*
15 *Psychopharmacology and Biological Psychiatry* **60**, 121–131.

16 Piato, Â.L., Capiotti, K.M., Tamborski, A.R., Oses, J.P., Barcellos, L.J.G., Bogo, M.R.,
17 Lara, D.R., Vianna, M.R. & Bonan, C.D. (2011) Unpredictable chronic stress
18 model in zebrafish (*Danio rerio*): Behavioral and physiological responses.
19 *Progress in Neuro-Psychopharmacology and Biological Psychiatry* **35**, 561–
20 567.

21 Rambo, C.L., Mocelin, R., Marcon, M., Villanova, D., Koakoski, G., de Abreu, M.S.,
22 Oliveira, T.A., Barcellos, L.J.G., Piato, A.L. & Bonan, C.D. (2017) Gender
23 differences in aggression and cortisol levels in zebrafish subjected to
24 unpredictable chronic stress. *Physiology & Behavior* **171**, 50–54.

25 Reddy, B.R., Babu, N.S., Das, T., Bhattacharya, D., Murthy, Ch.L.N., Kumar, A., Idris,

1 M.M. & Chakravarty, S. (2021) Proteome profile of telencephalon associates
2 attenuated neurogenesis with chronic stress induced mood disorder
3 phenotypes in zebrafish model. *Pharmacology Biochemistry and Behavior* **204**,
4 173170.

5 Reddy, R.G., Dachavaram, S.S., Reddy, B.R., Kalyankar, K.B., Rajan, W.D., Kootar,
6 S., Kumar, A., Das, S. & Chakravarty, S. (2018) Fellutamide B Synthetic Path
7 Intermediates with in Vitro Neuroactive Function Shows Mood-Elevating Effect
8 in Stress-Induced Zebrafish Model. *ACS Omega* **3**, 10534–10544. American
9 Chemical Society.

10 Reddy, R.G., Surineni, G., Bhattacharya, D., Marvadi, S.K., Sagar, A., Kalle, A.M.,
11 Kumar, A., Kantevari, S. & Chakravarty, S. (2019) Crafting Carbazole-Based
12 Vorinostat and Tubastatin-A-like Histone Deacetylase (HDAC) Inhibitors with
13 Potent in Vitro and in Vivo Neuroactive Functions. *ACS Omega* **4**, 17279–
14 17294. American Chemical Society.

15 Rosdy, M.S., Rofiee, M.S., Samsulrizal, N., Salleh, M.Z. & Teh, L.K. (2021)
16 Understanding the effects of *Moringa oleifera* in chronic unpredictable stressed
17 zebrafish using metabolomics analysis. *Journal of Ethnopharmacology* **278**,
18 114290.

19 Samsa, G. & Samsa, L. (2019) A Guide to Reproducibility in Preclinical Research.
20 *Academic Medicine* **94**, 47–52.

21 dos Santos Sampaio, T.I., de Melo, N.C., de Freitas Paiva, B.T., da Silva Aleluia, G.A.,
22 da Silva Neto, F.L.P., da Silva, H.R., Keita, H., Cruz, R.A.S., Sánchez-Ortiz,
23 B.L., Pineda-Peña, E.A., Balderas, J.L., Navarrete, A. & Carvalho, J.C.T. (2018)
24 Leaves of *Spondias mombin* L. a traditional anxiolytic and antidepressant:
25 Pharmacological evaluation on zebrafish (*Danio rerio*). *Journal of*

1 *Ethnopharmacology* **224**, 563–578.

2 Sequeira-Cordero, A., Salas-Bastos, A., Fornaguera, J. & Brenes, J.C. (2019)

3 Behavioural characterisation of chronic unpredictable stress based on

4 ethologically relevant paradigms in rats. *Scientific Reports* **9**, 17403. Nature

5 Publishing Group.

6 Sert, N.P. du, Hurst, V., Ahluwalia, A., Alam, S., Avey, M.T., Baker, M., Browne, W.J.,

7 Clark, A., Cuthill, I.C., Dirnagl, U., Emerson, M., Garner, P., Holgate, S.T.,

8 Howells, D.W., Karp, N.A., et al. (2020) The ARRIVE guidelines 2.0: Updated

9 guidelines for reporting animal research. *PLOS Biology* **18**, e3000410. Public

10 Library of Science.

11 Shams, S., Khan, A. & Gerlai, R. (2021) Early social deprivation does not affect cortisol

12 response to acute and chronic stress in zebrafish. *Stress* **24**, 273–281. Taylor

13 & Francis.

14 Song, C., Liu, B.-P., Zhang, Y.-P., Peng, Z., Wang, J., Collier, A.D., Echevarria, D.J.,

15 Savelieva, K.V., Lawrence, R.F., Rex, C.S., Meshalkina, D.A. & Kalueff, A.V.

16 (2018) Modeling consequences of prolonged strong unpredictable stress in

17 zebrafish: Complex effects on behavior and physiology. *Progress in Neuro-*

18 *Psychopharmacology and Biological Psychiatry* **81**, 384–394.

19 Strekalova, T. & Steinbusch, H. (2009) Factors of Reproducibility of Anhedonia

20 Induction in a Chronic Stress Depression Model in Mice. In *Mood and Anxiety*

21 *Related Phenotypes in Mice: Characterization Using Behavioral Tests* (ed T.D.

22 Gould), pp. 153–176. Humana Press, Totowa, NJ.

23 Thomson, J.S., Deakin, A.G., Cossins, A.R., Spencer, J.W., Young, I.S. & Sneddon,

24 L.U. (2020) Acute and chronic stress prevents responses to pain in zebrafish:

25 evidence for stress-induced analgesia. *Journal of Experimental Biology* **223**,

1 jeb224527.

2 Vesterinen, H.M., Sena, E.S., Egan, K.J., Hirst, T.C., Churolov, L., Currie, G.L.,
3 Antonic, A., Howells, D.W. & Macleod, M.R. (2014) Meta-analysis of data from
4 animal studies: A practical guide. *Journal of Neuroscience Methods* **221**, 92–
5 102.

6 Viechtbauer, W. (2010) Conducting Meta-Analyses in R with the metafor Package.
7 *Journal of Statistical Software* **36**, 1–48.

8 Weber-Stadlbauer, U. & Meyer, U. (2019) Challenges and opportunities of a-priori and
9 a-posteriori variability in maternal immune activation models. *Current Opinion*
10 *in Behavioral Sciences* **28**, 119–128.

11 Wilkinson, L. (2011) ggplot2: Elegant Graphics for Data Analysis by WICKHAM, H.
12 *Biometrics* **67**, 678–679.

13 Willner, P. (1997) Validity, reliability and utility of the chronic mild stress model of
14 depression: a 10-year review and evaluation. *Psychopharmacology* **134**, 319–
15 329.

16 Willner, P. (2017a) The chronic mild stress (CMS) model of depression: History,
17 evaluation and usage. *Neurobiology of Stress* **6**, 78–93.

18 Willner, P. (2017b) Reliability of the chronic mild stress model of depression: A user
19 survey. *Neurobiology of Stress* **6**, 68–77.

20 Willner, P., Towell, A., Sampson, D., Sophokleous, S. & Muscat, R. (1987) Reduction
21 of sucrose preference by chronic unpredictable mild stress, and its restoration
22 by a tricyclic antidepressant. *Psychopharmacology* **93**, 358–364.

23 Worp, H.B. van der, Howells, D.W., Sena, E.S., Porritt, M.J., Rewell, S., O'Collins, V.
24 & Macleod, M.R. (2010) Can Animal Models of Disease Reliably Inform Human
25 Studies? *PLOS Medicine* **7**, e1000245. Public Library of Science.

- 1 Zhang, R., Qiao, C., Liu, Q., He, J., Lai, Y., Shang, J. & Zhong, H. (2021) A Reliable
2 High-Throughput Screening Model for Antidepressant. *International Journal of*
3 *Molecular Sciences* **22**, 9505. Multidisciplinary Digital Publishing Institute.
- 4 Zhu, S., Shi, R., Wang, J., Wang, J.-F. & Li, X.-M. (2014) Unpredictable chronic mild
5 stress not chronic restraint stress induces depressive behaviours in mice.
6 *NeuroReport* **25**, 1151–1155.
- 7 Zimmermann, F.F., Altenhofen, S., Kist, L.W., Leite, C.E., Bogo, M.R., Cognato, G.P.
8 & Bonan, C.D. (2016) Unpredictable Chronic Stress Alters Adenosine
9 Metabolism in Zebrafish Brain. *Molecular Neurobiology* **53**, 2518–2528.

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