

Acute effects of ropivacaine hydrochloride on corneal endothelial cell ultrastructure of horses: *ex vivo* study

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ABSTRACT: The objective of this study was to evaluate the acute effects of ropivacaine hydrochloride on the corneal endothelium of horses. Forty-eight eyes were obtained from a commercial slaughterhouse and were randomly divided into three groups. In group *A*, the corneal endothelium was exposed to 0.75% ropivacaine hydrochloride for 60 seconds. In group *B*, the corneal endothelium was exposed to 0.75% ropivacaine hydrochloride for 15 minutes. In group *C*, the corneal endothelium was exposed to a balanced saline solution for 60 seconds. Afterwards, all samples were prepared for evaluation with scanning electron microscopy. Random electromicrographs were obtained from each sample. The images were analysed and, with the aid of software, areas with no endothelial cells were measured. The average endothelial loss, expressed as a percentage in relation to the total area, of the samples in group *A* was 5.28%. The average endothelial loss of samples from group *B*, expressed as a percentage in relation to the total area, was 20.39%. The damage to the corneal endothelium was significantly greater in group *B* compared to groups *A* and *C*. It was possible to conclude that 0.75% ropivacaine hydrochloride induced acute damage to corneal endothelium cells.

Key words: equine, intraocular surgery, cornea, intracameral anaesthesia.

Efeitos agudos do cloridrato de ropivacaína na ultraestrutura das células endoteliais da córnea de equinos: estudo *ex vivo*

RESUMO: Objetivou-se avaliar os efeitos agudos do cloridrato de ropivacaína no endotélio da córnea de equinos. Quarenta e oito olhos de equinos foram divididos aleatoriamente em três grupos. No grupo A o endotélio da córnea foi exposto a cloridrato de ropivacaína a 0,75% por 60 segundos. No grupo B o endotélio da córnea foi exposto a cloridrato de ropivacaína a 0,75% por 60 segundos. No grupo B o endotélio da córnea foi exposto a cloridrato de ropivacaína a 0,75% por 60 segundos. No grupo B o endotélio da córnea foi exposto a cloridrato de ropivacaína a 0,75% por 60 segundos. No grupo B o endotélio da córnea foi exposto a solução salina balanceada por 60 segundos. As amostras foram preparadas para avaliação com microscopia eletrônica de varredura. Eletromicrografias eletrônicas de varredura foram obtidas aleatoriamente de cada amostra. As imagens foram analisadas e, com o auxílio de um programa para morfometria foram medidas as áreas sem células endoteliais. A perda endotelial média foi expressa em porcentagem em relação à área total das amostras do grupo A foi de 5,28%. A perda endotelial média de amostras do grupo B foi expressa em porcentagem em relação à área total, foi de 20,39%. O dano ao endotélio da córnea foi significativamente maior no grupo B, comparado aos grupos A e C. O cloridrato de ropivacaína a 0,75% induziu dano agudo nas células do endotélio da córnea de equinos. **Palavras-chave**: equinos, cirurgia intraocular, córnea, anestesia intracameral.

1 INTRODUCTION

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A minimum number of endothelial cells 4 is vital for maintaining the transparency of the 5 cornea (PARIKH & EDELHAUSER, 2003). A loss 6 of corneal endothelial cells always occurs during 7 surgical procedures to remove cataracts. Moreover, 8 the corneal endothelium has minimal capacity for 9 mitosis and high cell losses may lead to vision loss. 10 In order to minimise the loss of endothelial cells and avoid corneal decompensation after intraocular surgery, it is important that solutions that are toxic to the corneal endothelium are not used inside the eye (ISHIKAWA, 2002). Any substance that may cause toxicity to the corneal endothelium must be carefully evaluated before being routinely used inside the anterior chamber. The use of intracameral anaesthesia has become frequent as an alternative to traditional blocks due to its ease of use and analgesic comfort during and after the procedure in humans 1

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(PARK et al. 2010; WANG, 2013). VÄ LIMÄKI 1 2 TÖRNBLOM compared viscoanaesthesia and 3 and intracameral anaesthesia with 1% lidocaine in 4 patients undergoing cataract surgery. Patients who 5 are given viscoanaesthesia may be at increased risk for postoperative corneal oedema (VÄLIMÄKI & 6 7 TÖRNBLOM, 2009). Some studies have already been developed to analyse the effects of intraocular 8 9 injection of lidocaine on the corneal endothelium. 10 Effect of intracameral ropivacaine on corneal 11 endothelium was studied (ÇAÇA et al, 2006). 12 Cataracts are an important cause of blindness in 13 horses and, at present, the only form of treatment is 14 surgical removal of the opaque lens (EDELMANN et 15 al, 2014; BROOKS et al. 2014; TOWNSEND, 2017). 16 To our knowledge, there are no studies evaluating 17 the toxicity of intracameral local anaesthetics on the 18 corneal endothelium of horses.

19 The aim of this study was to evaluate the 20 acute ex vivo effects of ropivacaine hydrochloride 21 in the ultrastructure of the corneal endothelial cells 22 of horses.

24 MATERIALS AND METHODS

26 Forty-eight corneas from 24 horses, male 27 or female, of different ages were studied. The eyes 28 were obtained from a licensed Brazilian commercial 29 slaughterhouse (xxx). The experiment was carried 30 out according to the standards of the Association for 31 Research in Vision and Ophthalmology (ARVO). 32 Immediately after slaughter, all eyes were examined. 33 With slit-lamp biomicroscopy (Portable Slit Lamp 34 SL 15, Kowa, Japan) the ocular surface, anterior chamber, and lens were examined. Corneal stain 35 with fluorescein (Fluorescein, Allergan, SP, Brazil) 36 37 was made. Eye bulbs that showed evidence of eye 38 disease were excluded. Immediately after the eye 39 exam, enucleation was performed, and the eye bulbs 40 were kept in a humid chamber until the corneas were 41 collected. The corneoscleral buttons were removed 42 with a scalpel and scissors. Corneas were then excised 43 with an eight mm diameter trephine. All corneas 44 were analyzed within 4 hours of death. Corneas were 45 randomly divided into three groups.

46 In group A (12 corneas), with a dropper 47 0.2 ml of 0.75% atracurium besylate (Cristália, São Paulo, Brazil) was dripped onto the endothelium. 48 Three minutes after the sample was rinsed with 49 balanced salt solution (BSS) (Halex Istar, ophthalmic 50 51 solution, GO, Brazil) to remove the atracurium 52 besylate. In group B (12 corneas), with a dropper 0.2 53 ml of 0.75% atracurium besylate was dripped onto the endothelium. Fiftheen minutes after the sample was rinsed with BSS to remove the atracurium besylate. In group C (24 corneas), with a dropper 0.2 ml of BSS was dripped onto the endothelium. All samples were kept in 2.5% glutaraldehyde, in 0.1M sodium cacodylate buffer and at pH 7.4 for 24 hours. The corneas were removed from the glutaraldehyde solution, washed in sodium cacodylate buffer solution and dehydrated in ascending concentrations of acetone (30, 50, 70, 80, 90 for 10 minutes at each concentration and for 20 min at 90% again). Afterwards the samples were left for 30 min in 100% acetone and were subjected to drying at a critical point with liquid carbon dioxide. The samples were fixed in stubs with adhesive tape and metallized with goldpalladium. The posterior endothelial surfaces were examined with a scanning electron microscope (JSM 6060, JEOL, Tokyo, Japan) operating at 15 kV. From each sample, five electron micrographs were obtained with 950x magnifications. Occasionally, images with a 30x magnification were obtained to have a panoramic view of each sample. The areas devoid of endothelial cells were calculated. The morphometric study was performed using ImageJ software (ImageJ 1.51k), which allowed calculation of the percentage of cell loss use surrounding areas with no endothelial cells. All analyses were performed by the same examiner. The distribution of variables was assessed using the Shapiro-Wilk test, which indicated a nonnormal distribution. The Friedman test was used to compare the sum of areas expressed as a percentage of the total between treatments. The Wilcoxon test was used to detect the differences and its P value was corrected by the Bonferroni test, due to the multiple comparisons made. Differences were considered statistically significant when $P \le 0.05$.

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RESULTS

With scanning electron microscopy (SEM), it was possible to observe, analyse and obtain images of the corneal endothelium in all samples analysed. In the samples of the control group, there were no areas with endothelial damage and the regular pattern of endothelial cells was observed in all samples (Figure 1). Endothelial losses were observed both in group A and group B samples (Figure 2). The average endothelial loss, expressed as a percentage in relation to the total area, of the samples in group A was 5.28±2.49%. There was a statistically significant difference significant among all compared groups. The average endothelial loss from samples from group B, expressed as a percentage in relation to the

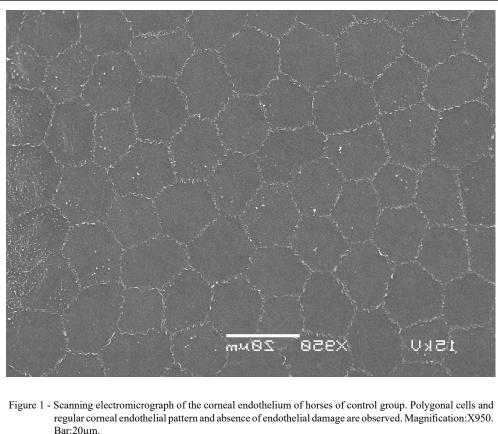
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1 total area, was $20.39 \pm 10.46\%$. The values found in 2 the sum of the areas expressed as a percentage of cell 3 loss were higher in group B when compared with 4 group A (P = 0.006) and group C (P = 0.003). There 5 was also a significant difference between the values 6 found in A in relation to C (P = 0.005).

8 DISCUSSION 9

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10 A minimum endothelial density is essential for the cornea to maintain its transparency. 11 In addition to the surgical procedure, numerous 12 other factors, including intraocular anaesthetics and 13 14 intraocular dyes, can cause endothelial damage. 15 These factors can compromise the function of 16 the endothelium, mainly because in most of the 17 studied species the mitotic activity is limited in 18 this layer (NAUTSCHER et al, 2015). To avoid 19 endothelial decompensation, it is important to 20 choose substances that do not damage the corneal endothelium (KHABAK et al. 2006; BORAZAN et

al. 2009; LEE et al, 2016). The loss of a large number 1 2 of endothelial cells can cause irreversible loss of corneal transparency (PARIKH & EDELHAUSER, 3 4 2003). In this sense, studies on the toxicity of drugs 5 in the corneal endothelium are extremely important. Intracameral anaesthesia have been popularised 6 7 as new techniques for use in cataract surgery in humans. These anaesthetic routes have potential 8 safety advantages over traditional techniques such 9 10 as retrobulbar and peribulbar anaesthesia (OLMEZ et al, 2004). Regarding intracameral anaesthesia 11 in humans and other animal species, studies were 12 carried out evaluating the toxicity of the corneal 13 endothelium and the comfort of patients (TAN & 14 BURTON, 2000; BORAZAN et al. 2009; PARK et al. 15 2010). However, there is a lack of research in regard 16 to equine species and intracameral anaesthesia. This, 17 and the importance of the theme, helped to motivate 18 this study. In addition, cataracts are an important 19 20 cause of blindness in horses and, at present, the only form of treatment is surgical removal of the opaque 21

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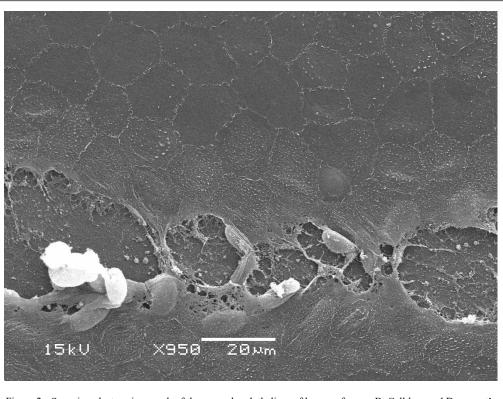


Figure 2 - Scanning electromicrograph of the corneal endothelium of horses of group B. Cell loss and Descemet's membrane exposure are observed. Magnification:X950. Bar:20µm.

lens (BROOKS et al, 2014; EDELMANN et al, 2014; 1 TOWNSEND, 2017). In the present research, an ex 2 3 vivo study was chosen. Normally, research related to 4 the toxicity of intraocular drugs using live animals is 5 carried out on laboratory animals. Previous studies carried out on animal eyes determined that within six 6 7 hours after death, the endothelium can be analysed 8 without structural changes occurring in this layer 9 (SIT et al, 2001; PIGATTO et al, 2005; PIGATTO et al, 2009; FAGANELLO et al, 2016). The use of 10 slaughtered animal eyes for endothelial analysis has 11 12 increasingly been shown to be an alternative to the 13 use of live animals to assess the toxicity of intraocular 14 drugs (PESCOSOLIDO et al, 2011; WEN et al, 2015; TERZARIOL et al, 2016; SILVA et al, 2018; JIANG 15 16 et al, 2018).

Corneal endothelial toxicity is related
to substances that come into contact with the
endothelium, based on their chemical compositions,
pH and osmolarities (PARIKH & EDELHAUSER,
2003). The final preparation of ropivacaine for
clinical use is presented with a pH ranging between

4.0 and 6.0 (RAMOS et al, 2000). Analysis of the 1 corneal thickness data and interpretation of the 2 3 scanning electron micrograph reveals that outside of the pH range of 6.5 to 8.5, structural and functional 4 alterations occur (GONNERING et al, 1979). 5 6 Endothelial losses were observed both in group A and 7 group B samples. In the present study, ropivacaine 8 was kept in direct contact with the endothelium and 9 direct contact with the corneal endothelium in order to verify whether it would induce endothelial damage. 10 In previous studies this methodology has already 11 been used with excellent results (CACA et al, 2006; 12 KHABAK et al, 2006; PESCOSOLIDO et al. 2011; 13 WEN et al, 2015; TERZARIOL et al, 2016; SILVA et 14 15 al, 2018). In other studies, however, an intracameral injection of the substance was performed, which was 16 analysed (BORAAN et al, 2009). Thus, dilution of 17 the substance in aqueous humour could minimise 18 its toxic effects on the endothelium. In the present 19 study, as we were unaware of the possible damage 20 21 of the tested substance, corneal trepanation and direct exposure of the anaesthetic to the corneal 22

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1 endothelium were chosen. In addition, with the 2 intracameral injection of the tested substance it would 3 be difficult to have a homogeneous direct contact 4 between ropivacaine and the corneal endothelium. 5 In the present study, ropivacaine directly contacted 6 with the endothelium across the entire area that was 7 later analysed. Ropivacaine is a local anaesthetic of the amide type with a long duration of action, and 8 9 its effect occurs through a reversible inhibition of the 10 influx of sodium ions in the nerve fibres. It has a high 11 degree of sensory motor differentiation, which can be 12 useful when motor block is undesirable (KUTHIALA & CHAUDHARY, 2011). Commercially available 13 14 ropivacaine does not contain preservatives in its 15 formulation. At this concentration, this anaesthetic 16 is widely used in blocking ophthalmology because 17 it promotes analgesia in some patients for up to 12 18 hours, being superior to lidocaine (KUTHIALA & CHAUDHARY, 2011). In previous studies in humans, 19 20 1% ropivacaine was more effective in analgesia and 21 did not promote significant endothelial cell loss when 22 compared to 2% lidocaine (MARTINI et al, 2002; 23 IACOBELLI et al. 2005).

24 In studies carried out on other animal 25 species, ropivacaine in concentrations between 0.1% 26 and 0.5% was demonstrated to be safe and provide 27 pain relief, but at concentrations of 1%, ropivacaine 28 induced lesions in the corneal endothelium (KLAMT 29 et al, 2003; CACA et al, 2006; BORAZAN et al, 2009). 30 In one study only, 0.75% ropivacaine was observed to 31 induce toxicity in the corneal endothelium of rabbits 32 (KHAZBAK et al, 2006). Among the techniques 33 normally used to prove the toxicity of drugs in the 34 corneal endothelium, optical microscopy associated 35 with alizarin red and SEM stand out (CACA et 36 al, 2006; SCHELINNI et al, 2007; SEGARRA 37 et al, 2018; SILVA et al, 2018). The use of optical 38 microscopy after staining the endothelium with vital 39 dyes has been shown to be a simple, fast and practical 40 way to detect cell damage as well as to analyse the 41 shape of endothelial cells (FAGANELLO et al, 2016). 42 In the present study, SEM was chosen because it has 43 already been widely used in studies related to corneal 44 morphology, the toxicity of intracameral drugs, the 45 effectiveness of means of corneal preservation and in the evaluation of the endothelium ultrastructure of 46 47 different species (OJEDA et al, 2001; PIGATTO et al, 48 2009; TERZARIOL et al, 2016). In the present study, 49 the methodology employed proved to be feasible, 50 allowing images to be obtained, and an analysis and quantification of endothelial cell losses. With SEM, due to the large increase in images, it is possible to delimit the edges of areas with cell loss, thus allowing

establishment of the percentage of endothelial loss. It was possible to visualise the acute toxic effects caused by direct exposure of ropivacaine hydrochloride to the corneal endothelium. Similar lesions in the corneal endothelium have been documented in other studies using local intraocular anaesthetics (EGGELING et al, 2000; TAN & BURTON 2000; KHABAK et al, 2006). In humans, the most commonly used anaesthetic within the eye is 1% lidocaine as it has less toxicity compared to other anaesthetics and concentrations (LIOU et al. 2004; BORAZAN et al, 2009; LEE et al, 2016).

CONCLUSION

According to the conditions proposed for this study, it was possible to conclude that ropivacaine 0.75% caused acute damage to the corneal endothelium of horses.

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BIOETHICS AND BIOSSECURITY COMMITTEE APPROVAL

This research was approved by the Research Committee of the xxx, and followed the ethical norms of the Association for Research in Vision and Ophthalmology (ARVO).

DECLARTION OF CONFLICT OF INTEREST

The authors declare no conflict of interest. The founding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

AUTHORS' CONTRIBUTIONS

The authors contributed equally to the manuscript.

REFERENCES

BORAZAN, M. et al. Induction of apoptosis of rabbit corneal endothelial cells by preservative-free lidocaine hydrochloride 2%, ropivacaine 1%, or levobupivacaine 0.75 **Journal of Cataract and Refractive Surgery**, v.35, n.4, p.753-758, 2009. Available from: https://doi.org/10.1016/j.jcrs.2008.12.016. Accessed: Jan. 20, 2017. doi: 10.1016/j.jcrs.2008.12.016.

BROOKS, D. E. et al. Visual outcomes of phacoemulsification cataract surgery in horses: 1990-2013. Veterinary Ophthalmology, v.17, n.1, p.117-128, 2014. Available from:

Ciência Rural, v.51, n.10, 2021.

5

<a>https://doi.org/10.1111/vop.12168>. Accessed: Sept. 21, 2015. doi: 10.1111/vop.12168. 2

4 ÇAÇA, I. et al. The histopathological effect of intracameral 5 ropivacaine in different concentrations on corneal endothelium. Annals of Ophthalmology, v.8, n.1, p.43-48, 2006. Available 6 7 from: <https://doi.org/10.1385/AO:38:1:43>. Accessed: Feb. 06, 2016. doi: 10.1385/AO:38:1:43. 8 C

10 EDELMANN, M. L. et al. Retrospective evaluation of phacoemulsification and aspiration in 41 horses (46 eyes): visual 11 12 outcomes vs. age, intraocular lens, and uveitis status. Veterinary Ophthalmology, v.17, n.1, p.160-167, 2014. Available from: 13 14 <https://doi.org/10.1111/vop.12185>. Accessed: Sept. 10, 2015. 15 doi: 10.1111/vop.12185. 16

17 EGGELING, P. et al. Corneal endothelial toxicity of different 18 lidocaine concentrations. Journal of Cataract and Refractive Surgery, v.26, n.9, p.1403-1408, 2000. Available from: https:// 19 doi.org/10.1016/S0886-3350(00)00379-5>. Accessed: Oct. 24, 20 21 2018. doi: 10.1016/S0886-3350(00)00379-5.

23 FAGANELLO, C. S. et al. Morphology of endothelial cells from 24 different regions of the equine cornea. Ciência Rural, v.45, n.12, p.2223-2228, 2016. Available from: https://doi.org/10.1590/0103- 25 8478cr20160216>. Accessed: Jun. 17, 2018. doi: 10.1590/0103-26 8478cr20160216. 27

29 GONNERING, R. et al. The pH tolerance of rabbit and human 30 corneal endothelium. Investigative Ophthalmology & Visual Science, v.18, p.373-390, 1979. Available from: https://pubmed. 31 32 ncbi.nlm.nih.gov/34576/>. Accessed: Oct. 24, 2018.

IACOBELLI, L. et al. Topical use of ropivacaine 1% vs lidocaine 34 35 2% in cataract surgery. Investigative Ophthalmology & Visual Science, v.46, n.13, p.795, 2005. Available from: https://iovs. 36 37 arvojournals.org/article.aspx?articleid=2400531>. Accessed: Sept. 38 10, 2015. 39

40 ISHIKAWA, A. Risk factors for reduced corneal endothelial 41 cell density before cataract surgery. Journal of Cataract and 42 Refractive Surgery, v.28, n.11, p.1982-1992, 2002. Available from: <https://doi.org/10.1016/S0886-3350(02)01502-X>. Accessed: Feb. 43 06, 2016. doi: 10.1016/S0886-3350(02)01502-X. 44 45

JIANG, G.; FAN T. Sodium ferulate attenuates lidocaine-46 47 induced corneal endothelial impairment. Oxidative Medicine 48 and Cellular Longevity, v.8, n.1, p.1-8, 2018. Available from: <a>https://doi.org/10.1155/2018/4967318>. Accessed: Oct. 24, 2018. 49 doi: 10.1155/2018/4967318. 50

52 KHAZBAK, L. et al. Effect of intracameral injection of lidocaine 53 and ropivacaine on rabbit corneal endothelium and trabecular 54 meshwork. Bulletin of the Ophthalmological Society of 55 Egypt, v.99, n.1, p.29-33, 2006. Available from: https://www. researchgate.net/publication/293654100. Accessed: Apr. 5, 2017. 56 57

KLAMT, J. G. et al. Continuous epidural anesthesia with 0.2% 58 59 ropivacaine associated to general anesthesia for upper abdominal 60 surgery in children. Revista Brasileira de Anestesiologia, v.53, 61 n.2, p.160-168, 2003. Available from: https://doi.org/10.1590/ S0034-70942003000200003>. Accessed: Jan. 20, 2017. doi: 62 10.1590/S0034-70942003000200003. 63 64

KUTHIALA, G.; CHAUDHARY, G. Ropivacaine: A review 65 66 of its pharmacology and clinical use. Indian Journal of Anaesthesia, v.55, n.2, p.104-110, 2011. Available from: https:// doi.org/10.4103/0019-5049.79875> Accessed: Aug. 29, 2016. doi: 10.4103/0019-5049.79875.

LEE, R. M. H. et al. Severe adverse events associated with local anaesthesia in cataract surgery: 1 year national survey of practice and complications in the UK. British Journal of Ophthalmology, v.100, n.6, p.772-776, 2016. Available from: http://dx.doi. org/10.1136/bjophthalmol-2015-307060>. Accessed: Jun. 17, 2018. doi: 10.1136/bjophthalmol-2015-307060.

LIOU, S. W. et al. Effect of intracameral injection of lidocaine and carbachol on the rabbit corneal endothelium. Journal of Cataract and Refractive Surgery, v.30, n.6, p.1351-1355, 2004. Available from: <https://doi.org/10.1016/j.jcrs.2003.10.032>. Accessed: Sept, 21, 2015. doi: 10.1016/j.jcrs.2003.10.032.

MARTINI, E. et al. Lidocaine versus ropivacaine for topical anesthesia in cataract surgery. Journal of Cataract and Refractive Surgery, v.28, n.6, p.1018-1022, 2002. Available from: https:// doi.org/10.1016/S0886-3350(01)01225-1>. Accessed: Jan. 20, 2017. doi: 10.1016/S0886-3350(01)01225-1.

NAUTSCHER, N. et al. Comparative morphological evaluation of domestic animal cornea. Veterinary Ophthalmology, v.19, n.4, p.297-304, 2015. Available from: https://doi.org/10.1111/ vop.12298>. Accessed: May, 08, 2016. doi: 10.1111/vop.12298.

OJEDA, J. L. et al. The three-dimentional microanatomy of the rabbit and human cornea. A chemical and mechanical microdissection-SEM approach. Journal of Anatomy, v.199, n.5, p.567-576, 2001. Available from: https://doi.org/10.1017/ S0021878201008512>. Accessed: Oct. 24, 2018. doi: 10.1017/ S0021878201008512.

OLMEZ, G. et al. Intraocular pressure and quality of blockade in peribulbar anesthesia using ropivacaine or lidocaine with adrenaline: a double-blind randomized study. The Tohoku Journal of Experimental Medicine, v.204, n.3, p.203-208, 2004. Available from: <https://doi.org/10.1620/tjem.204.203>. Accessed: Mar. 21, 2015. doi: 10.1620/tjem.204.203.

PARIKH, C. H.; EDELHAUSER, H.F. Ocular surgical pharmacology: corneal endothelial safety and toxicity. Current Opinion in Ophthalmology, v.14, n.4, p.178-185, 2003. Available from: <https://pubmed.ncbi.nlm.nih.gov/12888714/>. Accessed: May, 08, 2016. doi: 10.1097/00055735-200308000-00002.

PARK, S. A. et al. Evaluation of the analgesic effect of intracameral lidocaine hydrochloride injection on intraoperative and postoperative pain in healthy dogs undergoing phacoemulsification. American Journal of Veterinary Research, v.71, n.2, p.216-222, 2010. Available from: https://doi.org/10.2460/ajvr.71.2.216>. Accessed: Apr. 18, 2018. doi: 10.2460/ajvr.71.2.216.

PIGATTO, J. A. T. et al. Morphometric analysis of the corneal endothelium of rabbits using scanning electron microscopy, Acta Scientiae Veterinariae, v.33, n.1, p.41-45, 2005. Available from: <https://doi.org/10.22456/1679-9216.14441>. Accessed: May, 03, 2018. doi: 10.22456/1679-9216.14441.

PIGATTO, J. A. T. et al. Scanning electron microscopy of the corneal endothelium of ostrich. Ciência Rural, v.39, n.3, p.926-929, 2009. Available from: https://doi.org/10.1590/S0103- 84782009005000001>. Accessed: Nov. 11, 2017. doi: 10.1590/ S0103-84782009005000001.

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3 2000. Available from: https://bjan-sba.org/article/5e498c3d0aec5

4 119028b49e7/pdf/rba-50-6-442.pdf>. Accessed: Jul. 27, 2017.

5
6 SEGARRA, S. et al. A dose-escalation ex vivo study on the
7 effects of intracameral benzalkonium chloride in rabbits. BMC
8 Veterinary Research, v.14, n.39, p.1-9, 2018. Available from:
9 https://doi.org/10.1186/s12917-018-1349-8. Accessed: Sept. 03,
10 2018. doi: 10.1186/s12917-018-1349-8.

 SILVA, V. R. M. et al. Evaluation of equine corneal endothelium after exposure to 0.5% indocyanine green - in vitro study. Semina:
 Ciências Agrárias, v.39, n.2, p.613-620, 2018. Available from:
 https://doi.org/10.5433/1679-0359.2018v39n2p613. Accessed:
 Dec. 03, 2018. doi: 10.5433/1679-0359.2018v39n2p613.

17

18 SIT, M. et al. Corneal graft outcome study. Cornea,
19 v.20, n.2, p.129-133, 2001. Available from: https://doi.org/10.1097/00003226-200103000-00002. Accessed: Jul. 27,
2017. doi: 10.1097/00003226-200103000-00002.

TAN, J. H. Y., BURTON, R.L. Does preservative-free lignocaine
1% for hydrossection reduce pain during phacoemulsification?
Journal of Cataract and Refractive Surgery, v.26, n.5, p.733735, 2000. Available from: https://doi.org/10.1016/S0886-3350(00)00311-4> Accessed: Sept. 26, 2016. doi: 10.1016/S0886-

TERZARIOL, M. et al. Effects of intracameral brilliant blue on the corneal endothelium of swine: in vitro study. **Pesquisa Veterinária Brasileira**, v.36, n.8, p.775-780, 2016. Available from: http://dx.doi.org/10.1590/S0100-736X2016000800016. Accessed: Nov. 11, 2017. doi: 10.1590/S0100-736X2016000800016.

TOWNSEND, W. M. Disease and surgery of the equine lens. **Veterinary Clinics of North America:** Equine Practice, v.33, n.3, p.483-497, 2017. Available from: http://dx.doi.org/10.1016/j.cveq.2017.07.004. Accessed: Apr. 18, 2018. doi: 10.1016/j.cveq.2017.07.004.

VÄLIMÄKI, J., TÖRNBLOM, R. Viscoanaesthesia in cataract surgery: a prospective, randomized clinical trial. Acta **Ophthalmologica**, v.87, n.4, p.378-381, 2009. Available from: https://doi.org/10.1111/j.1755-3768.2008.01267.x. Accessed: Jul. 27, 2017. doi: 10.1111/j.1755-3768.2008.01267.x.

WANG, L. et al. Combined topical-intracameral anesthesia in manual small-incision cataract surgery: A Prospective, Randomized, Double-Masked, Placebo-Controlled Trial. **Asia-Pacific Journal of Ophthalmology**, v.1, p.9-14, 2013. Available from: https://journals.lww.com/apjoo/fulltext/2013/. Accessed: Sept. 26, 2016. doi: 10.1097/APO.0b013e318274c335.

WEN, Q. et al. Cytotoxicity of proparacaine to human corneal endothelial cells in vitro. **The Journal of Toxicological Sciences**, v.40, n.4, p.427-436, 2015. Available from: https://doi.org/10.2131/jts.40.427. Accessed: Apr. 18, 2018. doi: 10.2131/jts.40.427.

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