Fast surveillance response reveals the introduction of a new yellow fever virus sub-lineage in 2021, in Minas Gerais, Brazil

Miguel Souza Andrade¹, Fabrício Souza Campos²,³/+, Cirilo Henrique de Oliveira⁴, Ramon Silva Oliveira⁴, Aline Alves Scarpellini Campos⁵, Marco Antônio Barreto de Almeida⁵, Vagner de Souza Fonseca⁶,7, Danilo Simonini-Teixeira⁶, Anaiá da Paixão Sevá⁶, Andrea Oliveira Dias Temponi⁶, Fernando Maria Magalhães⁶, Danielle Costa Capistrano Chaves⁶, Maira Alves Pereira¹⁰, Ludmila Oliveira Lamounier¹⁰, Givaldo Gomes de Menezes⁶, Sandy Micaele Aquino-Teixeira⁴, Maria Eduarda Gonçalves-dos-Santos⁴, Sofía Bernal-Valle⁶, Nicolas Felipe Drumm Müller⁶, Jader da Cruz Cardoso⁶, Edmilson dos Santos⁶, Maria Angélica Mares-Guia¹¹, George Rêgo Albuquerque⁶, Alessandro Pecego Martins Romano¹², Ana Cláudia Franco⁶, Bergmann Morais Ribeiro¹, Paulo Michel Roeheȝ, Filipe Vieira Santos de Abreu⁴/⁺

¹Universidade de Brasília, Instituto de Ciências Biológicas, Departamento de Biologia Celular, Laboratório de Baculovírus, Brasília, DF, Brasil

BACKGROUND In Brazil, the yellow fever virus (YFV) is maintained in a sylvatic cycle involving wild mosquitoes and non-human primates (NHPs). The virus is endemic to the Amazon region; however, waves of epidemic expansion reaching other Brazilian states sporadically occur, eventually causing spillovers to humans.

OBJECTIVES To report a surveillance effort that led to the first confirmation of YFV in NHPs in the state of Minas Gerais (MG), Southeast region, in 2021.

METHODS A surveillance network was created, encompassing the technology of smartphone applications and coordinated actions of several research institutions and health services to monitor and investigate NHP epizootics.

FINDINGS When alerts were spread through the network, samples from NHPs were collected and YFV infection confirmed by reverse transcription-quantitative polymerase chain reaction (RT-qPCR) and genome sequencing at an interval of only 10 days. Near-complete genomes were generated using the Nanopore MinION sequencer. Phylogenetic analysis indicated that viral genomes were related to the South American genotype I, clustering with a genome detected in the Amazon region (state of Pará) in 2017, named YFV_{PA/MG} sub-lineage. Fast YFV confirmation potentialised vaccination campaigns.

MAIN CONCLUSIONS A new YFV introduction was detected in MG 6 years after the beginning of the major outbreak reported in the state (2015-2018). The YFV strain was not related to the sub-lineages previously reported in MG. No human cases have been reported, suggesting the importance of coordinated surveillance of NHPs using available technologies and supporting laboratories to ensure a quick response and implementation of contingency measures to avoid YFV spillover to humans.

Key words: yellow fever virus – Arbovirus – Flavivirus – non-human primate – epizootic – smartphone – MinION

doi: 10.1590/0074-02760220127

Financial support: CNPq/Decit/SCTIE/MS (grant 443215/2019-7).

Received 05 June 2022 Accepted 10 October 2022 In Brazil, the yellow fever virus (YFV) (family *Flaviviridae*, genus *Flavivirus*) exhibited two epidemiologically distinct transmission cycles: urban and sylvatic. In the urban cycle, which has not been recorded in Brazil since 1942, the virus is transmitted among humans by the vector *Aedes aegypti*.^(1,2) In the sylvatic/jungle cycle, the virus is transmitted by wild mosquitoes (mainly belonging to *Haemagogus* and *Sabethes* genera) to non-human primates (NHPs) (e.g., those belonging to *Alouatta* and *Callithrix* genera) and occasionally to un-



²Universidade Federal do Tocantins, Laboratório de Bioinformática e Biotecnologia, Gurupi, TO, Brasil

³Universidade Federal do Rio Grande do Sul, Instituto de Ciências Básicas da Saúde, Porto Alegre, RS, Brasil

Instituto Federal do Norte de Minas Gerais, Laboratório de Comportamento de Insetos, Salinas, MG, Brasil

⁵Secretaria Estadual de Saúde do Rio Grande do Sul, Centro Estadual de Vigilância em Saúde, Porto Alegre, RS, Brasil

⁶Organização Pan-Americana da Saúde/Organização Mundial da Saúde, Brasília, DF, Brasil

⁷Stellenbosch University, School of Data Science and Computational Thinking, Centre for Epidemic Response and Innovation, Stellenbosch, South Africa

⁸Universidade Estadual de Santa Cruz, Departamento de Agricultura e Ciências Ambientais, Ilhéus, BA, Brasil

⁹Secretaria de Saúde do Estado de Minas Gerais, Coordenação Estadual de Vigilância de Arbovírus, Belo Horizonte, MG, Brasil

¹⁰Fundação Ezequiel Dias, Laboratório Central de Saúde Pública, Belo Horizonte, MG, Brasil

¹¹Fundação Oswaldo Cruz-Fiocruz, Instituto Oswaldo Cruz, Laboratório de Flavivírus, Rio de Janeiro, RJ, Brasil

¹²Ministério da Saúde, Coordenação Geral de Vigilância de Arbovírus, Brasília, DF, Brasil

⁺ Corresponding author: camposvet@gmail.com/filipe.vieira@ifnmg.edu.br

https://orcid.org/0000-0002-5948-472X (FSC)

https://orcid.org/0000-0001-9768-4688 (FVSA)

vaccinated humans in close contact with forest areas.⁽³⁾ Yellow fever (YF) is endemic to the tropical rainforest of the Amazon region; from this region, waves of epidemic expansion spread toward other Brazilian regions at irregular intervals of time.⁽³⁾ During these waves, the virus usually reaches the states of Goiás (GO), in the Central-West region, and Minas Gerais (MG), in the Southeast region. Occasionally, the virus is spread to other southeast states, such as São Paulo (SP) (in 2000, 2008-2009, 2017-2018),⁽⁴⁻¹⁰⁾ Rio de Janeiro (RJ) and Espírito Santo (ES) (between 2017 and 2019).⁽¹¹⁻¹⁴⁾ Even less frequently, YFV can be detected in the southernmost states of the country: Rio Grande do Sul (RS) (2001, 2008-2009, 2020-2021),⁽¹⁵⁻¹⁷⁾ Paraná (PR) and Santa Catarina (SC) (2018-2020).⁽¹⁸⁾ The YFV spread outside endemic areas

raises at least three main concerns: 1) the possibility of re-emergence of an urban cycle; 2) the increased risk of YF in humans due to heterogeneous vaccination coverage outside the Amazon region; and 3) the extinction of threatened NHP species – especially the genus *Alouatta*, highly susceptible to YFV.^(5,7,19-23)

During sporadic expansion waves, MG has been an important corridor through which the virus travels before spreading to other Brazilian states. In recent times, major outbreaks occurred in MG in 2000-2003, 2010 and 2015-2018 (Fig. 1), with the latter being the largest sylvatic outbreak reported in the last 80 years. In the 2015-2018 outbreak, two viral sub-lineages were detected, crossing MG by different paths: the YFV MG/SP/ sub-lineage travelled through the west and southwest

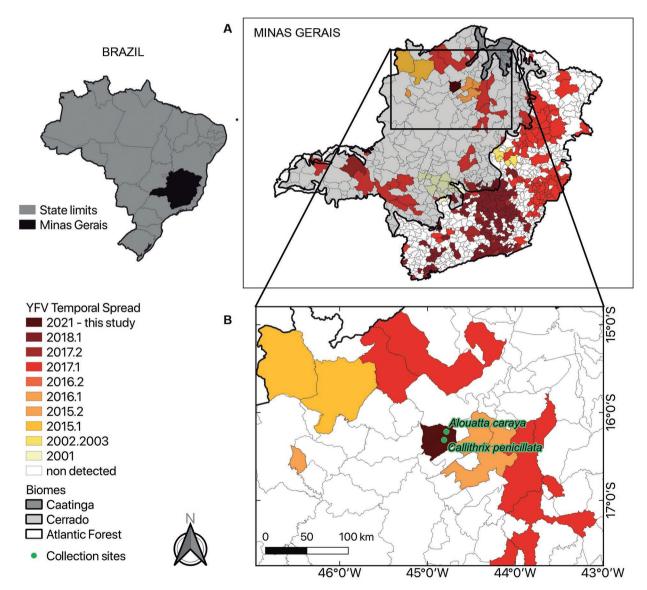


Fig. 1: on the left, Brazil map highlighting the state of Minas Gerais (MG). The Brazilian regional division consists of states and municipalities grouped into regions. MG map showing different municipalities with yellow fever virus (YFV) detections in non-human primates (NHPs) and/or humans per semester and year between 2001 to present (see the caption on the left). The different biomes in MG (Caatinga, Cerrado and Atlantic Forest) are shown in gray (A). Location of epizootics in MG registered on the Sistema de Informação em Saúde Silvestre Georreferenciado (SISS-Geo) platform during this investigation (B).

of the state, before reaching SP; the YFV_{MG/ES/RJ/BA} sublineage spread from the northwest, north, northeast and east of the state, before reaching ES and the state of Bahia (BA).^(14,17,28) Although epizootics have been reported since the end of 2020 (unfortunately, when no sampling was performed), the last reported viral detection in MG occurred in the first quarter of 2018.^(14,29) Nevertheless, the virus was detected in the states of Pará (PA), Tocantins (TO), GO, SP, PR, SC and RS from the end of 2020 to June 2022 (Supplementary Fig. 1),^(18,30,31) raising awareness of the possibility of reintroduction in MG.

In this work, we describe a surveillance effort encompassing the technology of smartphone applications and the concerted action of Coordenação-Geral de Vigilância Arboviroses, Secretaria de Estado de Saúde de Minas Gerais, Laboratório Central de Saude Pública, Brazilian Ministry of Health, and members of Febre Amarela BR project (Portuguese for Yellow Fever BR) that succeeded in the first YFV confirmation in NHPs in MG in 2021, followed by genome sequencing, in only 10 days. Phylogenetic analyses revealed the introduction of a new sub-lineage in the extra-Amazonian region.

MATERIALS AND METHODS

Study area – This study was conducted in MG, which has the largest number of municipalities (853) in Brazil, the second largest population in the country (21,411,923 inhabitants, 10.1% of the Brazilian population) and the fourth largest area (586,513.99 km², larger than Spain, for example) (https://www.ibge.gov.br/cidades-e-estados/mg.html). The study took place in the north of MG, which is predominantly covered by the Cerrado (a savannah-like biome) (Fig. 1). This region has well-defined dry and rainy periods. August, the month of this investigation, is the driest month of the year, during winter. December presents the highest average rainfall, during summer.

Establishing an information network to strengthen epizootic surveillance in northern MG – In 2020, a project sponsored by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) led to the formation of a network that aggregated research institutions and public health services, aimed at strengthening the surveillance of epizootics in several Brazilian regions. (32,33)

For these purposes, preliminary field expeditions to 12 municipalities in MG were conducted. In each municipality, meetings and lectures were organised with public health officials and environmental surveillance officials to educate them on the importance of epizootic surveillance as a prompt sign of YFV in circulation. The agents were trained to notify epizootics using the Sistema de Informação em Saúde Silvestre Georreferenciado (SISS-Geo) web app(34) (or georeferenced wildlife health information system) developed by Fundação Oswaldo Cruz-Fiocruz, attached to the Brazilian Ministry of Health. This platform works as a public repository for collaborative monitoring and disease surveillance of wild animals in Brazil. For monitoring, any person can send pictures of sylvatic animals and mortality (epizootic) in wildlife, as well as metadata. The app collects realtime geographic coordinates and sends information to state and federal surveillance centres. We also organised

fieldwork to train teams on vector and NHP sample collection. Finally, several WhatsApp application groups were created with all trainees, constituting an information network whose main aim was to distribute educational material and exchange news about epizootics. (32,33) An organisation chart showing the steps involved in organising the network is shown in Fig. 2. The Secretaria Estadual de Saúde supported all the field expeditions.

Investigation of the epizootics in northern MG – On August 24, 2021, the information network alerted us about the occurrence of epizootics in two municipalities, Icaraí de Minas and Ubaí, located in the northern region of MG, 595 km from Belo Horizonte, the capital of the state. Both municipalities remained unaffected during the 2015-2018 outbreak. On August 25, 2021, a team comprising members of the Febre Amarela BR project, members of the Secretaria Estadual de Saúde and officials from the affected municipalities were rapidly assembled and dispatched. On August 25 and 26, 2021, the team surveyed the riparian forests of those municipalities, searching for NHP carcasses to collect biological samples. All detected epizootics were registered in the SISS-Geo app and the Sistema de Informação de Agravos de Notificação (SINAN). Samples were collected following safety protocols,(35) preserved in liquid nitrogen (-196°C) and sent to the reference laboratory of the Sistema Único de Saúde (SUS) located at the Fundação Ezequiel Dias (FUNED) in MG, as well as to the laboratory of Febre Amarela BR project for viral diagnosis.

YFV detection – Samples (different fragments of viscera) were lysed with TRIzol™, stored in liquid nitrogen and sent to the sequencing laboratory on dry ice (-80°C). Total RNA extraction with TRIzol was performed according to the manufacturer's instructions. YFV RNA was detected using a previously published by reverse transcription-quantitative polymerase chain reaction (RT-qPCR) protocol. (36)

Genome sequencing - Samples were subjected to cDNA synthesis using the LunaScript™ RT Super-Mix Kit (NEB) following the manufacturer's instructions. Then, multiplex tiling PCR was performed using previously published YFV primers⁽³⁷⁾ with 40 amplification cycles (denaturation: 95°C for 15 s and annealing/extension: 65°C for 5 min) of PCR using Q5 high-fidelity DNA polymerase (NEB). A volume of 7.5 µL of the PCR product was used for DNA library preparation using the Rapid Barcoding Kit 96 SQK-RBK110.96 (Oxford Nanopore Technologies, Oxford, UK) following the manufacturer's instructions. The sequencing library was loaded onto an R9.4 flow cell (Oxford Nanopore Technologies) and sequenced for 6-18 h using the MiNKOW software. The resulting Fast5 files were base called and demultiplexed using Guppy (Version 4.4.2, Oxford Nanopore Technologies). Consensus sequences were generated by de novo assembly using Genome Detective. (38)

Phylogenetic analyses – To perform phylogenetic analyses, we selected all near-complete YFV sequences of South American genotype I, available at National Center for Biotechnology Information (NCBI) (n = 315),

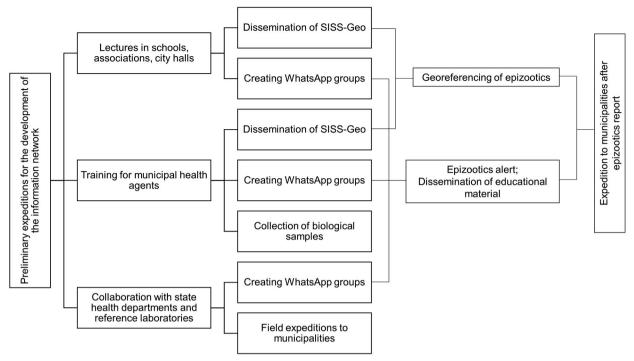


Fig. 2: organization chart showing steps for organising the information network to strengthen yellow fever surveillance. SISS-Geo: Sistema de Informação em Saúde Silvestre Georreferenciado.

excluding sequences < 8 kb and those of vaccine and patent-related viruses. Metadata such as sample collection dates and geographic positioning were retrieved from GenBank files or genome-associated publications (manual curation). The genome named MG66-L (reported here; details below) and 315 genomes from NCBI were aligned using MAFFT v.7.480.⁽³⁹⁾ The subsequent alignment was then used to infer a maximum likelihood tree topology in IQTREE version 2,⁽⁴⁰⁾ using GTR with 1,000 bootstrap replicates. The transfer bootstrap support for splits in the topology was inferred using Booster.⁽⁴¹⁾ The new genome sequences were sent to the NCBI GenBank database under accession numbers OL519587 to OL519589.

RESULTS

On August 25, 2021 (day 1), following the alert of our information network, members of the Febre Amarela BR project found and notified an epizootic affecting six black-and-gold howler monkeys (Alouatta caraya) in Icaraí de Minas (MG), of which only one carcass was suitable for sampling (liver, spleen and kidney samples were collected and named MG66-L, MG66-S and MG66-K, respectively). On August 26 (day 2), another epizootic was detected, affecting six black-tufted marmosets (Callithrix penicillata) in Ubaí, and only one carcass met the conditions for sampling collection (from the liver, spleen, kidney and brain, named MG67-L, MG67-S, MG67-k and MG67-B, respectively). Samples were sent to the laboratory (September 2, day 9) and tested (September 3, day 10). All sampled tissues were positive for YFV by RT-qPCR (Table).

The interval between the alert of the information network and YFV diagnosis was 10 days (Fig. 3). After confirming the virus detection, health authorities were immediately notified. Control measures were intensified, and further investigations were conducted in the affected and surrounding areas. Following diagnosis, a sequencing library was prepared, and sequencing was performed on the same day. There was only an 8-hour interval between RNA extraction and generation of the near-whole genome sequence.

Three near-complete YFV genomes were generated from three different fragments of viscera (liver, spleen and kidney) from the same carcass of *Alouatta caraya* (named MG66). A comparison of the three YFV genomes revealed no intra-host sequence differences. Therefore, the MG66-L genome was used for the phylogenetic analysis.

Such analyses revealed that the YFV genome generated here (MG66 – OL519587|Brazil|MG|Monkey|2021-08-25) was related to the South American genotype I, as expected, and clustered with an isolate (MF370546|Brazil|PA| Monkey|2017-04-05) sampled from an *Alouatta caraya* identified in PA (Amazonian region) in 2017 (Fig. 4A and Supplementary Fig. 2), indicating the introduction of a new YFV sub-lineage in MG, now called YFV Moreover, they were not related to the viruses detected in the same year (2021) in RS, the southernmost state of Brazil. Further analysis of the polyprotein encoded by genomes MF370546 (PA-2017) and OL519587 (MG-2021) revealed only five amino acid differences (T128I, X108I, N808D, V3060I and A3149V), whereas compari-

TABLE
Reverse transcription-quantitative polymerase chain reaction (RT-qPCR) test results,
according to tested species, local, tissues, date and Ct

Host species	Municipality	Geographical coordinates	Sample	Collection date	Ct
Alouatta caraya (MG66)	Icaraí de Minas	16°13'03.3"S 44°47'00.9"W	Liver	August 25, 2021	15
			Spleen	August 25, 2021	14
			Kidney	August 25, 2021	20
Callithrix penicillata (MG67)	Ubaí	16°18'42.0"S 44°48'36.0"W	Liver	August 26, 2021	31
			Spleen	August 26, 2021	30
			Kidney	August 26, 2021	31
			Brain	August 26, 2021	29

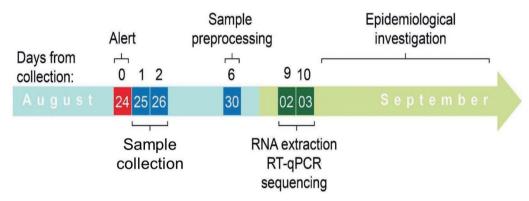


Fig. 3: timeline showing the time spent from the collection of samples to the generation of near-complete genomes. RT-qPCR: reverse transcription-quantitative polymerase chain reaction.

son of sequences OL519587 and MZ712133 (RS-2021) revealed nine amino acid differences. These findings confirm the expansion and circulation of at least two YFV sub-lineages (YFV_{PA/MG} and YFV_{MG/SP/RS}) in the extra-Amazon region in 2021. (17) The nucleotide difference matrix between the selected genomes is shown in Fig. 4B.

DISCUSSION

MG has historically been affected by several YFV introductions. However, owing to limited logistics, it has been difficult to conduct surveillance and collect biological samples promptly, both of which require a robust field and laboratory surveillance infrastructure. In this study, we describe the results of setting up an inter-institutional network capable of collaborative actions to investigate epizootics, which resulted in the first confirmation of YFV circulation in MG, Brazil, in 2021.

MG has played a role as an YFV dissemination route during expansion waves from the Amazon region. Despite the limited resources available in the past, studies on viral dispersion routes in the region have been conducted since 1938 (Supplementary Fig. 3).^(24,42) In the last 2 decades, outbreaks that resulted in human cases were recorded in 2001 (32 human cases), 2002-2003 (63 human cases) and two isolated cases between 2008-2009. The largest outbreak occurred between 2016-2018, with

1,006 human cases and 448 confirmed epizootic cases. (26,27,43,44) Importantly, there were no previous records of viral circulation in the two affected municipalities, suggesting the presence of a naive NHP population. (45,46)

Since 2019, to the best of our knowledge, despite the notification of several epizootics, no new reports on YFV circulation have been published in the north of MG.⁽²⁹⁾ The wide territorial extension, with huge rural areas far from urban and research centres, combined with socioeconomic inequality and the lack of trained professionals in several municipalities, made it difficult to implement appropriate surveillance of epizootics, thus significantly reducing the chances of timely sample collection for virus detection. These challenges reaffirm the need to introduce new tools and strategies to strengthen surveillance. Among them, the following points are worth mentioning. a) The use of smartphone applications to notify epizootics and collect the geographic coordinates of the occurrence in real time. In this sense, the SISS-Geo app, which has been progressively implemented in Brazil, speeds up the arrival of information to involved institutions and provides accurate geographic location and streamlines outbreak response. (34) b) The use of instant messaging apps increases the number of "watchers," providing information through real-time text transmission. Creating WhatsApp groups with a lo-

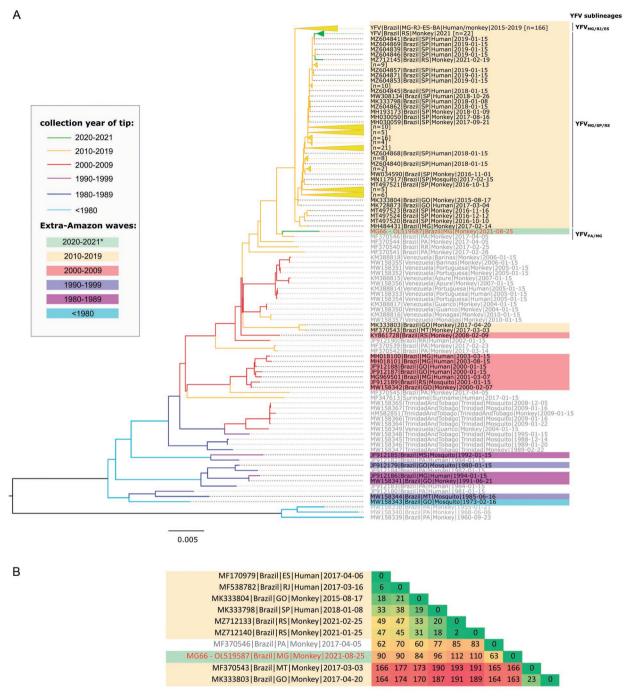


Fig. 4: maximum likelihood phylogenetic tree of South American genotype I yellow fever virus (YFV) strains, highlighting in red the MG66-L strain isolated in the southeastern state of Minas Gerais. Branches colours indicate the year of sample collection. Isolates from Amazon region are labelled in gray and isolates from Extra-Amazon region are labelled in black and highlighted by decade of wave (A). The sub-lineage YFV_{MG/ES/RJ} is collapsed at the top of the tree. Nucleotide differences between selected genomes (B).

cal or regional reach, comprising different members of the population (health agents, cyclists, hikers, students, rural workers, etc.), increases the chances of detecting epizootics and speeding up the dissemination of important information such as vaccination campaigns, NHPs and ecosystem conservation efforts. (32,33) Ultimately, this network of professional and citizen-driven wildlife dis-

ease surveillance serves both public health and science popularisation, the latter being a crucial yet neglected part in epidemic management. c) The creation of multi-institutional networks to facilitate and speed up sample collection and diagnosis. In the present study, research institutions linked to the Febre Amarela BR project provided an integrated, multidisciplinary team, aggregating

staff from universities, students and governmental organs at municipal, state and federal health secretariats whose coordinates and combined actions allowed the achievement of the results reported here.

Genomic surveillance efforts have shown that the virus circulating in MG was related to the virus circulating in PA, in the Amazon region, in 2017 (sub-lineage YFV_{PA/MG}). This reveals a new wave of viral expansion in MG, 6 years after the outbreak associated with the YFV_{MG/SP/RS} and YFV_{MG/ES/RJ/BA} sub-lineages, between 2015 and 2018. Interestingly, in 2021, at least two different viral sub-lineages were found circulating at the same time in the extra-Amazon region, represented by YFV_{MG/SP/RS}, detected in RS,⁽¹⁷⁾ and YFV_{PA/MG}. Notably, the latter was detected at the height of the dry season in one of the driest regions of MG, which makes it more difficult for the main species of mosquito vectors (i.e., *Haemagogus janthinomys* and *Hg. leucocelaenus*) to survive.⁽¹³⁾

In this study, the importance of coordinated efforts between local populations and different institutions is emphasised to increase surveillance capacity. Through this system, it was possible to confirm YFV occurrence, which enabled the establishment of timely response measures. Genomic surveillance has allowed the identification of a new YFV expansion wave in MG, which needs to be monitored to ensure that effective preventive measures are implemented in appropriate time and place. These include an increase in vaccine coverage in humans and improved communication regarding the risk of YFV to health professionals and the general population to avoid human cases. This study also highlights the need for awareness and public information on the role of NHPs as sentinels for the occurrence of YFV. Finally, this study highlights the importance of collaborative, integrated surveillance, allowing prompt action in the event of YF suspicion.

ACKNOWLEDGEMENTS

To the contributions of the Divisão de Vigilância Sanitária e Ambiental from MG, to the effort of the Grupo de Vigilância da Febre Amarela of MG that was at the forefront of the preparation for virus outbreak response and field investigations, to Beatrízio Rodrigues Almeida, Hermes Almeida and Joselio Ribeiro Paraíso, for their valuable help during the field work, to Aline Tátila Ferreira, for her help in creating images, to Fernando Lucas Melo, for their valuable help with genome sequencing, to countless colleagues from municipalities' health departments, who conducted the investigation of epizootics collecting samples in the field and to the Coordenação-Geral de Vigilância Arboviroses, Brazilian Ministry of Health. The Febre Amarela BR project (https://www.febreamarelabr.com. br/) is supported by grants from Conselho Nacional de Desenvolvimento Científico e Tecnológico/Departamento de Ciência e Tecnologia/Secretaria de Ciência, Tecnologia e Insumos Estratégicos/Brazilian Ministry of Health (CNPq/Decit/ SCTIE/MS) (grant number 443215/2019-7). MSA is granted a post-doctoral scholarship (DTI-A) from CNPq. ACF, FSC, BMR and PMR are CNPq research fellows. FVSA is on receipt of a grant (APQ-01403-21) from Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG) and (21/2020) from Instituto Federal do Norte de Minas Gerais (IFNMG).

AUTHORS' CONTRIBUTION

Conceived and designed the experiments: FVSA, MSA, FSC, AASC, JCC, ES, DST, GRA, ACF, BMR, PMR and MABA. Performed the experiments: MSA, VF, MAP, LOM, SMAT, MEGS, JCC, ES, LCB, CMDS, NFDM, CHO, AJJS, SBV, MAMG, GGM, LOM and MABA. Analysed the data: MSA, FSC, VF, AASC, FVSA, APS, JCC, ES, DST, APMR, BMR, AODT, PMR and MABA. Contributed reagents/materials/analysis tools: ACF, BMR, PMR, AODT, DCCC and MABA. Contributed to the writing of the manuscript: MSA, FSC, VF, AASC, MABA, APS, JCC, ES, AODT, DST, SBV, APMR, BMR, PMR and FVSA. All authors have read and agreed to the published version of the manuscript. We declare that no conflict of interest exists.

REFERENCES

- Monath TP, Vasconcelos PFC. Yellow fever. J Clin Virol. 2015; 64: 160-73.
- Franco O. História da febre amarela no Brasil. Rev Bras Malariol Doenças Trop. 1969; 21: 315-20.
- Possas C, Lourenço-de-Oliveira R, Tauil PL, Pinheiro F de P, Pissinatti A, Cunha RV da, et al. Yellow fever outbreak in Brazil: the puzzle of rapid viral spread and challenges for immunisation. Mem Inst Oswaldo Cruz. 2018; 113: e180278.
- Moreno ES, Agostini I, Holzmann I, Di Bitetti MS, Oklander LI, Kowalewski MM, et al. Yellow fever impact on brown howler monkeys (*Alouatta guariba clamitans*) in Argentina: a metamodelling approach based on population viability analysis and epidemiological dynamics. Mem Inst Oswaldo Cruz. 2015; 110: 865-76.
- Moreno ES, Rocco IM, Bergo ES, Brasil RA, Siciliano MM, Suzuki A, et al. Reemergência de febre amarela: detecção de transmissão no estado de São Paulo, Brasil, 2008. Rev Soc Bras Med Trop. 2011; 44: 290-6.
- Vasconcelos PF, Costa ZG, Travassos da Rosa ES, Luna E, Rodrigues SG, Barros VL, et al. Epidemic of jungle yellow fever in Brazil, 2000: implications of climatic alterations in disease spread. J Med Virol. 2001; 65: 598-604.
- Romano APM, Costa ZGA, Ramos DG, Andrade MA, Jayme V de S, Almeida MAB de, et al. Yellow fever outbreaks in unvaccinated populations, Brazil, 2008-2009. PLoS Negl Trop Dis. 2014; 8: e2740.
- Cunha MDP, Duarte-Neto AN, Pour SZ, Ortiz-Baez AS, Černý J, Pereira BB de S, et al. Origin of the São Paulo yellow fever epidemic of 2017-2018 revealed through molecular epidemiological analysis of fatal cases. Sci Rep. 2019; 9: 20418.
- Camargo-Neves VLF de, Poletto DW, Rodas LAC, Pachioli ML, Cardoso RP, Scandar SAS, et al. Entomological investigation of a sylvatic yellow fever area in São Paulo state, Brazil. Cad Saude Publica. 2005; 21: 1278-86.
- 10. Souza RP de, Petrella S, Coimbra TLM, Maeda AY, Rocco IM, Bisordi I, Silveira VR, et al. Isolation of yellow fever virus (YFV) from naturally infected *Haemagogus (Conopostegus) leucocelae-nus* (Diptera: Culicidae) in São Paulo State, Brazil, 2009. Rev Inst Med Trop S Paulo. 2011; 53: 133-9.
- 11. Fernandes NCC de A, Cunha MS, Guerra JM, Réssio RA, Cirqueira C dos S, Iglezias SD, et al. Outbreak of yellow fever among nonhuman primates, Espírito Santo, Brazil, 2017. Emerg Infect Dis. 2017; 23: 2038-41.

- Gómez MM, Abreu FVS de, Santos AACD, Mello IS de, Santos MP, Ribeiro IP, et al. Genomic and structural features of the yellow fever virus from the 2016-2017 Brazilian outbreak. J Gen Virol. 2018; 99: 536-48.
- 13. Abreu FVS de, Ribeiro IP, Ferreira-de-Brito A, Santos AAC dos, Miranda RM de, Bonelly I de S, et al. *Haemagogus leucocelaenus* and *Haemagogus janthinomys* are the primary vectors in the major yellow fever outbreak in Brazil, 2016-2018. Emerg Microbes Infect. 2019; 8: 218-31.
- 14. Delatorre E, Abreu FVS de, Ribeiro IP, Gómez MM, dos Santos AAC, Ferreira-de-Brito A, et al. Distinct YFV Lineages co-circulated in the central-western and southeastern Brazilian regions from 2015 to 2018. Front Microbiol. 2019; 10: 1079.
- Almeida MAB de, Santos E dos, Cardoso J da C, Fonseca DF da, Noll CA, Silveira VR, et al. Yellow fever outbreak affecting *Al-ouatta* populations in southern Brazil (Rio Grande do Sul State), 2008-2009. Am J Primatol. 2012; 74: 68-76.
- 16. Vasconcelos PF, Sperb AF, Monteiro HA, Torres MA, Sousa MR, Vasconcelos HB, et al. Isolations of yellow fever virus from *Hae-magogus leucocelaenus* in Rio Grande do Sul state, Brazil. Trans R Soc Trop Med Hyg. 2003; 97: 60-2.
- 17. Andrade M de S, Campos FS, Campos AAS, Abreu FVS, Melo FL, Sevá A da P, et al. Real-time genomic surveillance during the 2021 re-emergence of the yellow fever virus in Rio Grande do Sul state, Brazil. Viruses. 2021; 13: 1976.
- 18. Ministério da Saúde/Secretaria de Vigilância em Saúde. Informe quinzenal sarampo Brasil, semanas epidemiológicas 43 de 2020 a 1 de 2021. Bol Epidemiol. 2021; 52(4). Available from: https://www.gov.br/saude/pt-br/centrais-de-conteudo/publicacoes/boletins/epidemiologicos/edicoes/2021/boletim_epidemiologico_svs 4.pdf. [accessed on 24 September 2022].
- Holzmann I, Agostini I, Areta JI, Ferreyra H, Beldomenico P, Di Bitetti MS. 2010. Impact of yellow fever outbreaks on two howler monkey species (*Alouatta guariba clamitans* and *A. caraya*) in Misiones, Argentina. Am J Primatol. 2010; 72: 475-80.
- Bicca-Marques JC, Calegaro-Marques C, Rylands A, Strier K, Mittermeier R, Almeida MA, et al. Yellow fever threatens Atlantic Forest primates. Sci Adv. 2017; 3: e1600946/tab-e.
- Dietz JM, Hankerson SJ, Alexandre BR, Henry MD, Martins AF, Ferraz LP, et al. Yellow fever in Brazil threatens successful recovery of endangered golden lion tamarins.
 Sci Rep. 2019; 9: 12926.
- 22. Strier KB, Tabacow FP, de Possamai CB, Ferreira AIG, Nery MS, de Melo FR, et al. Status of the northern Muriqui (*Brachyteles hypoxanthus*) in the time of yellow fever. Primates. 2019; 60: 21-8.
- Berthet M, Mesbahi G, Duvot G, Zuberbühler K, Cäsar C, Bicca-Marques JC. Dramatic decline in a titi monkey population after the 2016-2018 sylvatic yellow fever outbreak in Brazil. Am J Primatol. 2021; 83: e23335.
- Laemmert HW, Hughes TP, Causey OR. The invasion of small forests by yellow fever virus as indicated by immunity in Cebus monkeys. Am J Trop Med 1949; s1-29: 555-65.
- 25. Pinheiro GG, Rocha MN, de Oliveira MA, Moreira LA, Andrade Filho JD. 2019. Detection of yellow fever virus in sylvatic mosquitoes during disease outbreaks of 2017-2018 in Minas Gerais state, Brazil. Insects 10(5): 136.
- 26. Costa ZGA. Estudo das características epidemiológicas da febre amarela no Brasil, nas áreas fora da Amazônia Legal, no período de 1999 a 2003 [dissertation]. Rio de Janeiro (RJ): Fundação Osvaldo Cruz, Escola Nacional de Saúde Pública Sérgio Arouca; 2005.
- 27. Ribeiro M, Antunes CM de F. Febre amarela: estudo de um surto. Rev Soc Bras Med Trop. 2009; 42: 523-31.

- Jesus JG de, Gräf T, Giovanetti M, Mares-Guia MA, Xavier J, Maia ML, et al. Yellow fever transmission in non-human primates, Bahia, northeastern Brazil. PLoS Negl Trop. 2020; 14: e0008405.
- 29. SES/SUBVS-SVE-DVAT-CEVARB. 2021. Nota Informativa SES/SUBVS-SVE-DVAT-CEVARB 1877/2021, SES-Nota Informativa 1877. Available from: https://www.saude.mg.gov.br/images/noticias_e_eventos/000_2021/SEI_1320.01.0038355_2021_82 NI SES 1877-2021 - ALERTA FA.pdf
- 30. Ministério da Saúde/Secretaria de Vigilância em Saúde. Monitoramento dos casos de arboviroses até a semana epidemiológica 29 de 2022. Bol Epidemiol. 2022; 53(Ago). Available from: https://www.gov.br/saude/pt-br/centrais-de-conteudo/publicacoes/boletins/epidemiologicos/edicoes/2022/boletim-epidemiologico-vol-53-no29. [accessed on 24 September 2022].
- 31. Ministério da Saúde/Sistema Único de Saude/DataSUS. Febre Amarela em humanos e primatas não-humanos – 1994 a 2021. Available from: https://opendatasus.saude.gov.br/dataset/febre-amarela-em-humanos-e-primatas-nao-humanos. [accessed on 25 September 2022].
- 32. Abreu FVS, Santos E dos, Gomes MQ, Vargas WP, Passos PH de O, Silva CNE, et al. Capture of *Alouatta guariba clamitans* for the surveillance of sylvatic yellow fever and zoonotic malaria: which is the best strategy in the tropical Atlantic Forest? Am J Primatol. 2019: 81: e23000.
- 33. Abreu FVS, Delatorre E, dos Santos AAC, Ferreira-de-Brito A, de Castro MG, Ribeiro IP, et al. Combination of surveillance tools reveals that yellow fever virus can remain in the same Atlantic Forest area at least for three transmission seasons. Mem Inst Oswaldo Cruz. 2019; 114: e190076.
- 34. Chame M, Barbosa H, Gadelha L, Augusto D, Krempser E, Abdalla L. SISS-Geo: leveraging citizen science to monitor wildlife health risks in Brazil. J Healthc Inform Res. 2019; 3: 414-40.
- 35. Romano A, Ramos D, Araujo F, Leal S, Costa Z, Silva A, et al. Guia de vigilância de epizootias em primatas não humanos e entomologia aplicada à vigilância da febre amarela. Brasília, DF: Ministério da Saúde; 2017.
- 36. Domingo C, Patel P, Yillah J, Weidmann M, Méndez JA, Nakouné ER, et al. Advanced yellow fever virus genome detection in point-of-care facilities and reference laboratories. J Clin Microbiol. 2012; 50: 4054-60.
- 37. Faria NR, Kraemer MUG, Hill SC, Goes de Jesus J, Aguiar RS, Iani FCM, et al. Genomic and epidemiological monitoring of yellow fever virus transmission potential. Science. 2018; 361: 894-9.
- 38. Vilsker M, Moosa Y, Nooij S, Fonseca V, Ghysens Y, Dumon K, et al. Genome Detective: an automated system for virus identification from high-throughput sequencing data. Bioinformatics. 2019; 35: 871-3.
- 39. Katoh K, Standley DM. MAFFT Multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol. 2013; 30: 772-80.
- Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. IQ-TREE: a fast and effective stochastic algorithm for estimating maximumlikelihood phylogenies. Mol Biol Evol. 2015; 32: 268-74.
- 41. Lemoine F, Domelevo Entfellner J-B, Wilkinson E, Correia D, Dávila Felipe M, De Oliveira T, et al. Renewing Felsenstein's phylogenetic bootstrap in the era of big data. 7702. Nature. 2018; 556: 452-6.
- 42. Casa de Oswaldo Cruz. Item 0005 Mapa demonstrando a disseminação da febre amarela no sul do Brasil, 1932-1942. Available from: http://arch.coc.fiocruz.br/index.php/y0luj. [accessed on 17 May 2022].

- 43. Secretaria de Estado de Saúde de Minas Gerais. Febre amarela silvestre em Minas Gerais. Boletim epidemiológico especial 18/10/2018. Available from: https://www.saude.mg.gov.br/images/ noticias_e_eventos/000_2018/BoletinsEpidemiologicos/Boletim_-_Febre_Amarela_2018_-_Consolidado_V3.pdf
- 44. Secretaria de Estado de Saúde de Minas Gerais). Febre amarela silvestre em Minas Gerais. Boletim epidemiológico 19/02/2019. Available from: https://www.saude.mg.gov.br/images/noticias_e_eventos/000_2019/jane_fev_mar/Febre_Amarela/Boletim_atualiza%C3%A7%C3%A3o_FA_12-02-2019.pdf
- 45. de Azevedo Fernandes NCC, Guerra JM, Díaz-Delgado J, Cunha MS, Saad L delC, Iglezias SD, et al. Differential yellow fever susceptibility in New World nonhuman primates, comparison with humans, and implications for surveillance. Emerg Infect Dis 2021; 27: 47-56.
- 46. Mares-Guia MAM de M, Horta MA, Romano A, Rodrigues CDS, Mendonça MCL, dos Santos CC, et al. Yellow fever epizootics in non-human primates, Southeast and Northeast Brazil (2017 and 2018). Parasit Vectors. 2020; 13: 90.