

UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL
FACULDADE DE MEDICINA
PROGRAMA DE PÓS-GRADUAÇÃO EM PSIQUIATRIA E CIÊNCIAS DO
COMPORTAMENTO

ELLEN SCOTTON

**O PAPEL DO BDNF E DA PRDX-1 NA PROTEÇÃO CONTRA O DANO
OXIDATIVO CENTRAL EM RATOS ANEDÔNICOS SUBMETIDOS A
UM PROTOCOLO DE ESTRESSE CRÔNICO MODERADO**

Porto Alegre, 2019

ELLEN SCOTTON

**O PAPEL DO BDNF E DA PRDX-1 NA PROTEÇÃO CONTRA O DANO
OXIDATIVO CENTRAL EM RATOS ANEDÔNICOS SUBMETIDOS A UM
PROTOCOLO DE ESTRESSE CRÔNICO MODERADO**

Dissertação apresentada como requisito parcial
para obtenção de título de Mestre em
Psiquiatria e Ciências do Comportamento à
Universidade Federal do Rio Grande do Sul,
Programa de Pós-Graduação em Psiquiatria e
Ciências do Comportamento.

Orientador: Prof. Dr. Maurício Kunz

Porto Alegre, 2019

CIP - Catalogação na Publicação

Scotton, Ellen

O PAPEL DO BDNF E DA PRDX-1 NA PROTEÇÃO CONTRA O
DANO OXIDATIVO CENTRAL EM RATOS ANEDÔNICOS SUBMETIDOS
A UM PROTOCOLO DE ESTRESSE CRÔNICO MODERADO / Ellen
Scotton. -- 2019.

114 f.

Orientador: Maurício Kunz.

Dissertação (Mestrado) -- Universidade Federal do
Rio Grande do Sul, Faculdade de Medicina, Programa de
Pós-Graduação em Psiquiatria e Ciências do
Comportamento, Porto Alegre, BR-RS, 2019.

1. Depressão. 2. Anedonia. 3. Ratos Wistar. 4.
Estresse oxidativo. 5. Fator neurotrófico derivado do
encéfalo. I. Kunz, Maurício, orient. II. Título.

FOLHA DE APROVAÇÃO DA BANCA EXAMINADORA

ELLEN SCOTTON

O PAPEL DO BDNF E DA PRDX-1 NA PROTEÇÃO CONTRA O DANO OXIDATIVO CENTRAL EM RATOS ANEDÔNICOS SUBMETIDOS A UM PROTOCOLO DE ESTRESSE CRÔNICO MODERADO

Dissertação apresentada como requisito parcial para obtenção de título de Mestre em Psiquiatria e Ciências do Comportamento à Universidade Federal do Rio Grande do Sul, Programa de Pós-Graduação em Psiquiatria e Ciências do Comportamento.

Porto Alegre, 29 de Março de 2019.

A comissão Examinadora, abaixo assinada, aprova a Dissertação “O papel do BDNF e da PRDX-1 na proteção contra o dano oxidativo central em ratos anedônicos submetidos a um protocolo de estresse crônico moderado.”, elaborada por Ellen Scotton como requisito parcial para a obtenção do grau de Mestre em Psiquiatria e Ciências do Comportamento.

Prof^a. Dr^a. Márcia Kauer Sant’Anna (UFRGS)

Dr. Dirson João Stein (UFRGS)

Prof^a. Dr^a. Rosa Maria Martins de Almeida (UFRGS)

AGRADECIMENTOS

Ao meu orientador, Maurício Kunz, pelos ensinamentos e incentivo durante a minha formação acadêmica.

Ao Rafa, meu irmão de coração, pela oportunidade de ter desenvolvido esse trabalho em parceria e por todo o auxílio durante essa etapa. Obrigada pela confiança, pela amizade sincera, pelo estímulo e pela presença constante.

À Adri, pelos momentos super produtivos de discussão e por sempre me incentivar.

À Giginha, pela ajuda, pelo companheirismo e pela amizade que vai muito além desse trabalho. Obrigada por ser um exemplo tão lindo na pesquisa e na vida.

A todos os amigos e colegas do Laboratório de Psiquiatria Molecular: Jéssica, Gabriela, Gabriel, Larissa, Bárbara, Érica, Fernanda, Luiza, Mailton, Daniel, Diego, Jacson, Gabi Berni e Letícia. Obrigada por toda a ajuda prática e pelo apoio emocional.

À toda equipe da UEA, em especial à Marta e à Dani, que sempre estiveram disponíveis para auxiliar durante o experimento com os animais.

Aos colaboradores externos, Tiago, Mirian, Alessandra e Tuani. Obrigada por terem adicionados suas expertises a esse trabalho.

À Gabix, minha amiga da vida toda, que esteve do meu lado em todos os momentos, mesmo quando longe. Obrigada pelo apoio, pelo incentivo, pelos momentos de descontração e por compreender tantas vezes a minha ausência.

Às amadas Bi Pfaffen e Bruna Maria, pela amizade e companheirismo que iniciaram no lab e seguirão por toda a vida. Vocês são uma grande inspiração, em todos os âmbitos.

Ao Ryan, por ter escolhido estar ao meu lado, principalmente na fase final dessa etapa. Obrigada por todo o amor, companheirismo, compreensão e por me fazer acreditar que no final, vai dar tudo certo, sempre.

À minha família... Mãe, Pai, Érica, Edna, Baccin e Dirceo, pelo suporte e pelo apoio constantes. Eu amo vocês, obrigada por tudo!

SUMÁRIO

LISTA DE ABREVIATURAS.....	7
APRESENTAÇÃO.....	10
PARTE I	11
RESUMO	12
ABSTRACT	13
1 INTRODUÇÃO.....	14
1.1 Depressão Maior	14
1.1.1 Neurobiologia	15
1.1.2 Hipótese inflamatória.....	16
1.1.2.1 Inflamação e dessensibilização do eixo HPA	19
1.1.3 Estresse oxidativo	21
1.1.3.1 Estresse oxidativo, inflamação e BDNF	24
1.2 Resiliência e suscetibilidade ao estresse	27
1.3 CUMS	28
1.4 Proteômica e biologia de sistemas na DM	30
2 JUSTIFICATIVA	32
3 OBJETIVOS	34
3.1 Objetivo geral	34
3.2 Objetivos específicos	34
PARTE II	35
4 ARTIGO	36
4.1 Carta de submissão	36
4.2 Manuscrito	37
PARTE III	72
5 CONSIDERAÇÕES FINAIS	73
6 PERSPECTIVAS	75
7 REFERÊNCIAS	76
ANEXO A	94
<i>Carta de aprovação do projeto sob número 150353 pela Comissão de Ética no Uso de Animais do Hospital de Clínicas de Porto Alegre (CEUA/HCPA).</i>	94
ANEXO B	95

LISTA DE ABREVIATURAS

ACTH	Corticotrofina (do inglês <i>adrenocorticotrophic hormone</i>)
BDNF	Fator neurotrófico derivado do cérebro (do inglês <i>brain-derived neurotrophic factor</i>)
BH4	Tetrahidrobiopterina
CAT	Catalase
CPF	Córtex pré-frontal
CRH	Hormônio liberador de corticotrofina (do inglês <i>corticotropin-releasing hormone</i>)
CUMS	Estresse crônico moderado e imprevisível (do inglês <i>chronic unpredictable mild stress</i>)
DM	Depressão Maior
DRT	Depressão resistente ao tratamento
DSM	Manual Diagnóstico e Estatístico de Transtornos Mentais (do inglês <i>Diagnostic and Statistical Manual of Mental Disorders</i>)
EROs	Espécies reativas de oxigênio
GCS	Glicocorticóides
GPx	Glutationa peroxidase
GR	Glutationa redutase
GSH	Glutationa reduzida
GSSG	Glutationa oxidada
H ₂ O ₂	Peróxido de hidrogênio
HPA	Hipotálamo-pituitária-adrenal
IDO	Indoleamina 2,3-dioxigenase

IFN	Interferon
IL	Interleucina
iNOS	Óxido nítrico sintase induzível (do inglês <i>inducible nitric oxide synthase</i>)
IRNDs	Inibidores da recaptação de noradrenalina e dopamina
IRSNs	Inibidores da recaptação de serotonina e noradrenalina
ISRSs	Inibidores seletivos da recaptação de serotonina
LPS	Lipopolissacarídeo
NF-κB	Fator nuclear Kappa B (do inglês <i>nuclear factor kappa B</i>)
NMDA	N-metil-d-aspartato
Nrf2	Fator nuclear eritróide 2 relacionado ao fator 2 (do inglês <i>nuclear factor erythroid 2-related factor 2</i>)
MAPK	Proteínas quinases ativadas por mitógeno (do inglês <i>mitogen-activated protein kinases</i>)
MDA	Malondialdeído
OMS	Organização Mundial da Saúde
PCR	Proteína C reativa
PRDX	Peroxirredoxina
RGs	Receptores de glicocorticoides
SNC	Sistema nervoso central
SOD	Superóxido dismutase
TAC	Capacidade antioxidante total (do inglês <i>total antioxidant capacity</i>)
TNF	Fator de necrose tumoral (do inglês <i>tumor necrosis factor</i>)

TOS Estado oxidante total (do inglês *total oxidant status*)

TrkB Receptor tropomiosina quinase B (do inglês *tropomyosin receptor kinase B*)

APRESENTAÇÃO

Este trabalho consiste na dissertação de mestrado intitulada “O papel do BDNF e da PRDX-1 na proteção contra o dano oxidativo central em ratos anedônicos submetidos a um protocolo de estresse crônico moderado”, apresentada ao Programa de Pós-Graduação em Psiquiatria e Ciências do Comportamento da Universidade Federal do Rio Grande do Sul, em 29 de março de 2019. O trabalho é apresentado em três partes, conforme segue:

- **Parte I:** Resumo, *Abstract*, Introdução, Justificativa e Objetivos.
- **Parte II:** Metodologia, Resultados e Discussão apresentados no formato do artigo científico intitulado: “*The protective role of BDNF and PRDX-1 against central oxidative damage in anhedonic rats submitted to a chronic unpredictable mild stress protocol*”. Os resultados discutidos são provenientes de duas frentes de investigação:
 - a) Estudo experimental: avaliamos o comportamento, peso corporal, peso relativo das glândulas adrenais, bem como medidas centrais do fator neurotrófico derivado do cérebro (BDNF) e parâmetros oxidativos em animais submetidos a um protocolo de estresse crônico moderado e imprevisível.
 - b) Bioinformática: realizamos uma análise de bancos de dados de proteômica de três estudos que realizaram protocolos experimentais semelhantes ao do presente trabalho, a fim de identificar proteínas diferencialmente expressas em processos biológicos envolvendo estresse oxidativo e inflamação em animais anedônicos.
- **Parte III:** Considerações Finais e Perspectivas.

Além disso, na sequência constam as Referências e na seção de Anexos encontram-se a carta de aprovação do projeto pela Comissão de Ética no Uso de Animais (CEUA/HCPA), e as instruções para a submissão de manuscritos na revista *Brain, Behavior and Immunity*.

PARTE I

RESUMO

A ativação crônica do eixo hipotálamo-pituitária-adrenal (HPA) e o aumento sustentado de glicocorticoides têm sido associados à depressão maior (DM), e também a alterações envolvendo neurotrofinas e marcadores de estresse oxidativo em resposta à inflamação. O presente estudo teve como objetivo avaliar medidas centrais do fator neurotrófico derivado do cérebro (BDNF), de dano oxidativo, e da capacidade antioxidante total de ratos submetidos ao estresse crônico moderado e imprevisível (CUMS), além de investigar a relação entre os níveis de BDNF e proteínas diferencialmente expressas envolvidas em processos biológicos de estresse oxidativo e inflamação. Ratos Wistar machos foram submetidos ao CUMS por seis semanas. Conforme as taxas de preferência por sacarose, os animais foram classificados como anedônicos ou não-anedônicos. Após a coleta dos tecidos cerebrais, foram avaliados o dano oxidativo, a capacidade antioxidante total e os níveis de BDNF. A fim de estender a discussão sobre possíveis mecanismos envolvendo os achados experimentais, adicionalmente, foi realizada uma análise de bioinformática reunindo resultados de proteômica de outros estudos com protocolo experimental semelhante, visando identificar proteínas envolvidas com o estresse oxidativo e vias inflamatórias diferencialmente expressas em animais anedônicos. O CUMS esteve associado ao aumento das concentrações de BDNF e à diminuição da capacidade antioxidante total, além de não ter sido identificado dano oxidativo aos lipídios e proteínas nos animais estressados. Além disso, a abordagem proteômica de bioinformática indicou que animais anedônicos apresentam um aumento na expressão de peroxirredoxina-1 (PRDX-1) e uma diminuição na expressão de proteínas envolvidas com sinalização apoptótica e inflamatória (RELA, ASK-1 e TAK-1) no hipocampo. Essas evidências sugerem que o BDNF e a PRDX-1 podem representar uma resposta inicial contra o estresse, com papel compensatório, prevenindo o dano oxidativo a lipídios e proteínas através da modulação da defesa antioxidante, principalmente em animais anedônicos.

Palavras-chave: Depressão maior; comportamento anedônico; estresse crônico moderado e imprevisível; preferência por sacarose; BDNF; PRDX-1; estresse oxidativo; espécies reativas de oxigênio; capacidade antioxidante total; proteômica.

ABSTRACT

Chronic activation of the HPA axis and sustained increase of glucocorticoids have been associated in major depression (MD), and also related to changes involving neurotrophins and oxidative stress markers in response to inflammation. This study aimed to evaluate central measures of brain-derived neurotrophic factor (BDNF), oxidative damage and total antioxidant capacity of rats submitted to chronic unpredictable mild stress (CUMS) and investigate the relationship between BDNF levels and differentially expressed proteins involved in oxidative stress and inflammatory biological processes. Wistar male rats were subjected to CUMS for six weeks. Based on the sucrose preference test, animals were divided into anhedonic and non-anhedonic clusters. After brain tissue collection, oxidative damage, total antioxidant capacity, and BDNF levels were evaluated. In order to extend discussion of possible mechanisms involving neurobiological findings, a bioinformatics approach was performed to identify proteins involved with oxidative stress and inflammation pathways that were differentially expressed in anhedonic animals from other studies with similar experimental protocol. CUMS was associated with an increase in BDNF concentrations accompanied by a decrease in total antioxidant capacity, besides the absence of oxidative damage to lipids and proteins in stressed animals. Withal, bioinformatics proteomic approach indicated that anhedonic animals showed a peroxiredoxin-1 (PRDX-1) up-regulation and a down-regulation of proteins involved with apoptotic and inflammation signaling (RELA, ASK-1 and TAK-1) in the hippocampus. These evidences suggest that BDNF and PRDX-1 may represent an initial response against stress, with a compensatory role, preventing oxidative damage to lipids and proteins through the modulation of antioxidant defense, mainly in anhedonic animals.

Keywords: Major depressive disorder; anhedonic behavior; chronic unpredictable mild stress; sucrose preference; BDNF; PRDX-1; oxidative stress; reactive oxygen species; total antioxidant capacity; proteomics

1 INTRODUÇÃO

1.1 Depressão Maior

A Depressão Maior (DM) é um transtorno psiquiátrico grave, crônico, altamente incapacitante e frequentemente associado a outras comorbidades, além de apresentar risco de suicídio substancialmente elevado (1–3). As manifestações da doença acarretam no prejuízo do funcionamento e na diminuição da qualidade de vida do indivíduo acometido, bem como elevam de forma importante os gastos no âmbito da saúde pública (4,5).

O diagnóstico da DM é realizado de acordo com critérios descritos no Manual Diagnóstico e Estatístico de Transtornos Mentais (DSM-V), e a sintomatologia varia de acordo com o endofenótipo apresentado. De forma geral, as manifestações incluem humor deprimido ou irritação, alterações do sono e do apetite, agitação ou retardo psicomotor, fadiga, sentimento de inutilidade ou culpa excessiva, falta de concentração, pensamento recorrente de morte e/ou ideação suicida (6–8). A anedonia, por sua vez, é um sintoma caracterizado pela ausência quase completa de sensação de prazer, motivação e interesse, sendo uma manifestação central no endofenótipo melancólico (9).

A prevalência da DM é estimada em 4,4%, sendo uma doença que se manifesta usualmente no início da idade adulta (10–13). Segundo dados recentes da Organização Mundial da Saúde (OMS), a DM é considerada a principal causa de incapacidade na população em geral, assumindo o quarto lugar entre as doenças com maior número de pessoas acometidas no mundo. Até o ano de 2020, a estimativa é de que a DM esteja em segundo lugar dentre as doenças com maior prevalência mundial (13).

Considerando as terapias farmacológicas disponíveis atualmente, apenas metade dos pacientes atinge a remissão completa dos sintomas, mesmo após várias tentativas de tratamento. Além disso, a remissão está vinculada a altos índices de recaída, principalmente durante o primeiro ano (14). Apesar dos antidepressivos serem responsáveis por resultados

importantes em parte dos indivíduos com DM, o tratamento convencional apresenta muitos efeitos adversos indesejáveis, como boca seca, tontura, ganho de peso e disfunção sexual, o que resulta na baixa adesão dos pacientes (2). Ainda, cerca de 20% dos indivíduos acometidos seguem sintomáticos após dois anos de tratamento (15).

Estima-se que 44% dos pacientes não respondem a duas tentativas consecutivas de tratamento com antidepressivos, apresentando depressão resistente ao tratamento (DRT). A elevada prevalência, somada à diminuição ainda mais acentuada da qualidade de vida, à baixa produtividade, ao maior número de hospitalizações, bem como seu alto impacto econômico, fazem da DRT um dos maiores desafios da psiquiatria moderna (16–19). Recentemente, Bergfeld e colaboradores reportaram que indivíduos com DRT apresentaram índices de tentativa e execução de suicídio, respectivamente, duas e dez vezes maiores, do que indivíduos depressivos não resistentes (20,21).

1.1.1 Neurobiologia

Alterações estruturais e funcionais em diversas regiões corticais envolvidas com o processamento de emoções, controle cognitivo, aprendizagem, formação de memórias e funções executivas estão presentes na DM (22,23). Estudos *post-mortem* com indivíduos depressivos reportam alterações em estruturas associadas fortemente com sinais clássicos da DM, como o córtex pré-frontal (CPF) e hipocampo (24–26). Além disso, estudos de imagem demonstram uma diminuição no volume dessas estruturas em pacientes depressivos, o que reforça a ideia de uma diminuição no número de neurônios e de células gliais em regiões córtico-límbicas de indivíduos com DM (27–31).

Desde os anos 50, a primeira linha de tratamento antidepressivo é composta por medicamentos que atuam sobre a recaptação de monoaminas, como os inibidores seletivos da recaptação de serotonina (ISRSs), inibidores da recaptação de serotonina e noradrenalina (IRSNs) e inibidores da recaptação de noraepinefrina e dopamina (IRNDs) (32). A utilização

desses fármacos é baseada na teoria monoaminérgica da DM, a qual propõe que as manifestações clínicas da doença são provenientes de uma redução na disponibilidade desses neurotransmissores ou de alterações funcionais da transmissão monoaminérgica de regiões cerebrais específicas (33,34).

A alteração no sistema serotoninérgico, por sua vez, pode ser considerada um mediador central na manifestação dos sintomas depressivos, uma vez que esse sistema tem origem nos núcleos dorsal e medial da rafe, e se projeta para diversas áreas corticais e subcorticais, tais como CPF, hipocampo, amígdala e estriado. Desse modo, a neurotransmissão serotoninérgica influencia os sistemas dopamínérigo, noradrenérgico, glutamatérgico, colinérgico e GABAérgico, tendo assim a capacidade de regular o humor, o sono e o apetite, funções que se encontram alteradas na DM (35).

Nesse contexto, alterações na biossíntese da serotonina também parecem estar envolvidas na fisiopatologia da DM, como postulado por Lapin e Oxenkrug em 1969, com a criação da “hipótese serotoninérgica”. Essa teoria propõe que alterações no metabolismo do triptofano, precursor da serotonina, resultem na diminuição da síntese desse mediador químico, levando à ocorrência de sintomas depressivos (36). Sugere-se ainda que o prejuízo na síntese de serotonina seja mediado por fatores ambientais, como a exposição crônica ao estresse, e pela modulação da resposta inflamatória via ativação do eixo HPA.

1.1.2 Hipótese inflamatória

Como descrito previamente, um alto índice de pacientes deprimidos tratados com os antidepressivos convencionais não apresenta remissão completa dos sintomas ou demonstra ser refratário ao tratamento (37,38). Evidências crescentes também indicam que a DM está associada a comorbidades de cunho inflamatório, como *diabetes mellitus* (39), obesidade (40) e doenças reumáticas (41), além de estar relacionada a outros fatores que aumentam a produção de mediadores inflamatórios, como o estresse psicossocial, diminuição no sono,

abuso sexual na infância e isolamento social (42,43). Nesse sentido, a DM parece apresentar um importante componente inflamatório em suas bases etiológicas (44,45).

Diversos estudos demonstram que os níveis periféricos de proteínas envolvidas com a resposta inflamatória encontram-se alterados em pacientes com DM. Uma meta-análise de Dowlati e colaboradores indicou aumento significativo nos níveis circulantes do fator de necrose tumoral alfa (TNF- α) e de interleucina (IL) 1 beta (IL-1 β) (46), enquanto outra meta-análise demonstrou aumento plasmático de proteína C reativa (PCR), IL-1 β e IL-6, ambos em pacientes depressivos (47). Ainda, um estudo epidemiológico demonstrou que níveis elevados de PCR e IL-6 detectados no início do estudo foram associados à presença de sintomas da DM após doze anos de seguimento (48).

Além das citocinas estarem aumentadas perifericamente, esses marcadores inflamatórios podem atingir o sistema nervoso central (SNC) atravessando a barreira hematoencefálica através dos órgãos circunventriculares ou pela via aferente do nervo vago (49–51). No SNC as citocinas podem ser produzidas pela microglia, astrócitos e neurônios e atuam como transmissores gliais, podendo modificar funções neuronais através de sua ligação com um amplo número de receptores (44).

As células da microglia são macrófagos residentes no SNC, os quais atuam na regulação da resposta neuroimune central. Em condições fisiológicas, a microglia se apresenta como uma rede amplamente distribuída que estabelece contato com dendritos e sinapses próximas, podendo remodelar essas conexões e secretar neurotrofinas como o BDNF, o qual exerce papel neuroprotetor, preservando as funções desse microambiente (52). Frente a um sinal gerado por dano celular, a microglia altera sua morfologia passando a secretar citocinas e quimiocinas que recrutam outras células do sistema imune, a fim de promover o reparo e remodelamento tecidual (53).

Tipicamente, a ativação da microglia com fenótipo tipo "M2" está associada a um perfil anti-inflamatório, representado pelo aumento de BDNF, da IL-4, e da atividade fagocítica. Em contraste, a ativação do fenótipo "M1" está associada à maior expressão da enzima óxido nítrico sintase induzível (iNOS), produção de espécies reativas de oxigênio (EROs) e de IL-1 β , além da diminuição da secreção de fatores neurotróficos, configurando um perfil pró-inflamatório. Embora a ativação da microglia não possa ser dicotomizada entre ativação tipo "M1" e ativação tipo "M2", sugere-se que a intensidade dessas respostas antagônicas possa determinar se a atividade microglial resultará na depuração de detritos teciduais e na resolução da resposta inflamatória, ou se será estabelecido um processo de neuroinflamação crônica, levando a disfunção e morte celular (52).

As citocinas pró-inflamatórias tais como interferon (IFN), IL-1 β e TNF- α podem reduzir a disponibilidade de monoaminas como serotonina, dopamina e noradrenalina através do aumento da expressão e função dos seus transportadores pré-sinápticos. Ainda, o aumento de citocinas pode resultar na geração de EROs, provocando a oxidação de um cofator enzimático essencial para a síntese de dopamina, a tetrahidrobiopterina (BH4) (54,55). Esses processos podem provocar um prejuízo na via de recompensa, estando associados com a manifestação da anedonia (56).

O aumento da resposta inflamatória também pode diminuir a disponibilidade de monoaminas, desviando o triptofano, precursor da serotonina, para a via das quinureninas, através da ativação da enzima indolamina 2,3-dioxigenase (IDO). A microglia ativada pode converter quinurenina em ácido quinolínico, um agonista do receptor N-metil-d-aspartato (NMDA), o que juntamente com a redução na recaptAÇÃO e maior liberação astrocitária de glutamato pode resultar na amplificação da neurotransmissão glutamatérgica, além de diminuir a concentração de BDNF e interferir na sinalização de seu receptor tropomiosina quinase B (TrkB), afetando os processos de neurogênese e plasticidade neuronal. Esses

desfechos possivelmente estão relacionados com a piora cognitiva dos pacientes com DM (54,55,57,58).

Quando a via das quinureninas é estimulada ocorre a elevação da condutância da membrana ao cálcio, o aumento da lipoperoxidação e da produção de mediadores inflamatórios, o estímulo da iNOS e da apoptose neuronal, além do surgimento de sinais clínicos da DM (57,58). Nesse sentido, além de diminuir a disponibilidade de triptofano, o aumento da atividade da IDO em resposta à inflamação resulta na amplificação da neurotransmissão glutamatérgica, no aumento de estresse oxidativo e intensifica o processo de neurodegeneração (59). Em modelos pré-clínicos o bloqueio dos receptores NMDA através da utilização de quetamina, ou a utilização de inibidores da IDO, diminuiu o comportamento tipo-depressivo em camundongos (60,61).

Em suma, uma vez no cérebro as citocinas podem influenciar os sistemas de neurotransmissão, induzir apoptose, diminuir a neurogênese e prejudicar os mecanismos de neuroplasticidade, podendo afetar circuitos que regulam o comportamento e estão relacionados com a manifestação de sintomas clássicos da DM, como a anedonia e a ansiedade (54,55,62).

1.1.2.1 Inflamação e dessensibilização do eixo HPA

O estresse é considerado um estado de desequilíbrio homeostático frente à exposição a estressores psicológicos, ambientais ou fisiológicos. Esses eventos geram múltiplas alterações neuroquímicas, imunes e hormonais, levando a ativação de uma resposta mediada principalmente pelo eixo HPA (31). A partir de um estímulo estressor, ocorre a liberação do hormônio liberador de corticotrofina (CRH) pelo hipotálamo com a consecutiva síntese e liberação da corticotrofina (ACTH) pela adenóhipófise, o qual atua sobre a glândula adrenal, estimulando a síntese de glicocorticoides (GCs), principalmente o cortisol, a espécie humana (63).

Em uma resposta aguda ao estresse os GCs apresentam propriedades imunossupressoras e anti-inflamatórias, inibem a proliferação de linfócitos, reduzem a expressão de citocinas pró-inflamatórias como TNF- α e IL-6 e estimulam o aumento da expressão de citocinas anti-inflamatórias, como a IL-10 (64). Além disso, os receptores de GCs (RGs) – principalmente os receptores de GCs α (RG α) – respondem promovendo um *feedback* negativo sobre a produção de CRH, a fim de preservar a homeostase do organismo. O estresse crônico, por sua vez, aumenta de forma significativa os níveis periféricos de IL-6, o que acaba por intensificar a liberação de cortisol e diminuir a sensibilidade do eixo HPA (42,65).

Em geral, respostas sustentadas ao estresse envolvem concentrações persistentemente altas de GCs, além de níveis aumentados de adrenalina e noradrenalina, o que acaba por prejudicar o funcionamento do eixo HPA, estimulando a inflamação (66). A ativação prolongada do eixo HPA e aumento crônico de GCs têm sido evidenciados na DM, bem como a diminuição da sensibilidade do eixo HPA desses indivíduos frente ao tratamento com dexametasona, um corticoide com potente ação anti-inflamatória (67,68). Essa dessensibilização do eixo HPA parece ser intermediada pela resistência aos GCs, uma vez que na DM e em situações de estresse crônico ocorre a diminuição da expressão dos RGs no hipotálamo, prejudicando o *feedback* negativo necessário para desativar o eixo HPA (65,69). Ainda, o prejuízo no *feedback* negativo potencializa a sinalização imune, através do aumento dos níveis de citocinas e de células pró-inflamatórias (70,71).

Da mesma forma que a resistência aos GCs potencializa a inflamação, as citocinas pró-inflamatórias atuam sobre os RGs, diminuindo sua expressão e função, agravando ainda mais o cenário inflamatório (72). Achados de Carvalho e colaboradores evidenciam o aumento de cortisol e de IL-6 em sangue periférico de pacientes com DRT (73). Os mesmos indivíduos também apresentaram indícios de resistência aos GCs em um ensaio *in vitro*

envolvendo a exposição de suas células linfomononucleares a um insulto e posterior tratamento com cortisol e dexametasona (74). Em conjunto, os achados envolvendo a resistência aos GCs, o aumento na produção de cortisol, bem como a resposta inflamatória, demonstram que esses mecanismos são coexistentes em muitos pacientes depressivos, evidenciando a relação entre a inflamação e a resposta ao estresse na fisiopatologia da DM.

1.1.3 Estresse oxidativo

Durante o metabolismo celular energético aproximadamente 5% do oxigênio não é reduzido diretamente a água, gerando moléculas altamente reativas conhecidas como EROS. As principais EROS incluem o ânion superóxido, o radical hidroxila e o peróxido de hidrogênio (H_2O_2) (75,76). Em baixas concentrações as EROS desempenham funções importantes na regulação de processos como a fagocitose, apoptose, ativação de fatores de transcrição e sinalização celular, estando em equilíbrio com os sistemas antioxidantes (77). Condições adversas podem gerar um desequilíbrio no estado redox celular, tanto pela produção aumentada de EROS, como pela diminuição na capacidade de defesa antioxidante, levando ao estresse oxidativo (76). Essa situação pode provocar dano em biomoléculas como lipídios, proteínas e DNA. O cérebro, por sua vez, é um órgão altamente suscetível aos efeitos prejudiciais das EROS, devido à sua alta demanda metabólica e aos menores níveis de antioxidantes (78).

A carbonilação de proteínas é um marcador muito associado a dano irreversível promovido pelo estresse oxidativo. Esse processo pode resultar no prejuízo das atividades transportadoras e receptoras das proteínas, levando à perda de sua funcionalidade e também à oxidação de lipídios (79,80). Estruturalmente os ácidos graxos são um dos componentes principais das membranas celulares, sendo importantes na manutenção de sua integridade e fluidez. A peroxidação lipídica é um processo que age sobre a composição e organização celular, promovendo alterações covalentes de biomoléculas e prejudicando a homeostasia do

organismo. Adicionalmente, a peroxidação lipídica tem um envolvimento bem documentado nos processos de morte celular e em várias doenças (81). Diversos estudos já identificaram níveis séricos aumentados de malondialdeído (MDA), um subproduto reativo da lipoperoxidação, em pacientes com DM (82–84). Uma meta-análise recente mostrou que compostos associados com a peroxidação lipídica e com a oxidação do DNA, 8-hidroxi-2-deoxiguanosina e F2-isoprostanos, respectivamente, são os marcadores de estresse oxidativo mais consistentemente associados com a DM (85).

O sistema de defesa enzimático é composto por muitas enzimas. Dentre elas, destacam-se a superóxido dismutase (SOD), a catalase (CAT) e a glutationa peroxidase (GPx). As suas atividades representam a primeira linha de proteção antioxidante, tendo um papel fundamental nos mecanismos e estratégias de defesa do organismo (86). Em um cenário de estresse oxidativo, a SOD é a primeira enzima antioxidante responsável por proteger as células dos danos ocasionados pelas EROs, seguida da ação da CAT (87,88). Alterações na atividade da SOD são usualmente encontradas em pacientes deprimidos, mas os achados são inconsistentes quanto à direção dessas alterações (89). Algumas evidências apontam para uma diminuição da atividade da SOD (90,91), bem como da CAT (92) na DM. Em contrapartida, o aumento das atividades de ambas as enzimas também tem sido demonstrado em outros estudos (93,94). Recentemente, Tsai e Huang encontraram níveis séricos de SOD e CAT aumentados durante a fase aguda da DM (95).

As peroxirredoxinas (PRDX), por sua vez, são proteínas antioxidantes que agem em conjunto com outros sistemas de defesa diminuindo os substratos utilizados na formação do H₂O₂ e modulando a sua sinalização (96–98). Kim e colaboradores demonstraram que a PRDX-1 estava *up-regulated* na microglia, após exposição ao lipopolissacarídeo (LPS). A reversão desse aumento esteve associada à morte celular mediada por H₂O₂, sugerindo um papel protetor da PRDX-1 frente a um contexto pró-inflamatório (99). Ainda, outro estudo

identificou que níveis diminuídos de PRDX-6 estavam associados a alterações da morfologia celular e morte neuronal no córtex de roedores expostos ao tratamento com corticosterona (100).

Além das defesas antioxidantes enzimáticas, alguns compostos não enzimáticos importantes são provenientes da dieta, como as vitaminas E (tocoferol) e C (ácido ascórbico), o ácido úrico e o zinco (101). A glutationa, por sua vez, é o composto não enzimático antioxidante mais abundante no organismo, e desempenha um papel importante na proteção celular e de biomoléculas contra as EROs, além de ser um marcador endógeno sensível de estresse oxidativo. Ela pode ser encontrada na forma reduzida (GSH) ou oxidada (GSSG), por ação das enzimas glutationa redutase (GR) e GPx, respectivamente, sendo a razão GSH/GSSG amplamente utilizada para estimar o estado redox dos sistemas biológicos (102,103). Poucos estudos investigaram anormalidades relacionadas à GSH na DM, mas um estudo *post-mortem* detectou níveis diminuídos no CPF de indivíduos deprimidos (104).

A capacidade antioxidante total (TAC), por sua vez, pode fornecer informações sobre o estado antioxidante total de um indivíduo, incluindo antioxidantes ainda não reconhecidos ou não facilmente detectados (105). Contudo, esse ensaio avalia a capacidade antioxidante proveniente de compostos não enzimáticos, através de reações de oxidação e redução, geralmente não avaliando as atividades enzimáticas (106). Poucos estudos avaliaram a TAC em pacientes depressivos, todavia, alguns achados indicam que indivíduos com DM apresentam TAC diminuída, em detrimento do estado oxidante total (TOS) aumentado (107). Ainda, uma meta-análise conduzida por Liu e colaboradores corrobora esses dados, uma vez que também identificou níveis diminuídos de TAC e níveis aumentados de radicais livres, bem como de produtos provenientes de dano oxidativo em pacientes depressivos quando comparados a controles (108).

Níveis elevados de estresse oxidativo estão usualmente associados com prejuízo cognitivo e são considerados um dos potenciais mecanismos envolvidos com a neuropatologia e o envelhecimento precoce nos transtornos de humor (88). Um grande número de evidências indica que a produção exacerbada de EROs, levando ao aumento do estresse oxidativo, pode ser responsável por alterações neuronais que induzem a morte celular, promovendo, consequentemente, a atrofia de regiões específicas (109). Um menor volume hipocampal tem sido associado à resposta antidepressiva mais lenta e ao aumento de estresse oxidativo periférico na depressão tardia (110–112).

Em conjunto, essas evidências dão suporte ao envolvimento do estresse oxidativo na fisiopatologia da DM. Estudos clínicos e pré-clínicos que acessaram os efeitos de antidepressivos indicam que eles podem agir sobre as EROs sequestrando os radicais livres e suprimindo a via de estresse oxidativo. A ação dos antidepressivos contra os danos induzidos pelo desequilíbrio do estado redox parece mediar a remissão dos sintomas depressivos, bem como a recuperação dos pacientes (113). Crescentes evidências demonstram um potencial antioxidante dos antidepressivos, indicando que eles são capazes de restaurar e normalizar a atividade de enzimas como SOD, CAT e GPx (114,115), além de aumentar as concentrações de GSH e TAC (116), bem como diminuir os níveis de oxidação do DNA, de lipídios e de proteínas, atenuando também a morte celular induzida pela sinalização do H₂O₂ (117).

1.1.3.1 Estresse oxidativo, inflamação e BDNF

Evidências indicam que as bases biológicas da DM envolvem a interação entre fatores biológicos e ambientais, além de má adaptação na resposta ao estresse. Contudo, a forma como esses fatores são orquestrados e interagem entre si ainda não está bem elucidada (9). O aumento de marcadores de estresse oxidativo e de inflamação tem sido associado com a DM (89,118). Nesse contexto, as EROs, além de provocar dano às biomoléculas, atuam como mediadores na transdução de sinal de vias inflamatórias envolvendo o fator de transcrição

fator nuclear Kappa B (NF-κB) e as proteínas quinases ativadas por mitógeno (MAPK). Sugere-se ainda que a não adaptação das células frente às alterações do estado redox, a subsequente morte celular, e os danos provocados pelos mediadores inflamatórios estejam amplamente associados com a neuropatologia da DM. Dessa forma, a ativação do sistema imune e o aumento de estresse oxidativo induzem uma cascata de eventos orquestrada por fatores de transcrição como o fator nuclear eritróide 2 relacionado ao fator 2 (Nrf2) e o NF-κB, apresentando efeitos sinérgicos sobre a patogênese da DM (119).

O fator de transcrição Nrf2 exerce um papel fundamental na homeostase redox. Em baixas concentrações de EROS, ele é ativado e estimula a transcrição de genes com papel antioxidante, levando a efeitos protetores (119). Mellon e colaboradores reportaram que genes regulados pelo Nrf2 estavam aumentados em pacientes com DM, sugerindo que uma resposta antioxidante estava sendo requerida, e tiveram sua expressão diminuída após o tratamento antidepressivo (120). Além disso, um recente estudo pré-clínico mostrou que a suscetibilidade à DM era resultado de um estado persistente de estresse oxidativo, mediado por uma disfunção do Nrf2, o que foi revertido após tratamento com antioxidantes (121).

Em contrapartida, o aumento de EROS promove a ativação e translocação do NF-κB para o núcleo, podendo ativar cascatas inflamatórias, pró-oxidantes ou antioxidantes, dependendo do contexto celular (119). A ativação inicial do eixo HPA e a elevação de GCs são tipicamente associadas a respostas anti-inflamatórias, incluindo o bloqueio da sinalização do NF-κB (122,123). No entanto, o estresse pode aumentar os efeitos do NF-κB, intensificando a inflamação (124–126). Koo e colaboradores concluíram que a sinalização IL-1 β /NF-κB é ativada pelo estresse crônico, e que essa via foi necessária para o desenvolvimento de comportamento anedônico, bem como para os efeitos anti-neurogênicos observados. Ainda, sugere-se que o bloqueio do NF-κB possa inibir a ação de outras citocinas pró-inflamatórias implicadas na resposta ao estresse e na DM (127).

Strawbridge e colaboradores reportaram que concentrações aumentadas de biomarcadores inflamatórios estavam relacionadas à resposta parcial ao tratamento com antidepressivos, enquanto a inflamação persistentemente aumentada foi capaz de predizer os indivíduos não respondedores (128). Nesse contexto, outro estudo já havia demonstrado que os pacientes depressivos com concentrações mais elevadas de IL-6 eram menos propensos a responder ao tratamento antidepressivo (129).

Inúmeras evidências clínicas e pré-clínicas indicam que a diminuição dos níveis de BDNF, uma neurotrofina amplamente distribuída no cérebro e na periferia, pode ser revertida com tratamento antidepressivo, havendo, em partes, a melhora de sintomas da DM. Desse modo, fica claro que o mecanismo de resposta farmacológica dos antidepressivos está diretamente relacionado à ação do BDNF (130). Somado ao desequilíbrio do eixo HPA e à diminuição dos níveis de BDNF, Kunugi e colaboradores mostraram que os receptores RG interagem diretamente com os receptores TrkB, e que a exposição persistente aos GCs reduz essa interação, diminuindo a sinalização BDNF/TrkB e, consequentemente, colaborando com prejuízo dos efeitos neurotróficos (131).

O BDNF participa diretamente em processos como o desenvolvimento neuronal, neurogênese, plasticidade sináptica e arborização dendrítica, além de participar da consolidação da memória (132). De forma interessante, além de suas funções associadas à resposta imune, o fator de transcrição NF- κ B também pode atuar na sobrevivência celular e na plasticidade sináptica. Uma revisão recente reuniu diversas evidências, sugerindo que o NF- κ B regula a expressão do BDNF, e que o BDNF pode induzir a ativação do NF- κ B. É proposto que esse *feedback* positivo possa estar envolvido com as ações antidepressivas que levam ao aumento da neurogênese e da plasticidade neuronal, com a restauração da transmissão sináptica e com alterações comportamentais positivas, embora esse mecanismo não esteja completamente elucidado (133).

A ativação prolongada do eixo HPA com aumento persistente de GCs, bem como alterações envolvendo o estado redox e a inflamação em resposta ao estresse, usualmente levam ao prejuízo da neurogênese, o que parece estar associado à neuropprogressão na DM (89,134). Em conjunto, esses resultados sugerem que o prejuízo nos níveis de BDNF e na sua via de sinalização, bem como o perfil pró-inflamatório e pró-oxidativo podem estar relacionados com a diminuição da resposta ao tratamento antidepressivo.

1.2 Resiliência e suscetibilidade ao estresse

Perante a exposição ao estresse, a maior parte dos indivíduos exibe uma resposta adaptativa efetiva que viabiliza a superação da adversidade, caracterizando-os como resilientes. Em contrapartida, uma parcela menor da população demonstra uma habilidade limitada de lidar com esses eventos, apresentando um comprometimento das respostas adaptativas e consequentemente sendo mais suscetível ao desenvolvimento de doenças associadas ao estresse, como a DM (135,136).

Em um recente estudo clínico, García-Leon e colaboradores reportaram que a resiliência está relacionada com diferentes manifestações subjetivas de estresse em adultos saudáveis. Seus resultados sugerem que a resiliência atua como um amortecedor da percepção ao estresse, e que a diminuição da auto percepção permite que os eventos estressores sejam enfrentados com mais sucesso (137). Ainda, estudos comparando técnicas comportamentais, moleculares e eletrofisiológicas indicam que a resposta não adaptativa ao estresse envolve alterações em circuitos neurais específicos que regulam a recompensa, a resposta emocional, o medo e o comportamento social, prejudicando o enfrentamento do evento estressor (138). É importante destacar que o fenótipo resiliente parece estar relacionado a um processo neurobiológico distinto, não representando apenas a ausência de vulnerabilidade (139).

Dois modelos animais têm sido muito utilizados para estudar o paradigma de suscetibilidade e resiliência ao estresse. No modelo de derrota social os animais demonstram

diferenças individuais mais pronunciadas, apresentando uma desvantagem em comparação ao modelo de estresse crônico moderado e imprevisível (CUMS) (140). Contudo, a exposição a esse modelo geralmente identifica um maior número de animais resilientes ao estresse, o que se assemelha ao contexto clínico da DM, onde a vulnerabilidade individual interage com os eventos adversos que o indivíduo experimenta ao longo da vida, sendo um bom modelo para estudar os mecanismos envolvidos com a resiliência ao estresse (141). No modelo de CUMS, por sua vez, tem sido demonstrado, que cerca da metade dos animais expostos ao protocolo de estresse desenvolvem comportamento anedônico, sugerindo que ele seja uma ferramenta robusta para o estudo da suscetibilidade ao estresse (142–145). Alguns resultados baseados no protocolo de CUMS indicam que os animais suscetíveis apresentam diferenças, por exemplo, na neurogênese hipocampal (146) na performance cognitiva (147) na resposta inflamatória (143) e nos padrões de estresse oxidativo (148) em relação aos animais resilientes.

1.3 CUMS

Os modelos animais neuropsiquiátricos representam uma ferramenta pré-clínica extremamente desafiadora, dada a natureza subjetiva de muitos sintomas, a ausência de biomarcadores específicos, e as dificuldades de transpor os achados experimentais para o contexto clínico dos transtornos psiquiátricos. No entanto, o progresso na compreensão da fisiopatologia, bem como o avanço acerca dos possíveis tratamentos e a melhora da resposta terapêutica podem ser beneficiados por essa abordagem (149).

Idealmente, a fim de que um modelo animal seja adequado para o estudo de alguma doença, ele deve obedecer a três validades: de face, de construto e preditiva. A validade de face se refere à semelhança entre a sintomatologia clássica da doença e o comportamento desenvolvido pelos animais frente ao protocolo experimental. A validade de construto consiste em reproduzir características das bases biológicas da doença e a validade preditiva,

por sua vez, avalia o quanto as alterações comportamentais podem ser prevenidas ou revertidas através de fármacos classicamente utilizados no tratamento clínico (150).

Dentre os modelos animais utilizados no estudo da DM, destaca-se o modelo de CUMS, que surgiu em meados dos anos 80, baseado na perda de resposta à recompensa em animais submetidos a um cronograma variável de estressores e na reversão desse comportamento após tratamento antidepressivo (151,152). No CUMS, os animais são expostos diariamente a diferentes estressores, a fim de estimular a resposta ao estresse prevenindo uma possível adaptação. Dentre os estudos descritos na literatura, encontram-se variações nos protocolos experimentais. Contudo, uma recente revisão de Willner indicou que os estressores mais utilizados consistem na exposição à maravilha molhada, inclinação da caixa moradia, inversão do ciclo claro-escuro, privação de água e de comida e a superpopulação de roedores em uma mesma caixa (153).

Após verificar-se alta reproduzibilidade, ficou estabelecido que a exposição ao CUMS é capaz de induzir a diminuição do consumo ou da preferência por sacarose em animais suscetíveis, evidenciando a manifestação do comportamento anedônico. Esses achados promoveram de forma crescente o interesse da comunidade científica em utilizar esse modelo experimental para o estudo da DM. Transpondo para o contexto clínico, essa característica faz referência a um sintoma central da DM, a anedonia, e, portanto, fornece validade de face ao modelo experimental (154,155).

O modelo de CUMS também é amplamente utilizado como um indutor do desequilíbrio do eixo HPA e de inflamação. Um estudo conduzido por Liu e colaboradores identificou um aumento na concentração plasmática de IFN, TNF- α , e na atividade da IDO, além da diminuição nas concentrações de serotonina no CPF de ratos expostos ao CUMS (156). Ainda, um estudo de Rosseti e colaboradores demonstrou que o comportamento anedônico estava associado à presença de neuroinflamação, caracterizada pelo aumento de IL-

1, IL-6 e do marcador de ativação microglial CD11b. Essas alterações foram detectadas nos animais que demonstraram uma diminuição no consumo da sacarose, mas não nos animais resilientes (143). Recentemente, Xie e colaboradores reportaram que animais estressados com altos níveis de IL-1 β e IL-6 apresentaram menores concentrações de BDNF no CPF (157). Além disso, já havia sido demonstrado que a IL-1 β se comportou como um mediador do efeito antineurogênico, e que estava associada ao comportamento anedônico em ratos submetidos ao estresse agudo ou crônico (158).

Diferenças no padrão de estresse oxidativo de animais submetidos ao CUMS também tem sido descritas (159–161). Entretanto, um número limitado de estudos avaliando os parâmetros oxidativos e o estado redox em estruturas do SNC de animais suscetíveis e resilientes ao estresse foi realizado. Wang e colaboradores reportaram o aumento da oxidação proteica e da atividade da CAT, e diminuição da atividade da SOD em hipocampo e córtex de animais com comportamento anedônico em comparação a animais resilientes e controles. Ao mesmo tempo, esse estudo não identificou alterações referentes à ocorrência de peroxidação lipídica nessas estruturas (148).

Considerando os dados apresentados, conclui-se que o modelo de CUMS apresenta as três validades – de face, de construto e preditiva – requeridas, sendo representativo para o estudo da DM. Ainda, considerando a forte relação existente entre a resposta ao estresse e o desenvolvimento da DM, sugere-se que o CUMS seja uma ferramenta robusta para estudar as características e os mecanismos envolvidos na resiliência e suscetibilidade envolvendo o estresse oxidativo, a inflamação e a neurogênese.

1.4 Proteômica e biologia de sistemas na DM

Abordagens independentes de hipótese, como a proteômica e a genômica, são consideradas ferramentas robustas para identificar mecanismos envolvidos com a fisiopatologia de diversas doenças, incluindo transtornos psiquiátricos. Considerando as

modificações pós-traducionais e a provável inconsistência na regulação do processo de tradução, a proteômica parece apresentar algumas vantagens como instrumento de investigação. Devido à dificuldade de acessar amostras de líquor e de tecido cerebral em humanos, e considerando evidências que apontam para uma disfunção na permeabilidade da barreira hematoencefálica na DM – viabilizando o trânsito de proteínas entre o cérebro e a periferia – alguns estudos têm investido na avaliação proteômica a partir de sangue periférico de pacientes depressivos. Xu e colaboradores, por exemplo, identificaram alterações na expressão de proteínas envolvendo o metabolismo lipídico e imunoregulação em pacientes depressivos, reforçando o envolvimento desses mecanismos na DM (162).

Além disso, o modelo animal de CUMS também têm sido utilizado como um instrumento de estudo para avaliações proteômicas, evidenciando proteínas diferencialmente expressas em estruturas relevantes para a patogênese da DM, a partir de análises via bioinformática. Um estudo de Yang e colaboradores identificou alterações em processos biológicos envolvendo o metabolismo da glutationa e o metabolismo energético no CPF de animais expostos ao CUMS (163). Achados recentes também sugerem um prejuízo no metabolismo de aminoácidos, a desregulação do metabolismo do glutamato, alterações no metabolismo de ácidos graxos bem como a expressão anormal de proteínas relacionadas à sinapse no hipocampo de animais suscetíveis ao estresse (164), além de alterações envolvendo proteínas importantes para a neurogênese (165). Ainda, Zhou e colaboradores reportaram a alteração de proteínas relacionadas à neurotransmissão glutamatérgica e ao processo de sinapse na amigdala de ratos expostos ao estresse crônico, sugerindo que mudanças na regulação e na estrutura dessas proteínas possam estar envolvidas com o prejuízo dos mecanismos de neuroplasticidade em animais suscetíveis ao estresse (166).

2 JUSTIFICATIVA

A DM é um transtorno psiquiátrico grave e altamente incapacitante que apresenta um elevado risco de suicídio (1–3). Atualmente, estima-se que apenas metade dos pacientes atinja a remissão completa dos sintomas, havendo altos índices de recaída, principalmente durante o primeiro ano (14). Evidências indicam que a DM é uma doença heterogênea e complexa, que resulta da interação entre diversos fatores biológicos e ambientais, o que dificulta o completo entendimento de sua neurobiologia. Um aspecto consistentemente associado à depressão é a hiperativação do eixo HPA, acompanhada do aumento persistente de GCs e da alteração da resposta adaptativa ao estresse (9). Cerca de 85% dos estudos envolvendo a DM reportam resistência aos GCs e exacerbação da sinalização inflamatória nos pacientes (167).

Ainda, condições adversas podem gerar um desequilíbrio no estado redox celular. Desse modo, sugere-se que uma não adaptação das células frente a essas alterações, com subsequente morte celular e danos provocados pelos mediadores inflamatórios, estejam relacionados com a neuropatologia da DM (88,119). A alteração de marcadores de estresse oxidativo e de inflamação parece afetar parâmetros neurotróficos como o BDNF, o que pode estar relacionado à redução de neuroplasticidade associada à DM (89,118,131). No entanto, ainda não está claro como a susceptibilidade ao estresse crônico e a anedonia estão associadas a essas alterações.

O CUMS é um modelo amplamente utilizado para promover o desequilíbrio do eixo HPA e a inflamação, além de ser capaz de reproduzir o paradigma de resiliência e vulnerabilidade ao estresse, induzindo a manifestação do comportamento anedônico em animais suscetíveis (142–145). Nesse sentido, visando compreender melhor a relação entre os componentes já descritos na patogênese da DM e a resposta ao estresse, se faz necessário estudar os efeitos do estresse crônico sobre o comportamento, parâmetros oxidativos e antioxidantes, bem como sobre as concentrações de BDNF em animais submetidos ao CUMS.

Ainda, considerando a complexidade dos mecanismos envolvidos no comportamento anedônico, torna-se válido executar através da bioinformática, a avaliação de bancos de dados de proteômica provenientes de estudos com protocolos semelhantes ao desse experimento. Nesse sentido, informações referentes ao enriquecimento de processos biológicos envolvendo estresse oxidativo e inflamação, além da identificação de proteínas diferencialmente expressas em animais anedônicos, podem ser úteis para corroborar os resultados experimentais do presente estudo e sugerir mecanismos envolvidos com a resposta ao estresse.

3 OBJETIVOS

3.1 Objetivo geral

Investigar os níveis de BDNF, a ocorrência de dano oxidativo e a capacidade antioxidante total em animais submetidos ao protocolo de CUMS.

3.2 Objetivos específicos

- Avaliar o comportamento tipo-depressivo utilizando os testes de preferência por sacarose, nado forçado e campo aberto;
- Classificar os animais em anedônicos ou não-anedônicos de acordo com alterações na preferência por sacarose;
- Verificar a relação entre o peso da adrenal e a massa corporal nos diferentes grupos experimentais;
 - Avaliar a peroxidação lipídica, a carbonilação de proteínas e a capacidade antioxidante total no CPF e hipocampo dos diferentes grupos experimentais;
 - Mensurar a concentração de BDNF hipocampal nos diferentes grupos experimentais;
 - Analisar bancos de dados de proteômica de estudos com protocolo experimental semelhante, e identificar proteínas envolvidas com o estresse oxidativo e com a sinalização da resposta inflamatória diferencialmente expressas em animais anedônicos;
- Relacionar a resposta ao estresse com os desfechos avaliados.

PARTE II

4 ARTIGO

4.1 Carta de submissão

14/03/2019

Email – Ellen Scotton – Outlook

Track your co-authored submission to Brain Behavior and Immunity

Brain Behavior and Immunity <EvideSupport@elsevier.com>

Qui, 14/03/2019 15:31

Para: ellensc7@hotmail.com <ellensc7@hotmail.com>

Dear Miss Scotton,

Submission no: BBI_2019_225

Submission title: The protective role of BDNF and PRDX-1 against central oxidative damage in anhedonic rats submitted to a chronic unpredictable mild stress protocol

Corresponding author: Dr Mauricio Kunz

Listed co-author(s): Professor Rafael Colombo, Professor Adriane Rosa, Miss Fernanda E. Valiati, Dr Tiago F. Lopez, Miss Giovana Bristot, Miss Alessandra E. Guerra, Mr Gabriel H. Hizo, Miss Gabriela M.P. Possebon, Miss Ellen Scotton, Miss Túani M. Silva, Dr Mirian Salvador, Miss Jéssica C. Reis, Miss Luiza P.Géa

Dr Kunz has submitted a manuscript to Brain Behavior and Immunity and listed you as a co-author. This email is to let you know we will be in contact with updates at each decision stage of the submission process.

The link below takes you to a webpage where you can sign in to our submission system using your existing Elsevier profile credentials or register to create a new profile. You will then have the opportunity to tailor these updates and view reviewer and editor comments once they become available.

http://www.evise.com/profile/api/navigate/BBI?resourceUrl=%2Fco-author%2F%3Fdgcid%3Dinvite_email_cauthoroutreach08891591%23%2FBBI%2Fsubmission%2FBBI_2019_225

If you are not a co-author of this manuscript, please contact Researcher Support at:
<https://service.elsevier.com>

Thank you very much for your submission and we will be in touch as soon as we have any news to share.

Brain Behavior and Immunity

If you do not wish to receive further update emails on your co-authored submission, you can unsubscribe via this link:

http://www.evise.com/co-author/#/BBI/unsubscribe/ellensc7@hotmail.com/M8MYIWzXgwhIGATljAdun5UJxFb_AguFVPoJ42fI6ByMyP6zAq7k7Q2A7F4soX4PGuJJPwZaAKbpJucC1sFVQ

4.2 Manuscrito

Title page

Title: The protective role of BDNF and PRDX-1 against central oxidative damage in anhedonic rats submitted to a chronic unpredictable mild stress protocol.

Authors: Ellen Scotton^{a,b}; Rafael Colombo^{a,c}; Jéssica C. Reis^a; Gabriela M.P. Possebon^a Gabriel H. Hizo^a; Fernanda E. Valiati^{a,d}; Luiza P.Géa^{a,e}; Giovana Bristot^{a,d}; Mirian Salvador^f; Tuani M. Silva^f, Alessandra E. Guerra^g; Tiago F. Lopez^h; Adriane R. Rosa^{a,b,e}; Maurício Kunz^{a,b}

^a Laboratório de Psiquiatria Molecular, Hospital de Clínicas de Porto Alegre, Porto Alegre, RS, Brazil

^b Programa de Pós-Graduação em Psiquiatria e Ciências do Comportamento, Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, RS, Brazil.

^c Universidade de Caxias do Sul (UCS), Caxias do Sul, RS, Brazil

^d Programa de Pós-Graduação em Bioquímica, UFRGS, Porto Alegre, RS, Brazil

^e Programa de Pós-Graduação em Farmacologia e Terapêutica, UFRGS, Porto Alegre, RS, Brazil

^f Laboratório de estresse oxidativo e antioxidantes, Instituto de Biotecnologia, UCS, Caxias do Sul, RS, Brazil

^g Easy Search Acessoria em Pesquisa, Grupo Diagnose, Caxias do Sul, RS, Brazil

^h Centro de Pesquisa Experimental, Hospital de Clínicas de Porto Alegre, Porto Alegre, RS, Brazil

Authors' e-mail addresses:

Ellen Scotton: ellensc7@hotmail.com

Rafael Colombo: rcolombo1@ucs.br

Jéssica C. Reis: reis.jessica96@gmail.com

Gabriela M.P. Possebon: gabrielappossebon@gmail.com

Gabriel H. Hizo: gh231483@gmail.com

Fernanda E. Valiati: fe.e.valiati@gmail.com

Luiza P.Géa: lupgea@gmail.com

Giovana Bristot: giubristot@gmail.com

Mirian Salvador: msalvado@ucs.br

Tuani M. Silva: tuanimendes@yahoo.com.br

Alessandra E. Guerra: aeggodoy@gmail.com

Tiago F. Lopez: tflopez@hcpa.edu.br

Adriane R. Rosa: adrianerrosa@gmail.com

Maurício Kunz: maukunz@gmail.com

***Corresponding author:**

Maurício Kunz, PhD

Department of Psychiatry

Federal University of Rio Grande do Sul, Porto alegre- RS, Brazil

Phone: +55 51 3359 8021

Email: maukunz@gmail.com

Word count: 4471

Abstract

Chronic activation of the HPA axis and sustained increase of glucocorticoids have been associated in major depression, and also related to changes involving neurotrophins and oxidative stress markers in response to inflammation. This study aimed to evaluate central measures of brain-derived neurotrophic factor (BDNF), oxidative damage and total antioxidant capacity of rats submitted to chronic unpredictable mild stress (CUMS) and investigate the relationship between BDNF levels and differentially expressed proteins involved in oxidative stress and inflammatory biological processes. Wistar male rats were subjected to CUMS for 6 weeks. Based on the sucrose preference test, animals were divided into anhedonic and non-anhedonic clusters. After brain tissue collection, oxidative damage, total antioxidant capacity, and BDNF levels were evaluated. In order to extend discussion of possible mechanisms involving neurobiological findings, a bioinformatics approach was performed to identify proteins involved with oxidative stress and inflammation pathways that were differentially expressed in anhedonic animals from other studies with similar experimental protocol. CUMS was associated with an increase in BDNF concentrations accompanied by a decrease in total antioxidant capacity, besides the absence of oxidative damage to lipids and proteins. Withal, bioinformatics proteomic approach indicated that anhedonic animals showed a peroxiredoxin-1 (PRDX-1) up-regulation and a down-regulation of proteins involved with apoptotic and inflammation signaling (RELA, ASK-1 and TAK-1) in the hippocampus. These evidences suggest that BDNF and PRDX-1 may represent an initial response against stress, with a compensatory role, preventing oxidative damage to lipids and proteins through the modulation of antioxidant defense, mainly in anhedonic animals.

Keywords

Major depression; anhedonic behavior; CUMS; sucrose preference; BDNF; PRDX-1; oxidative stress; proteomics

Highlights

- Increased BDNF levels, but no oxidative damage in the hippocampus from animal exposed to the CUMS.
- Anhedonic rats show PRDX-1 up-regulation and RELA, ASK-1 and TAK-1 down-regulation.
- BDNF and PRDX-1 may have a compensatory role and attenuate oxidative damage.

1. Introduction

Major depressive disorder (MDD) is a severe, chronic, highly disabling psychiatric disorder often associated with comorbidities, besides presenting a high risk for suicide (Buckner et al., 2019; Cameron et al., 2014; Lépine and Briley, 2011). Anhedonia is a central hallmark of MDD, which is characterized by lack of pleasure and loss of reactivity to positive stimuli (Nasca et al., 2015; Otte et al., 2016). MDD symptoms lead to impairment in functioning and decrease in quality of life, representing an important outgoing in public health costs (Ferrari et al., 2013; Luppa et al., 2007).

According to the World Health Organization (WHO), MDD is considered the primary cause of disability in the population. By the year 2020, MDD is estimated to rank second place among the diseases with the highest global prevalence (“WHO | Depression and Other Common Mental Disorders,” n.d.). Evidence indicates that the biological bases of MDD include interactions between genetic, epigenetic, biochemical and environmental factors, as well as hormonal changes related to stress response. Both oxidative stress and enhancement of the inflammatory signal play a role in the susceptibility to stress. However, the way these factors are orchestrated and interact is not well elucidated yet (Otte et al., 2016).

Most individuals exhibit an effective adaptive response to stress, which characterizes them as resilient. In contrast, others demonstrate a limited ability to coping with stress, presenting maladaptive responses that lead to stress-related diseases, such as MDD (Hjemdal et al., 2011; Wang et al., 2014). Under an acute stress response, glucocorticoids (GCs) exhibit immunosuppressive and anti-inflammatory properties (Liu et al., 2017). In turn, chronic stress significantly increases the peripheral levels of pro-inflammatory cytokines, which intensifies cortisol release, promotes desensitization of the hypothalamic-pituitary-adrenal (HPA) axis and enhance oxidative stress (Pariante and Miller, 2001; Raedler, 2011).

Persistent activation of the HPA axis and a sustained increase of GCs have been described in MDD (Hasler et al., 2004; Ising et al., 2007). Recently, Horowitz and colleagues have shown that about 85% of MDD studies report resistance to GCs and exacerbation of inflammatory signaling in depressive patients (Horowitz and Zunszain, 2015). Stress and augmented GCs concentration are associated with increased production of reactive oxygen species (ROS) and oxidative stress (Bjelaković et al., 2007; Flaherty et al., 2017; You et al., 2009). Several studies have already identified elevation in serum levels of malondialdehyde (MDA) in patients with MDD (Islam et al., 2018; Khajehnasiri et al., 2013; Mazereeuw et al., 2015). Also, a recent meta-analysis showed that increased lipid peroxidation and DNA oxidation are the most consistently stress markers elevated in MDD, usually with small to moderate effect sizes (Black et al., 2015). Moreover, antioxidant enzymes superoxide dismutase (SOD) and catalase (CAT) present alterations in their activity, and increased activities of both enzymes might be indicators of acute depressive episodes in MDD patients (Tsai and Huang, 2016).

Peroxiredoxins (PRDX), in turn, are antioxidant enzymes that act by decreasing the substrates used in hydrogen peroxide (H_2O_2) formation and modulating intracellular signaling (Antunes and Brito, 2017; Sies et al., 2017; Won et al., 2012). Findings showed decreased PRDX-6 levels in rodents exposed to corticosterone treatment, and this reduction was associated with impairments in cell morphology and neuronal death in the cerebral cortex (Skynner et al., 2006). In addition to modulation in the antioxidant enzymes activity, a postmortem study has reported lower GSH concentrations in the PFC of individuals with MDD (Gawryluk et al., 2011). A recent meta-analysis conducted by Liu and colleagues identified reduced levels of total antioxidant capacity (TAC) and increased levels of free radicals as well as oxidative damage products in depressive patients when compared to controls (Liu et al., 2015).

Beyond damage biomolecules, ROS act as mediators in signal transduction of inflammatory pathways involving nuclear factor kappa B (NF-κB). Furthermore, immune system activation and increased oxidative stress induce a cascade of events orchestrated by transcription factors such as the nuclear factor derived from erythroid 2 (Nrf2) and NF-κB, revealing synergistic effects on the pathogenesis of MDD. At higher levels of oxidative stress, NF-κB is activated and depending on the cellular context inflammatory cascades, pro-oxidant or antioxidant genes can be triggered. At low concentrations, ROS activate Nrf2 and stimulate the transcription of genes with antioxidant function, including the expression of antioxidant enzymes (Bakunina et al., 2015). BDNF is a neurotrophin that induces complex neuronal signaling cascades critical for cellular changes underlying synaptic plasticity. Also, a recent study showed that BDNF play a crucial role as an inducer of neuronal antioxidant responses inducing Nrf2 nuclear translocation (Bruna et al., 2018).

An increase in oxidative stress and inflammation are usually associated with cognitive impairment and are considered to be one of the potential mechanisms involved with neuroprogression and early aging in mood disorders (Maurya et al., 2016). A crescent body of evidence indicates that the excessive production of ROS and reduced antioxidant capacity lead to an increase in oxidative stress and may be responsible for neuronal changes inducing cell death, thus promoting the atrophy of specific regions involved in behavior (Michel et al., 2012). Therefore, we aim to evaluate central measures of BDNF, oxidative damage and antioxidant capacity of rats submitted to chronic unpredictable mild stress (CUMS), and investigate the relationship between BDNF levels and differentially expressed proteins involved in oxidative stress and inflammatory biological processes according to the different patterns of stress-susceptibility.

2. Materials and methods

2.1. Animals

Thirty-five male Wistar rats (45-day-old, 220-250g) were obtained from the Central Animal House of the Universidade Federal do Rio Grande do Sul, Porto Alegre, state of Rio Grande do Sul, Brazil. They were housed singly in standard polycarbonate rat cages under standardized environmental conditions: 12h light/dark cycle (lights on between 7:00 a.m. and 7:00 p.m.), controlled temperature ($22 \pm 1^{\circ}\text{C}$), and food and water available ad libitum. All experimental procedures were approved by the ethics committee for animal research of the Hospital de Clínicas de Porto Alegre (protocol 150353) and were carried out following the eighth edition of the National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals. All efforts were made to minimize the number of animals and their suffering.

2.2. Sucrose preference test (SPT)

After two weeks of laboratory habituation and housing conditions, the animals were exposed to water and 1% sucrose solution in eight baseline tests conducted twice a week in their home cage. After 12h of food and water deprivation, two bottles, one with 1% sucrose solution and another with water, stayed available for animals during 1h. Bottles were weighed before and after the test to evaluate sucrose intake. All analyses were performed half an hour after the beginning of the dark cycle. Based on sucrose preference rates in the final baseline test, animals with unstable and/or low basal sucrose preference (under 60%) were excluded. The remaining animals were divided into two paired groups: control (n= 7) and “to-be-stressed” (n= 16). The reduced sucrose preference, which is used as an index of anhedonia, was calculated according to the following formula: sucrose consumption/[water consumption + sucrose consumption] ×100%. Subsequently, sucrose preference was monitored once a week, under similar conditions, until the end of the study.

2.3. CUMS

CUMS protocol was based on previous studies (Willner, 2017). The control animals were housed and handled in a separate room and had no contact with the stressed group. They were deprived of food and water during 12 h before each sucrose test, but otherwise, food and water were freely available. The CUMS group was daily exposed to different stressors during six weeks. Based on most common components of the CUMS regime, the stress protocol consisted in food and water deprivation (24h), tail pinch (1min), cage tilt (24h), the light-dark reversal (24h), overcrowding (2h), immobilization (2h) and wet bedding (24h) (Fig. 1). Also, all animals were weighted on the first day of CUMS and after the last stressor, at the end of the protocol.

2.4. Anhedonic and non-anhedonic clusters

After six weeks of CUMS, sucrose preference test was the first behavioral outcome evaluated. Based on Nasca and colleagues, following statistical approach was used for the assignment of animals subjected to CUMS in anhedonic and non-anhedonic clusters, according to their sucrose preference. Animals which fell into the standard deviation from the mean of the control group were considered non-anhedonic since they showed a behavior similar to unstressed rats. Animals that fell outside the standard deviation of the mean of the control group were designated as anhedonic (Nasca et al., 2015).

2.5. Open field test (OFT)

An open-field test was used to assess locomotor activity. The test was performed in a 77 cm diameter field enclosed by a 50 cm high acrylic wall with an open top. The dark floor of the arena was divided into quadrants, and its internal space was empty. Individually and randomly, the animals were placed at the center of the apparatus, without previous habituation, to explore the open field for 5 min. The device was continuously cleaned with alcohol between the tests. We used a video camera connected to a computer to record the

behavior of each animal. The videos were analyzed using the ANYmaze behavioral tracking software, which provided the number of crossings and total distance traveled by each animal.

2.6. Forced swim test (FST)

FST test was performed as described previously by Porsolt, with minor modifications (Porsolt et al., 1977). In brief, rats were placed individually in a black cylinder (height: 50 cm, diameter: 30 cm) filled with 30 cm of water at 25°C. In this cylinder, rats may not touch the bottom or escape. In the first exposure, the training session, rats were placed on the water for 15 min of forced swimming. Then, 24h later rats were placed on the cylinder again for 5 min, to perform the test session. We used a video camera connected to a computer to record the behavior of each animal. The videos were analyzed using the Boris software (Friard and Gamba, 2016), which provide total duration (seconds) of immobility. Water in the tank was changed after every five sessions.

2.7. Euthanasia and tissue collection

Post-behavioral analysis, rats were euthanized by decapitation and brain were dissected. The brain tissue were immediately frozen on dry ice in eppendorf tubes and then stored at -80 °C for further analyses. PFC and one hemisphere of the hippocampus were used for determination of oxidative lipid and protein damage and trolox equivalent antioxidant capacity (TEAC). Furthermore, BDNF measurement was performed using the other hippocampus hemisphere, and adrenal glands were collected and weighted nearly after euthanasia.

2.8. Determination of thiobarbituric acid reactive substances (TBARS)

Lipid peroxidation was monitored by the formation of TBARS during an acid-heating reaction, according to a protocol adapted from Wills (Wills, 1966). Specifically, 400 µL of supernatant from each sample was combined with 600 µL of 15% trichloroacetic acid and

0,67% thiobarbituric acid. The mixture was heated at 100°C for 15 min. After cooling to room temperature (RT), the samples were centrifuged at 5200 xg for 5 min. The supernatants were isolated, and their absorbance was measured at 530 nm. Hydrolyzed 1,1,3,3-tetramethoxypropane (TMP) was used as a standard, and the results were expressed as nmol MDA/mg. Total protein levels were evaluated using the Bradford method (Bradford, 1976).

2.9. Determination of protein carbonyl

Oxidative damage to proteins was measured based on the reaction of protein carbonyl groups with 2,4-dinitrophenylhydrazine (DNPH) (Levine et al., 1990). For the assay, 200 µL of DNPH (10 mM) or 200 µL of HCl (2 M) were added to 50 µL of supernatants. The reaction mixture was incubated in the dark for 30 min, and vortexed every 10 min. 250 µL of 20 % trichloroacetic acid was added to each reaction mixture and centrifuged at 3300 xg for 10 min. The supernatants from each sample were discarded, and the pellets were washed three times with ethanol-ethyl acetate (1:1) to remove free reagents. The samples were centrifuged, and the pellets were resuspended in 1000 µL of urea solution (8 M) at 37 °C, during 15 min. Absorbance was read at 365 nm, and the results were expressed as nmol DNPH/mg. Total protein levels were evaluated using the Bradford method (Bradford, 1976).

2.10. Determination of TEAC

The screening of TAC was performed by TEAC method, through the ability of the sample to scavenging the radical ABTS⁺ [2,2-azino-bis (3-etylbenzotiazolin) -6-sulfonic acid]. The ABTS⁺ solution is formed from the reaction of 7 mM ABTS with 2.45 mM potassium persulphate. This solution was kept in the dark at RT for 12-16 h before use. The solution ABTS • + was diluted with 5mM phosphate buffer saline (PBS pH 7.4) until an absorbance of 0.700 ± 0.35 at 734 nm. Then, 1.0 mL of ABTS • + diluted solution was added to 10 µL of sample (Re et al., 1999). The absorbance was read exactly 6 minutes after the initial mixture. For quantification, a standard curve was used with Trolox solution, and the results were

expressed in $\mu\text{mol TEAC/mg}$. Total protein levels were evaluated using the Bradford method (Bradford, 1976).

2.11. Determination of BDNF protein levels

BDNF serum levels were determined by sandwich-ELISA using monoclonal antibodies specific for BDNF (R&D Systems, USA). Briefly, microtiter plates (96-well flat-bottom) were coated overnight at RT with monoclonal anti-BDNF antibody at 4 $\mu\text{g/mL}$ in phosphate-buffered saline (PBS). After that, plates were washed three times with wash buffer and blocked with PBS containing 5 % nonfat milk powder for 1 hour at RT. After washing, plates were incubated two hours at RT with the samples diluted 1:5 in sample diluent (PBS with 1 % bovine serum albumin - BSA) and the standard curve ranged from 15.63 to 1,000 pg/mL of BDNF. Plates were washed, and biotinylated anti-BDNF antibody at 0.2 $\mu\text{g/mL}$ in PBS was added, which was incubated for 2 hours at RT. After washing, incubation with streptavidin-peroxidase conjugate (diluted 1:1,000 in sample diluent) for 1 hour at RT was performed, and subsequently, plates were rewashed and incubated with the substrate (3,3',5,5'-Tetramethylbenzidine) for 20 minutes at room temperature. Finally, the stop solution was added, and the amount of BDNF was determined by measuring absorbance at 450 nm. Total protein levels were evaluated using the Bradford method (Bradford, 1976). The standard curve demonstrates a direct relation between optical density and BDNF concentration and the results were expressed in pg/mg of total protein.

2.12. Proteomics analyses: a bioinformatic approach

Studies on chronic stress were selected based on their methodology similarity and ability to identifying groups of animals that were susceptible to stress. The following process was performed: (1) "chronic mild stress" AND "proteomics" keywords retrieved 39 articles on PubMed; (2) only 7 studies performed proteomic evaluation in the hippocampus of rats submitted to chronic mild stress; and finally (3) 3 studies had similar methodology and used

bioinformatic approaches to evaluate their data. All studies used sucrose consumption to classify animals in anhedonic and non-anhedonic, using the same rationale and methods for this outcome. Input data was prepared using all differentially expressed proteins (DEP) in the three works (Han et al., 2015; Henningsen et al., 2012; Zhang et al., 2018).

2.13. Statistical analyses

Statistical analyses of experimental outcomes were performed using SPSS 18.0 version and p -values < 0.05 were considered statistically significant. The results were checked for normality by Shapiro-Wilk test and analyzed with one-way ANOVA followed by Bonferroni's test for post-hoc comparisons. In proteomic analyses, to all DEP, we assigned the fold change values of $+/- 1$ and p -value < 0.05 , only to use them in the enrichment analysis. Kyoto Encyclopedia of Genes and Genomes (KEGG) results and Gene Ontology Biological Process pathways were rechecked and plotted using the R software environment v.3.4.4 (Team, 2013) and the R package pathfindR v.1.2.1 (Ulgen et al., 2018) considering as significantly enriched pathways those with FDR < 0.05 in the function run_pathfindR.

3. Results

3.1. Effects of CUMS in behavioral assessment

Behavior outcomes are exposed in Fig. 2. Anhedonia is a hallmark symptom of MDD that can be evaluated in rodents through a decrease in the sucrose preference compared to water in the SPT. Remarkably, anhedonic rats showed a significant reduction in sucrose preference after CUMS compared to non-anhedonic and control groups ($p < 0.001$). On the other hand, locomotor activity in OFT and total immobility time in FST remained unchanged between all groups.

3.2. Body weight and adrenal glands relative weight

The following morphometric variables are described in Fig. 3. Body weight was recorded in the beginning and in the end of the CUMS period. The control group presented a higher weight gain compared to animals exposed to stress, both anhedonic ($p= 0.004$) and non-anhedonic ($p= 0.001$). Furthermore, anhedonic rats had significant higher adrenal relative weight in comparison to the control group ($p= 0.014$).

3.3. Oxidative parameters

Assessment of TBARS, protein carbonyl and TEAC in PFC and hippocampus of anhedonic, non-anhedonic and control groups are given in Table 1. Lipid peroxidation and protein carbonylation did not present differences between groups. However, lower TEAC levels were found in the PFC and hippocampus, respectively, of anhedonic ($p= 0.00$ and $p= 0.006$) and non-anhedonic ($p= 0.026$ and $p= 0.005$) animals compared to controls.

3.4. BDNF levels

Hippocampus BDNF protein levels among groups are reported in Fig. 4. One-way ANOVA test revealed significant differences between CUMS exposed animals and control group. In particular, the anhedonic group showed an augmentation of 1,4-fold ($p= 0.001$) and the non-anhedonic group presented an increase of 1,2-fold ($p= 0.009$) in BDNF protein compared to controls.

3.5. Proteomics findings

Interestingly, differently expressed groups of proteins were observed when incorporating all data, with no intersection among studies (Han et al., 2015; Henningsen et al., 2012; Zhang et al., 2018). The biological processes enriched in the studies are indicated in Fig. 5. Thus, Map3k5 (apoptosis signal-regulating kinase 1(ASK-1)), Map3k7 (transforming growth factor beta-activated kinase 1 (TAK-1)) and RELA (transcriptional factor p65 known as NF- κ B p65

subunit) were down-regulated, while PRDX1 was up-regulated in the hippocampus of animals presenting anhedonic behavior (see Table 2).

4. Discussion

The findings of our study indicate that CUMS protocol was associated with an increase in hippocampus BDNF concentrations accompanied by a decrease in TEAC, despite the absence of oxidative damage to lipids and proteins. Withal, bioinformatics proteomic approach indicated that anhedonic animals show a PRDX-1 up-regulation and RELA, ASK-1 and TAK-1 down-regulation in the hippocampus. Taken together, these data suggest that, mainly in animals with anhedonic behavior, BDNF may have a compensatory role, attenuating neuronal damage through the modulation of enzymatic antioxidant activity.

A same stressful event may be more or less significant for an individual according to previous experiences and with their vulnerability, even in identical twins or isogenic rodents (Fraga et al., 2005; Freund et al., 2013). It is generally thought that chronic stress can lead to depressive-like behavior in animals. Indeed, reduced sucrose preference is an indicator of anhedonic-like behavioral change, the core symptom of MDD (Antoniuk et al., 2019; Freund et al., 2013). Our findings demonstrate that CUMS decreased sucrose preference in rats susceptible to stress-induced anhedonic behavior, in association to a lower body weight gain and an increase of relative adrenal glands weight compared to controls, which is in accordance with previous findings (Lucca et al., 2008; Vollmayr and Henn, 2003; Zhang et al., 2014). In contrast, no changes in immobility time and locomotor activity were found.

Ulrich and colleagues have shown that an increased adrenal weight after chronic stress is due to hyperplasia and hypertrophy in specific adrenal subregions, which is associated with increased maximal corticosterone responses to HPA axis (Ulrich-Lai et al., 2006). GCs are the crucial hormones involved in stress adaptation because of their role in feedback regulation on the functioning of the HPA axis. Therefore, corticosterone expression can be regarded as a

signal of stress injury during exposure to a stressor. Usually, chronic stress is associated with impairment in HPA axis response, leading to persistent higher GCs levels and decreased levels of BDNF (McEwen, 2006; Zhang et al., 2014).

Interestingly, although our findings indicate that stress-induced hypertrophy in adrenal glands - mainly in anhedonic rats - an increase in hippocampal BDNF was found. Previous studies showed that acute stress with increased plasma corticosterone levels was accompanied by a high expression of BDNF and tropomyosin receptor kinase (TrkB) in the hippocampus (Marmigère et al., 2003; Shi et al., 2010). Furthermore, Adlard and colleagues reported that a chronic immobilization protocol resulted in a significant increase in hippocampal BDNF, as well as intermittent immobilization and chronic cold stress also demonstrated the same trend (Adlard et al., 2004). Taken together, these findings suggest that augmented BDNF levels may be part of a compensatory response to preserve hippocampal homeostasis, or a way of neuronal plasticity to cope with stressor stimuli.

Moreover, in addition to the stressed animals present increased levels of BDNF compared to controls, the anhedonic rats showed slightly higher levels than the non-anhedonic ones, which may point a more pronounced biological response in these animals, even though there was no statistic difference between groups. BDNF is a neurotrophin involved in central nervous system (CNS) homeostasis, as well as in synaptic transmission and plasticity, playing an essential role in survival, maintenance, and growth of neurons (Bathina and Das, 2015; Lipsky and Marini, 2007; Ninan, 2014; Yamada et al., 2002). In addition, BDNF modulates the expression of a range of other genes, interacting with neurotransmitter systems such as glutamatergic (Carvalho et al., 2008), dopaminergic (Berton et al., 2006) and serotonergic (Martinowich and Lu, 2008; Pezawas et al., 2008).

Neurotrophins such as BDNF, can also contribute to neuronal cell protection against oxidative stress (Ichim et al., 2012). As mentioned before, BDNF can induce Nfr2 nuclear

translocation, promoting the transcription of antioxidant genes and leading to protective effects against ROS (Bruna et al., 2018). Using differentiated rat pheochromocytoma cells (PC12) with neuron-like characteristics, Ogura and colleagues reported that subtoxic levels of oxidative stress induced by H₂O₂ stimulated BDNF expression, supporting the involvement of a sensitive mechanism to ROS underlying BDNF neuroprotective effects (Ogura et al., 2014). In this line, our results identified the absence of oxidative lipid and protein damage in animals exposed to CUMS. Although, a decrease in TEAC was detected in the PFC and hippocampus of these animals, which is in accordance with clinical findings that showed reduced TAC in acute episodes of depressed patients (Liu et al., 2015), probably indicating a rapid response against modifications in the redox state. Nevertheless, the TEAC assay measures the antioxidant capacity provided by non-enzymatic compounds (e.g., glutathione), based on redox chemical reactions, usually excluding the evaluation of enzymatic activities (Bartosz, 2010).

Interestingly, based on bioinformatics approach, it was evidenced that some enriched biological processes include H₂O₂ pathways (Fig. 5). At the same time, up-regulation of PRDX-1 was found in anhedonic animals (Table 2). In accordance, Palmfeldt and colleagues recently demonstrated that PRDX-1 and PRDX-2 were down-regulated in resilient animals compared to anhedonic animals, possibly indicating that resilient animals were submitted to lower oxidative stress (Palmfeldt et al., 2016). It has been proposed that PRDX reduce more than 90% of cellular H₂O₂ (Adimora et al., 2010; Cox et al., 2009). Thus, their central role as peroxide scavenging enzymes in the cellular arsenal of antioxidant enzymes, such as CAT and glutathione peroxidase (GPx), has been recently recognized (Knoops et al., 2016; Perkins et al., 2015). Considering that PRDX-1 is highly sensitive to H₂O₂ and that BDNF may also mediate antioxidant effects, it may be suggested that up-regulation of PRDX-1 along with the increase of BDNF, might represent an adjacent mechanism in the modulation of moderate

levels of oxidative stress, preventing oxidative damages such as lipid peroxidation and protein carbonylation. According to STICH: chemical association networks (“STITCH network view,” n.d.), the combined score for interactions between H₂O₂ and BDNF is high, meaning that these two biomolecules have a great biological interaction (Fig. 6).

In addition to up-regulation of PRDX-1 indicated by proteomic analyses, it was found downregulation of ASK-1 and TAK-1, kinases involved in signaling for cell apoptosis, and RELA, a subunit of NF-κB, in anhedonic animals (Table 2). At low levels of oxidative stress, Nrf2 is activated (even through BDNF) and initiates transcription of antioxidative genes (e.g., antioxidant enzymes), leading to cytoprotective effects. At higher levels of oxidative stress, NF-κB is activated, and depending on the cellular context, can activate inflammatory cascades, pro-oxidant or antioxidant genes (Bakunina et al., 2015). Anti-apoptotic proteins are also suggested to be necessary for physiological functioning of neurons and synapses as well as for resilience to stress. Shishkina and colleagues found that both short-term stress and acute glucocorticoid exposures induce anti-apoptotic proteins expression (e.g., Bcl-2) and may reflect an adaptive attempt of the brain to reduce potential apoptotic effects of these treatments on neuronal cells (Shishkina et al., 2015).

Based on our results, we may suggest that CUMS might have induced the generation of ROS in low levels, activating sensitive antioxidant responses through increased BDNF levels and PRDX-1 up-regulation instead of activating apoptotic pathways, which is supported by down-regulation of ASK-1, TAK-1, and RELA. It has been reported that unpredictable or acute stress tends to provide an excited state with augmented expression of BDNF, contributing to protection against stress (Marmigère et al., 2003; McEwen, 2006). Thus, BDNF and GCs appear to oppose each other, with BDNF reversing the excitability in hippocampal neurons induced by stress levels of corticosterone. However, it does not mean that these pathways would remain inactive if exposure to stress were more persistent. With

stress' maintenance, if the excitation is not attenuated, a decrease in BDNF expression may occur (Shi et al., 2010). Moreover, Hiroshi and colleagues showed that glucocorticoid receptor (GR) interacts directly with TrkB and persistent exposure to GCs reduces this interaction, decreasing BDNF/TrkB signaling and, consequently, collaborating to the impairment of neurotrophic effects (Kunugi et al., 2010).

5. Conclusion

To sum up, the absence of oxidative damage to lipids and proteins, as well as the decreased TEAC concentration and down-regulation of ASK1, TAK1 and RELA may suggest that even in animals with an anhedonic behavior, structural damage exerted by oxidative stress and inflammation in response to activation of HPA axis and increased corticosterone, may be attenuated by a compensatory response via increased BDNF and up-regulation of PRDX-1.

BDNF and PRDX-1 seem to be sensitive biomarkers to ROS (e.g., H₂O₂), and they might represent an initial response to stress to the maintenance of redox homeostasis, preventing oxidative damage to lipids and proteins, mainly in anhedonic rats. Nonetheless, further investigation is needed to elucidate the relationship among the stress response and biological compensatory mechanisms involving coping to chronic stress.

6. Conflict of interest statement

The authors declare no conflicts of interest.

Acknowledgements

The authors would like to thank the Fundo de Incentivo à Pesquisa – Hospital de Clínicas de Porto Alegre (FIPE-HCPA 150353), and the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) for funding support. Adriane R. Rosa would like to thank to CNPq, PQ number 305707-2015/09.

References

- Adimora, N.J., Jones, D.P., Kemp, M.L., 2010. A model of redox kinetics implicates the thiol proteome in cellular hydrogen peroxide responses. *Antioxid. Redox Signal.* 13, 731–743.
- Adlard, P.A., Perreau, V.M., Cotman, C.W., 2004. Chronic immobilization stress differentially affects the expression of BDNF mRNA and protein in the mouse hippocampus. *Stress and Health* 20, 175–180.
- Antoniuk, S., Bijata, M., Ponimaskin, E., Włodarczyk, J., 2019. Chronic unpredictable mild stress for modeling depression in rodents: Meta-analysis of model reliability. *Neurosci Biobehav Rev* 99, 101–116.
- Antunes, F., Brito, P.M., 2017. Quantitative biology of hydrogen peroxide signaling. *Redox Biology* 13, 1.
- Bakunina, N., Pariante, C.M., Zunszain, P.A., 2015. Immune mechanisms linked to depression via oxidative stress and neuroprogression. *Immunology* 144, 365–373.
- Bartosz, G., 2010. Non-enzymatic antioxidant capacity assays: Limitations of use in biomedicine. *Free Radic. Res.* 44, 711–720.
- Bathina, S., Das, U.N., 2015. Brain-derived neurotrophic factor and its clinical implications. *Arch Med Sci* 11, 1164–1178.
- Bertón, O., McClung, C.A., Dileone, R.J., Krishnan, V., Renthal, W., Russo, S.J., Graham, D., Tsankova, N.M., Bolanos, C.A., Rios, M., Monteggia, L.M., Self, D.W., Nestler, E.J., 2006. Essential role of BDNF in the mesolimbic dopamine pathway in social defeat stress. *Science* 311, 864–868.
- Bjelaković, G., Beninati, S., Pavlović, D., Kocić, G., Jevtović, T., Kamenov, B., Saranac, L.J., Bjelaković, B., Stojanović, I., Basić, J., 2007. Glucocorticoids and oxidative stress. *J Basic Clin Physiol Pharmacol* 18, 115–127.
- Black, C.N., Bot, M., Scheffer, P.G., Cuijpers, P., Penninx, B.W.J.H., 2015. Is depression associated with increased oxidative stress? A systematic review and meta-analysis. *Psychoneuroendocrinology* 51, 164–175.

- Bradford, M.M., 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 72, 248–254.
- Bruna, B., Lobos, P., Herrera-Molina, R., Hidalgo, C., Paula-Lima, A., Adasme, T., 2018. The signaling pathways underlying BDNF-induced Nrf2 hippocampal nuclear translocation involve ROS, RyR-Mediated Ca²⁺ signals, ERK and PI3K. *Biochem. Biophys. Res. Commun.* 505, 201–207.
- Buckner, J.D., Lewis, E.M., Tucker, R.P., 2019. Mental Health Problems and Suicide Risk: The Impact of Acute Suicidal Affective Disturbance. *Arch Suicide Res* 1–20.
- Cameron, C., Habert, J., Anand, L., Furtado, M., 2014. Optimizing the management of depression: primary care experience. *Psychiatry Res* 220 Suppl 1, S45-57.
- Carvalho, A.L., Caldeira, M.V., Santos, S.D., Duarte, C.B., 2008. Role of the brain-derived neurotrophic factor at glutamatergic synapses. *Br J Pharmacol* 153, S310–S324.
- Cox, A.G., Winterbourn, C.C., Hampton, M.B., 2009. Mitochondrial peroxiredoxin involvement in antioxidant defence and redox signalling. *Biochem. J.* 425, 313–325.
- Ferrari, A.J., Charlson, F.J., Norman, R.E., Patten, S.B., Freedman, G., Murray, C.J.L., Vos, T., Whiteford, H.A., 2013. Burden of depressive disorders by country, sex, age, and year: findings from the global burden of disease study 2010. *PLoS Med.* 10, e1001547.
- Flaherty, R.L., Owen, M., Fagan-Murphy, A., Intabli, H., Healy, D., Patel, A., Allen, M.C., Patel, B.A., Flint, M.S., 2017. Glucocorticoids induce production of reactive oxygen species/reactive nitrogen species and DNA damage through an iNOS mediated pathway in breast cancer. *Breast Cancer Res* 19.
- Fraga, M.F., Ballestar, E., Paz, M.F., Ropero, S., Setien, F., Ballestar, M.L., Heine-Suñer, D., Cigudosa, J.C., Urioste, M., Benitez, J., Boix-Chornet, M., Sanchez-Aguilera, A., Ling, C., Carlsson, E., Poulsen, P., Vaag, A., Stephan, Z., Spector, T.D., Wu, Y.-Z., Plass, C., Esteller, M., 2005. Epigenetic differences arise during the lifetime of monozygotic twins. *Proc. Natl. Acad. Sci. U.S.A.* 102, 10604–10609.

- Freund, J., Brandmaier, A.M., Lewejohann, L., Kirste, I., Kritzler, M., Krüger, A., Sachser, N., Lindenberger, U., Kempermann, G., 2013. Emergence of individuality in genetically identical mice. *Science* 340, 756–759.
- Friard, O., Gamba, M., 2016. BORIS: a free, versatile open-source event-logging software for video/audio coding and live observations. *Methods in Ecology and Evolution* 7, 1325–1330.
- Gawryluk, J.W., Wang, J.-F., Andreazza, A.C., Shao, L., Young, L.T., 2011. Decreased levels of glutathione, the major brain antioxidant, in post-mortem prefrontal cortex from patients with psychiatric disorders. *Int. J. Neuropsychopharmacol.* 14, 123–130.
- Han, X., Shao, W., Liu, Z., Fan, S., Yu, J., Chen, J., Qiao, R., Zhou, J., Xie, P., 2015. iTRAQ-based quantitative analysis of hippocampal postsynaptic density-associated proteins in a rat chronic mild stress model of depression. *Neuroscience* 298, 220–292.
- Hasler, G., Drevets, W.C., Manji, H.K., Charney, D.S., 2004. Discovering endophenotypes for major depression. *Neuropsychopharmacology* 29, 1765–1781.
- Henningsen, K., Palmfeldt, J., Christiansen, S., Baiges, I., Bak, S., Jensen, O.N., Gregersen, N., Wiborg, O., 2012. Candidate hippocampal biomarkers of susceptibility and resilience to stress in a rat model of depression. *Mol. Cell Proteomics* 11, M111.016428.
- Hjemdal, O., Vogel, P.A., Solem, S., Hagen, K., Stiles, T.C., 2011. The relationship between resilience and levels of anxiety, depression, and obsessive-compulsive symptoms in adolescents. *Clin Psychol Psychother* 18, 314–321.
- Horowitz, M.A., Zunszain, P.A., 2015. Neuroimmune and neuroendocrine abnormalities in depression: two sides of the same coin. *Ann. N. Y. Acad. Sci.* 1351, 68–79.
- Ichim, G., Tauszig-Delamasure, S., Mehlen, P., 2012. Neurotrophins and cell death. *Exp. Cell Res.* 318, 1221–1228.
- Ising, M., Horstmann, S., Kloiber, S., Lucae, S., Binder, E.B., Kern, N., Künzel, H.E., Pfennig, A., Uhr, M., Holsboer, F., 2007. Combined dexamethasone/corticotropin releasing hormone test predicts treatment response in major depression - a potential biomarker? *Biol. Psychiatry* 62, 47–54.

- Islam, Md Rabiul, Islam, Md Reazul, Ahmed, I., Moktadir, A.A., Nahar, Z., Islam, Mohammad Safiqul, Shahid, S.F.B., Islam, S.N., Islam, Md Saiful, Hasnat, A., 2018. Elevated serum levels of malondialdehyde and cortisol are associated with major depressive disorder: A case-control study. *SAGE Open Med* 6, 2050312118773953.
- Khajehnasiri, F., Mortazavi, S.B., Allameh, A., Akhondzadeh, S., Hashemi, H., 2013. Total antioxidant capacity and malondialdehyde in depressive rotational shift workers. *J Environ Public Health* 2013, 150693.
- Knoops, B., Argyropoulou, V., Becker, S., Ferté, L., Kuznetsova, O., 2016. Multiple Roles of Peroxiredoxins in Inflammation. *Mol Cells* 39, 60–64.
- Kunugi, H., Hori, H., Adachi, N., Numakawa, T., 2010. Interface between hypothalamic-pituitary-adrenal axis and brain-derived neurotrophic factor in depression. *Psychiatry and Clinical Neurosciences* 64, 447–459.
- Lépine, J.-P., Briley, M., 2011. The increasing burden of depression. *Neuropsychiatr Dis Treat* 7, 3–7.
- Levine, R.L., Garland, D., Oliver, C.N., Amici, A., Climent, I., Lenz, A.G., Ahn, B.W., Shaltiel, S., Stadtman, E.R., 1990. Determination of carbonyl content in oxidatively modified proteins. *Meth. Enzymol.* 186, 464–478.
- Lipsky, R.H., Marini, A.M., 2007. Brain-derived neurotrophic factor in neuronal survival and behavior-related plasticity. *Ann. N. Y. Acad. Sci.* 1122, 130–143.
- Liu, T., Zhong, S., Liao, X., Chen, J., He, T., Lai, S., Jia, Y., 2015. A Meta-Analysis of Oxidative Stress Markers in Depression. *PLoS ONE* 10, e0138904.
- Liu, Y.-Z., Wang, Y.-X., Jiang, C.-L., 2017. Inflammation: The Common Pathway of Stress-Related Diseases. *Front Hum Neurosci* 11, 316.
- Lucca, G., Comim, C.M., Valvassori, S.S., Pereira, J.G., Stertz, L., Gavioli, E.C., Kapczinski, F., Quevedo, J., 2008. Chronic mild stress paradigm reduces sweet food intake in rats without affecting brain derived neurotrophic factor protein levels. *Curr Neurovasc Res* 5, 207–213.
- Luppa, M., Heinrich, S., Angermeyer, M.C., König, H.-H., Riedel-Heller, S.G., 2007. Cost-of-illness studies of depression: a systematic review. *J Affect Disord* 98, 29–43.

- Marmigère, F., Givalois, L., Rage, F., Arancibia, S., Tapia-Arancibia, L., 2003. Rapid induction of BDNF expression in the hippocampus during immobilization stress challenge in adult rats. *Hippocampus* 13, 646–655.
- Martinowich, K., Lu, B., 2008. Interaction between BDNF and serotonin: role in mood disorders. *Neuropsychopharmacology* 33, 73–83.
- Maurya, P.K., Noto, C., Rizzo, L.B., Rios, A.C., Nunes, S.O.V., Barbosa, D.S., Sethi, S., Zeni, M., Mansur, R.B., Maes, M., Brietzke, E., 2016. The role of oxidative and nitrosative stress in accelerated aging and major depressive disorder. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 65, 134–144.
- Mazereeuw, G., Herrmann, N., Andreazza, A.C., Khan, M.M., Lanctôt, K.L., 2015. A meta-analysis of lipid peroxidation markers in major depression. *Neuropsychiatr Dis Treat* 11, 2479–2491.
- McEwen, B.S., 2006. Protective and damaging effects of stress mediators: central role of the brain. *Dialogues Clin Neurosci* 8, 367–381.
- Michel, T.M., Pülschen, D., Thome, J., 2012. The role of oxidative stress in depressive disorders. *Curr. Pharm. Des.* 18, 5890–5899.
- Nasca, C., Bigio, B., Zelli, D., Nicoletti, F., McEwen, B.S., 2015. Mind the gap: glucocorticoids modulate hippocampal glutamate tone underlying individual differences in stress susceptibility. *Mol Psychiatry* 20, 755–763.
- Ninan, I., 2014. Synaptic regulation of affective behaviors; role of BDNF. *Neuropharmacology* 76.
- Ogura, Y., Sato, K., Kawashima, K.-I., Kobayashi, N., Imura, S., Fujino, K., Kawaguchi, H., Nedachi, T., 2014. Subtoxic levels of hydrogen peroxide induce brain-derived neurotrophic factor expression to protect PC12 cells. *BMC Res Notes* 7, 840.
- Otte, C., Gold, S.M., Penninx, B.W., Pariante, C.M., Etkin, A., Fava, M., Mohr, D.C., Schatzberg, A.F., 2016. Major depressive disorder. *Nat Rev Dis Primers* 2, 16065.

- Palmfeldt, J., Henningsen, K., Eriksen, S.A., Müller, H.K., Wiborg, O., 2016. Protein biomarkers of susceptibility and resilience to stress in a rat model of depression. *Mol. Cell. Neurosci.* 74, 87–95.
- Pariante, C.M., Miller, A.H., 2001. Glucocorticoid receptors in major depression: relevance to pathophysiology and treatment. *Biol. Psychiatry* 49, 391–404.
- Perkins, A., Nelson, K.J., Parsonage, D., Poole, L.B., Karplus, P.A., 2015. Peroxiredoxins: guardians against oxidative stress and modulators of peroxide signaling. *Trends Biochem. Sci.* 40, 435–445.
- Pezawas, L., Meyer-Lindenberg, A., Goldman, A.L., Verchinski, B.A., Chen, G., Kolachana, B.S., Egan, M.F., Mattay, V.S., Hariri, A.R., Weinberger, D.R., 2008. Evidence of biologic epistasis between BDNF and SLC6A4 and implications for depression. *Mol. Psychiatry* 13, 709–716.
- Porsolt, R.D., Le Pichon, M., Jalfre, M., 1977. Depression: a new animal model sensitive to antidepressant treatments. *Nature* 266, 730–732.
- Raedler, T.J., 2011. Inflammatory mechanisms in major depressive disorder. *Curr Opin Psychiatry* 24, 519–525.
- Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M., Rice-Evans, C., 1999. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology and Medicine* 26, 1231–1237.
- Shi, S.-S., Shao, S.-H., Yuan, B.-P., Pan, F., Li, Z.-L., 2010. Acute stress and chronic stress change brain-derived neurotrophic factor (BDNF) and tyrosine kinase-coupled receptor (TrkB) expression in both young and aged rat hippocampus. *Yonsei Med. J.* 51, 661–671.
- Shishkina, G.T., Kalinina, T.S., Bulygina, V.V., Lanshakov, D.A., Babluk, E.V., Dygalo, N.N., 2015. Anti-Apoptotic Protein Bcl-xL Expression in the Midbrain Raphe Region Is Sensitive to Stress and Glucocorticoids. *PLOS ONE* 10, e0143978.
- Sies, H., Berndt, C., Jones, D.P., 2017. Oxidative Stress. *Annual Review of Biochemistry* 86, 715–748.

Skynner, H.A., Amos, D.P., Murray, F., Salim, K., Knowles, M.R., Munoz-Sanjuan, I., Camargo, L.M., Bonnert, T.P., Guest, P.C., 2006. Proteomic analysis identifies alterations in cellular morphology and cell death pathways in mouse brain after chronic corticosterone treatment. *Brain Res.* 1102, 12–26.

STITCH network view. <http://stitch.embl.de/cgi/network.pl?taskId=h07KR7iXK0cu> (accessed 3.8.19).

Team, R.C., 2013. R: A language and environment for statistical computing.

Tsai, M.-C., Huang, T.-L., 2016. Increased activities of both superoxide dismutase and catalase were indicators of acute depressive episodes in patients with major depressive disorder. *Psychiatry Res* 235, 38–42.

Ulgen, E., Ozisik, O., Sezerman, O.U., 2018. pathfindR: An R Package for Pathway Enrichment Analysis Utilizing Active Subnetworks. *bioRxiv* 272450.

Ulrich-Lai, Y.M., Figueiredo, H.F., Ostrander, M.M., Choi, D.C., Engeland, W.C., Herman, J.P., 2006. Chronic stress induces adrenal hyperplasia and hypertrophy in a subregion-specific manner. *Am. J. Physiol. Endocrinol. Metab.* 291, E965-973.

Vollmayr, B., Henn, F.A., 2003. Stress models of depression. *Clinical Neuroscience Research* 3, 245–251.

Wang, M., Perova, Z., Arenkiel, B.R., Li, B., 2014. Synaptic modifications in the medial prefrontal cortex in susceptibility and resilience to stress. *J. Neurosci.* 34, 7485–7492.

WHO: World Health Organization, 2017. Depression and Other Common Mental Disorders http://www.who.int/mental_health/management/depression/prevalence_global_health_estimates/en/ (accessed 3.7.19).

Willner, P., 2017. The chronic mild stress (CMS) model of depression: History, evaluation and usage. *Neurobiol Stress* 6, 78–93.

Wills, E.D., 1966. Mechanisms of lipid peroxide formation in animal tissues. *Biochem. J.* 99, 667–676.

- Won, H., Lim, S., Jang, M., Kim, Y., Rashid, M.A., Jyothi, K.R., Dashdorj, A., Kang, I., Ha, J., Kim, S.S., 2012. Peroxiredoxin-2 upregulated by NF-κB attenuates oxidative stress during the differentiation of muscle-derived C2C12 cells. *Antioxid. Redox Signal.* 16, 245–261.
- Yamada, K., Mizuno, M., Nabeshima, T., 2002. Role for brain-derived neurotrophic factor in learning and memory. *Life Sci.* 70, 735–744.
- You, J.-M., Yun, S.-J., Nam, K.N., Kang, C., Won, R., Lee, E.H., 2009. Mechanism of glucocorticoid-induced oxidative stress in rat hippocampal slice cultures. *Can. J. Physiol. Pharmacol.* 87, 440–447.
- Zhang, L., Luo, J., Zhang, M., Yao, W., Ma, X., Yu, S.Y., 2014. Effects of curcumin on chronic, unpredictable, mild, stress-induced depressive-like behaviour and structural plasticity in the lateral amygdala of rats. *Int. J. Neuropsychopharmacol.* 17, 793–806.
- Zhang, Y., Yuan, S., Pu, J., Yang, L., Zhou, X., Liu, L., Jiang, X., Zhang, H., Teng, T., Tian, L., Xie, P., 2018. Integrated Metabolomics and Proteomics Analysis of Hippocampus in a Rat Model of Depression. *Neuroscience* 371, 207–220.

Figures

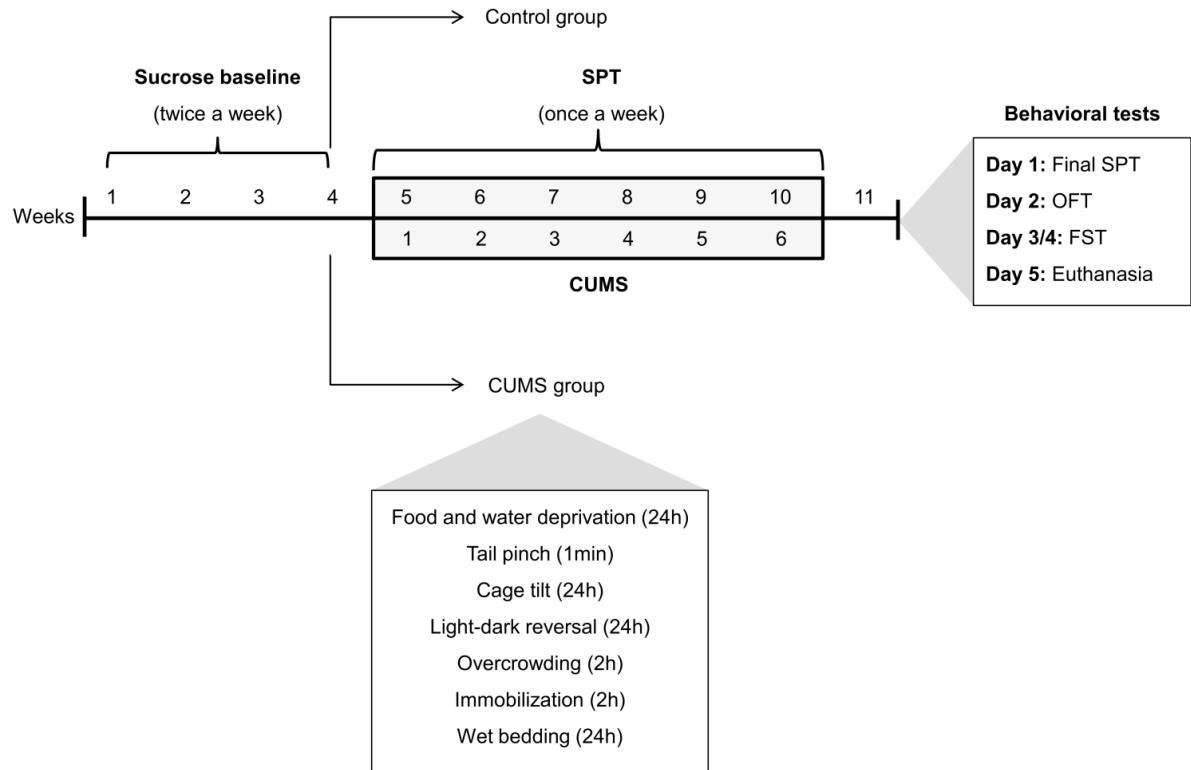


Fig.1. Schematic representation of the experimental procedures. CUMS, chronic unpredictable mild stress; FST, forced swim test; OFT, open field test; SPT, sucrose preference test.

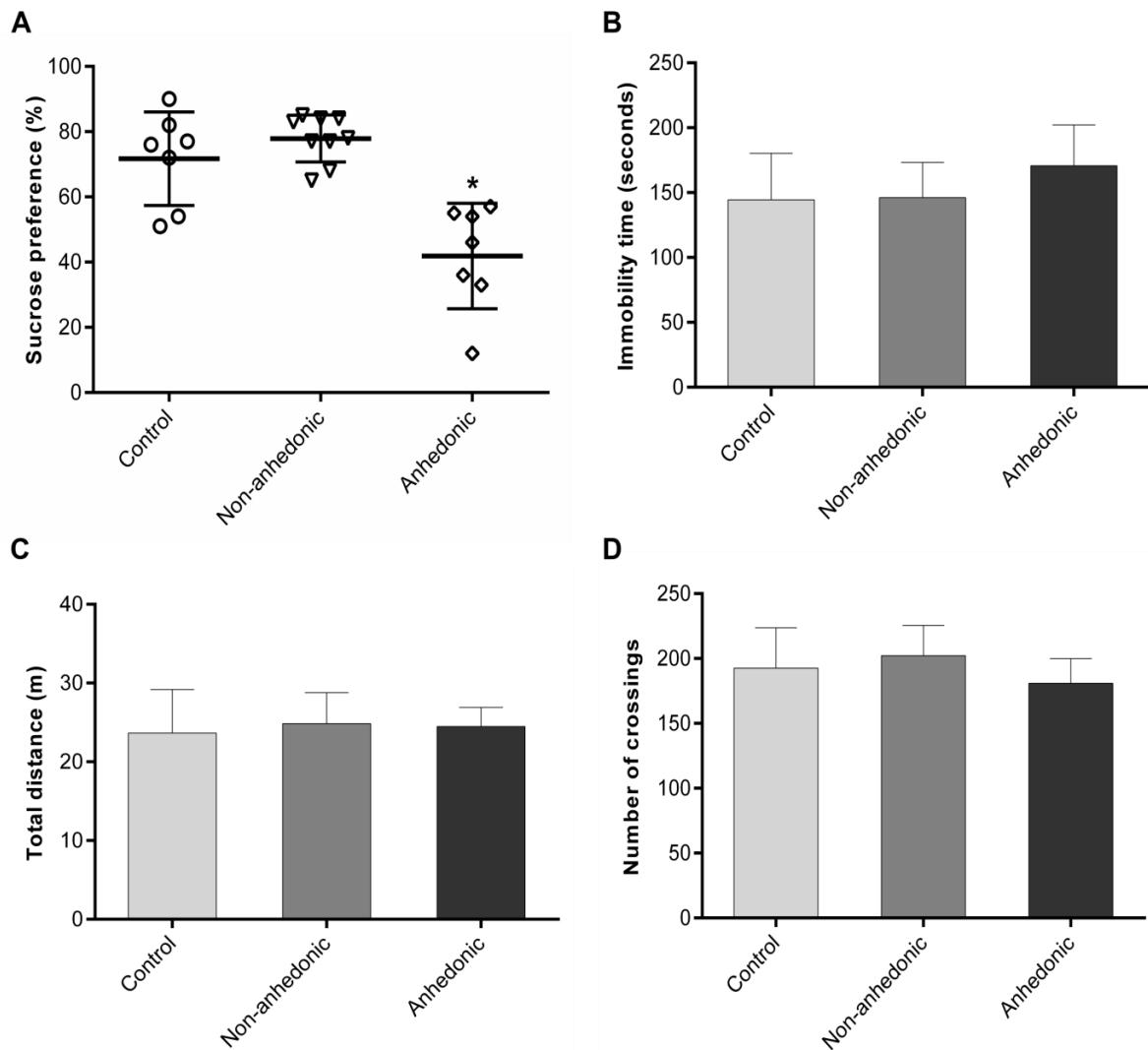


Fig. 2. Effects of CUMS on behavior assessments. (A) Sucrose preference test was performed to assign anhedonic and non-anhedonic clusters after six weeks of CUMS, according to sucrose preference rate. (B) Total immobility time was assessed by forced swim test, and the open-field test indicates (C) total distance traveled and (D) number of crossings. All values are shown as mean \pm SD. Groups: anhedonic ($n=7$), non-anhedonic ($n=9$) and control ($n=7$).

* $p \leq 0.001$ according to one-way ANOVA followed by Bonferroni's post-hoc test. CUMS, chronic unpredictable mild stress; SD, standard deviation.

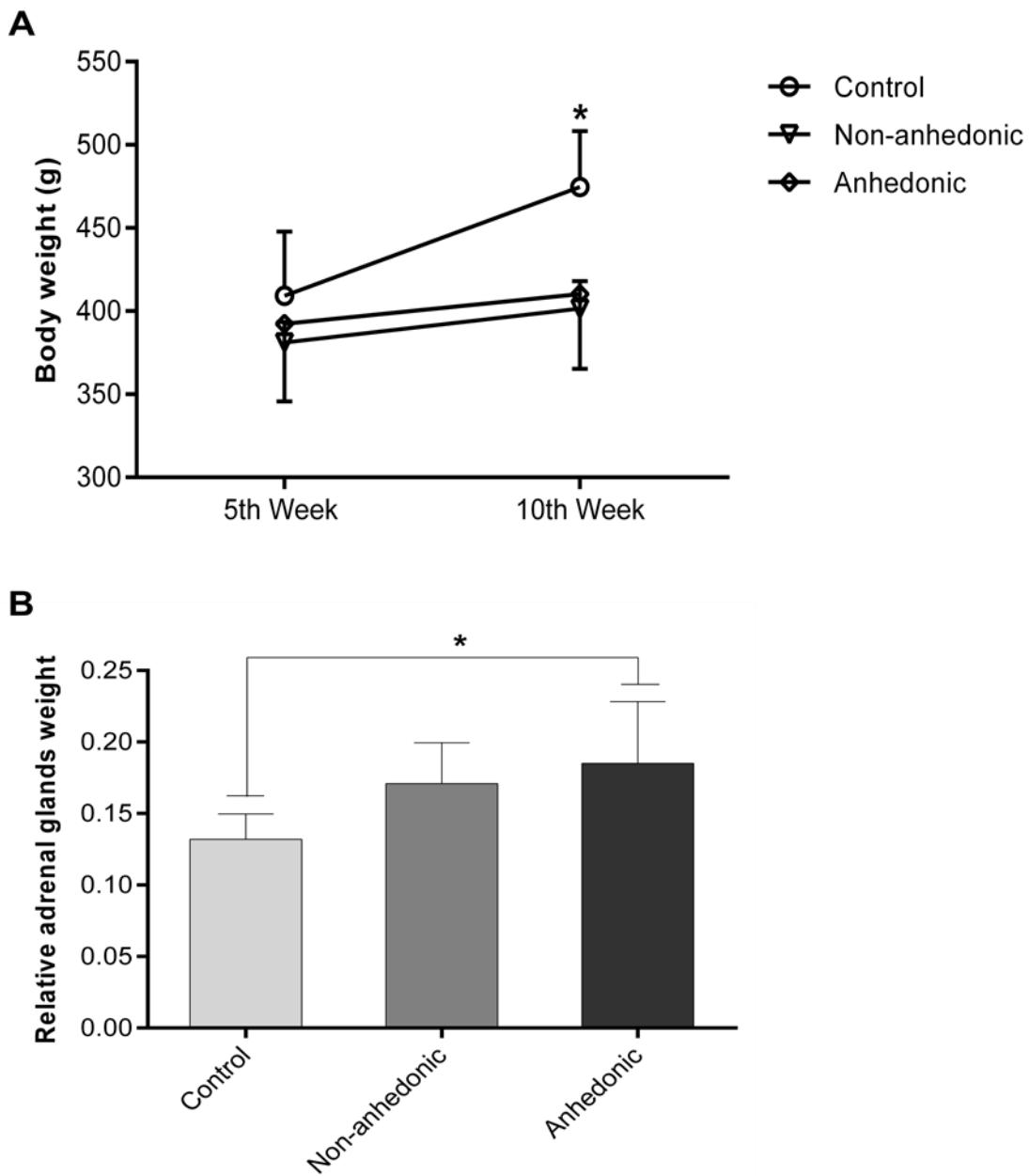


Fig. 3. Effects of CUMS in morphometric variables. (A) Animals' body weight before and after CUMS protocol and (B) relative adrenal glands weight (adrenals weight/body weight) among the groups. All values are shown as mean \pm SD. Groups: anhedonic (n= 7), non-anhedonic (n= 9) and control (n= 7). * $p\leq 0.01$ or ** $p\leq 0.001$ according to one-way ANOVA followed by Bonferroni's post-hoc test. CUMS, chronic unpredictable mild stress; SD, standard deviation.

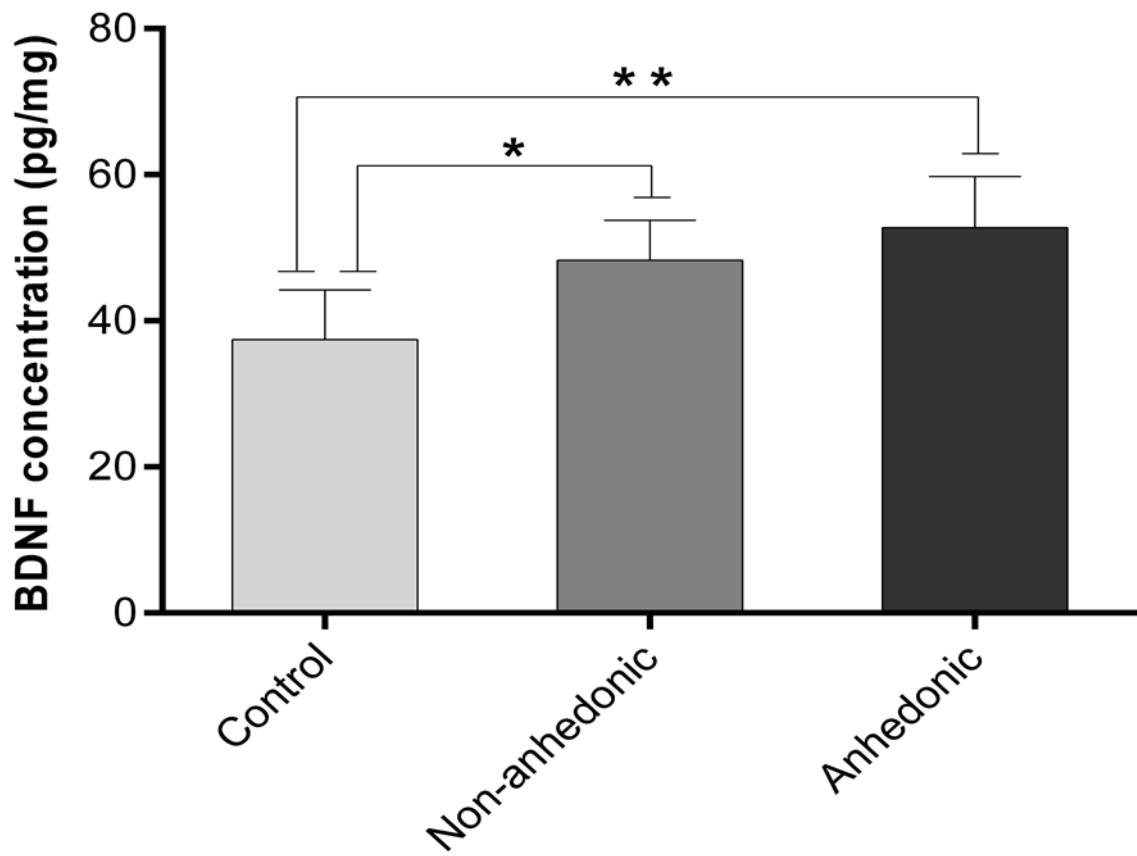


Fig. 4. Effects of CUMS in BDNF protein levels in the hippocampus. All values are shown as mean \pm SD. Groups: anhedonic (n= 7), non-anhedonic (n= 9) and control (n= 7). *p \leq 0.01 or **p \leq 0.001 according to one-way ANOVA followed by Bonferroni's post-hoc test. CUMS, chronic unpredictable mild stress; SD, standard deviation.

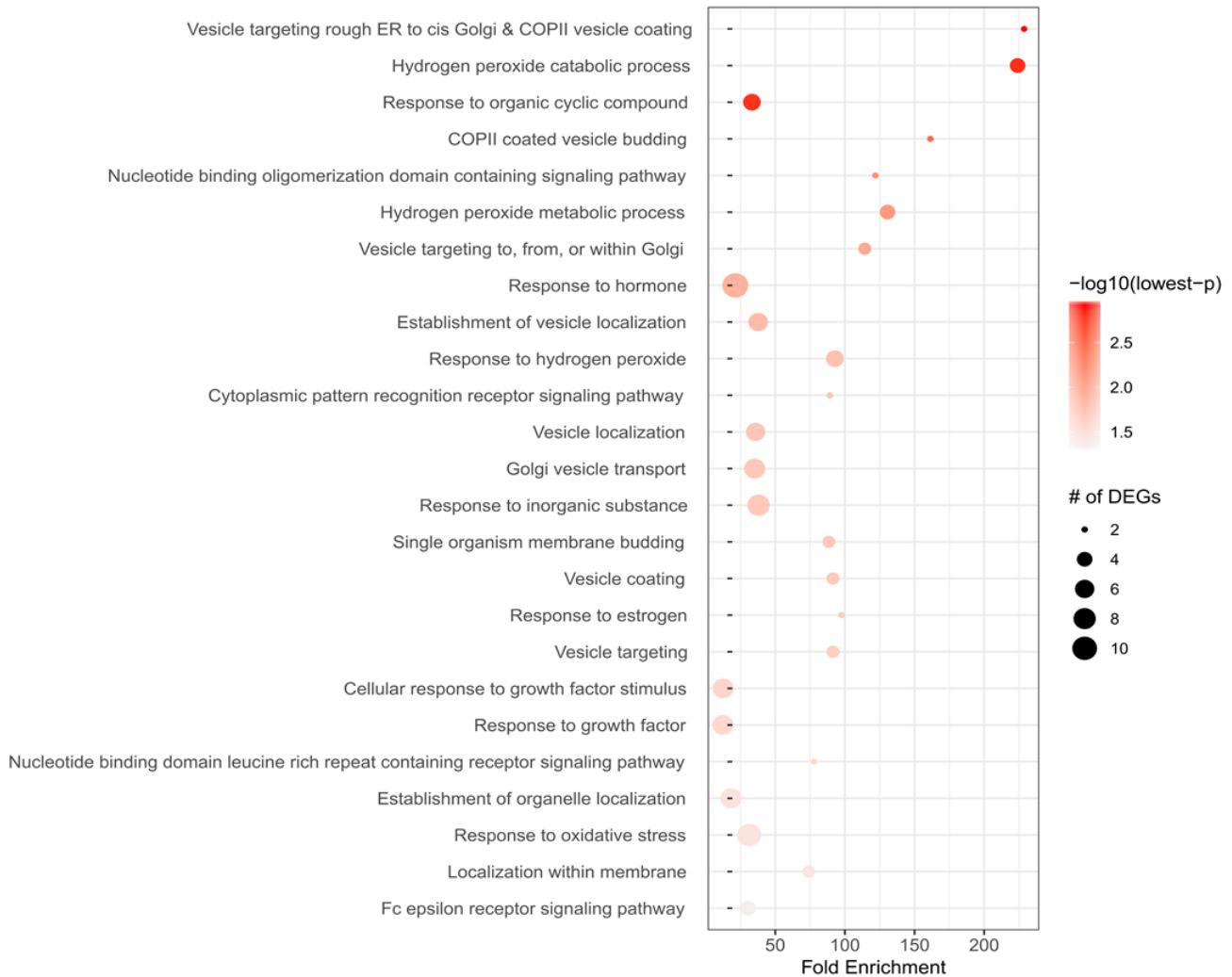


Fig. 5. Enrichment analysis by pathfindR - all pathways-GO-BP. DEGs, differentially expressed genes.

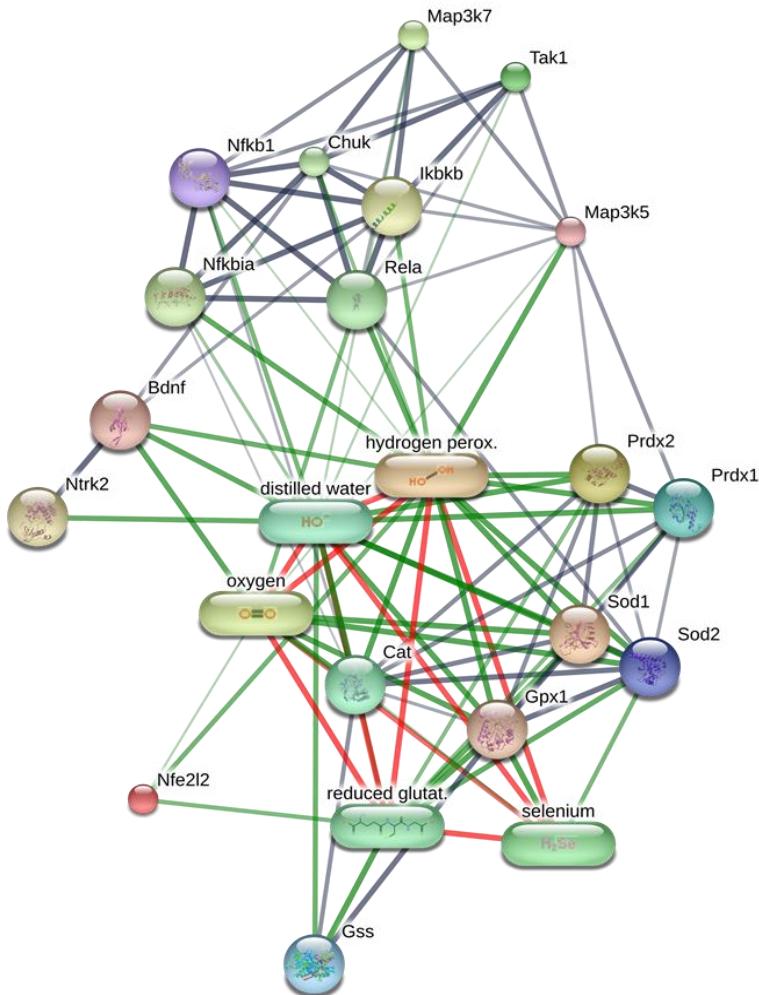


Fig. 6. Chemical association networks by STITCH - proteins involved with BDNF signaling, oxidative stress and inflammation (BDNF x H₂O₂ Combined Score = 0.891). Bdnf, brain-derived neurotrophic factor; Cat, catalase; Chuk, conserved helix-loop-helix ubiquitous kinase; Gpx, glutathione peroxidase 1; Gss, glutathione synthase; Ikbkb, inhibitor of nuclear factor kappa b kinase subunit beta; Map3k5 = Ask1, apoptosis signal-regulating kinase 1; Map3k7 = Tak1, transforming growth factor beta-activated kinase 1; Nfe2l2 = Nrf2, nuclear factor (erythroid-derived 2)-like 2; NF-κB, factor nuclear kappa b; NFκB1, nuclear factor kappa b subunit 1; Prdx1, peroxirredoxin 1; Prdx2, peroxirredoxin 2; Rela, transcription factor p65; Sod1, superoxide dismutase-1; Sod2, superoxide dismutase-2. Link to access complete results: <http://stitch.embl.de/cgi/network.pl?taskId=h07KR7iXK0cu>.

Table 1. Effects of CUMS in central TBARS, Carbonyl Protein, and TEAC.

	PC		TBARS		TEAC	
	PFC	Hippocampus	PFC	Hippocampus	PFC	Hippocampus
	0.21 ± 0.09 (n= 7)	0.25 ± 0.14 (n= 7)	0.44 ± 0.21 (n= 7)	0.33 ± 0.08 (n= 7)	15.89 ± 5.45 (n= 6)	36.09 ± 24.34 (n= 7)
Control						
	0.32 ± 0.15 (n= 9)	0.17 ± 0.07 (n= 9)	0.39 ± 0.08 (n= 9)	0.22 ± 0.09 (n= 8)	9.04 ± 4.97* (n= 9)	9.71 ± 5.58* (n= 9)
Non-anhedonic						
	0.28 ± 0.12 (n= 7)	0.16 ± 0.09 (n= 7)	0.45 ± 0.21 (n= 7)	0.28 ± 0.08 (n= 7)	7.14 ± 0.73* (n= 6)	7.95 ± 2.47* (n= 6)
Anhedonic						

All values are shown as mean ± SD. Outliers were identified by GraphPad quick calc: outlier calculator, and removed from the analyses. *p≤ 0.01 compared to the control group, according to one-way ANOVA followed by Bonferroni's post-hoc test. DNPH, 2,4-dinitrophenylhydrazine; PC, protein carbonyl; PFC, prefrontal cortex; SD, standard deviation; TBARS, thiobarbituric acid reactive substances; TEAC, Trolox equivalent antioxidant capacity.

Table 2. Enrichment Analysis of Gene Ontology Biological Process (GO-BP).

Pathway ID	Pathway description	Up-regulated	Down-regulated
GO:0042744	Hydrogen peroxide catabolic process	Prdx-1	Hba1, Hba2, Hbb
GO:0042743	Hydrogen peroxide metabolic process	Prdx-1	Hba1, Hba2, Hbb
GO:0042542	Response to hydrogen peroxide		Hba1, Hba2, Hbb Map3k5, Rela
	Cytoplasmatic pattern		
GO:0002753	recognition receptor signaling pathway		Map3k7, Rela
GO:0010035	Response to inorganic substance	mt3	Cdkn1b, Hba1, Hba2, Hbb, Map3k5, Pef1, Rela
GO:0006979	Response to oxidative stress	Cpeb2, Mt3, Prdx-1	Hba1, Hba2, Hbb, Map3k5, Naprt, Rela

Cdkn1b, cyclin-dependent kinase inhibitor 1; Hba1, hemoglobin alpha 1; Hba2, hemoglobin alpha 2; Hbb, hemoglobin beta; Map3k5 = Ask1, apoptosis signal-regulating kinase 1; Map3k7 = Tak1, transforming growth factor beta-activated kinase 1 p65; Naprt, nicotinate phosphoribosyltransferase; Pef1, penta-ef-hand domain containing 1; Prdx-1, peroxirredoxin 1; Rela, transcription factor.

PARTE III

5 CONSIDERAÇÕES FINAIS

Os achados deste estudo indicam que o CUMS esteve associado a um aumento nas concentrações de BDNF. Neurotrofinas são biomoléculas associadas à neurogênese, podendo também contribuir para a proteção das células neuronais contra o estresse oxidativo (168). O BDNF, por sua vez, pode induzir a translocação nuclear do fator Nrf2, ativando a transcrição de genes antioxidantes e resultando em efeitos protetores contra as EROs (169). Um estudo com células tipo-neuronais indicou que níveis sub-tóxicos de estresse oxidativo induzido por H₂O₂ induziram a expressão de BDNF, sustentando o envolvimento de um mecanismo sensível às EROs, subjacente aos efeitos neuroprotetores dessa neurotrofina (170).

Curiosamente, a análise dos bancos de dados de proteômica evidenciou que alguns processos biológicos enriquecidos se referem a vias relacionadas ao H₂O₂. Ao mesmo tempo, a *up-regulation* de PRDX-1 foi identificada em animais anedônicos. Considerando que a PRDX-1 é altamente sensível ao H₂O₂ e que o BDNF também pode mediar efeitos antioxidantes, sugere-se que, em conjunto, o aumento desses marcadores pode representar um mecanismo modulatório frente a níveis moderados de estresse oxidativos em animais anedônicos.

Ainda, a ausência de dano oxidativo, bem como a diminuição da TEAC e a *down-regulation* de ASK-1, TAK-1 e RELA, sugerem que o dano estrutural que poderia ser provocado por alterações no estresse oxidativo e inflamação em animais com comportamento anedônico, pode estar sendo atenuado por uma resposta compensatória via aumento de BDNF e *up-regulation* de PRDX-1. Nesse sentido, sugere-se que o CUMS possa ter induzido a geração de EROs em níveis moderados, ativando respostas antioxidantes sensíveis através via BDNF e PRDX-1 em detrimento da ativação da sinalização apoptótica e inflamatória.

O presente estudo apresenta limitações, como: I) o modelo experimental de CUMS está em fase de implementação em nosso grupo de pesquisa, sendo necessárias adaptações no

protocolo para adequá-lo às nossas investigações; II) a eutanásia e coleta de amostras biológicas foi realizada cinco dias após o último estressor, devido a realização dos testes comportamentais; III) as quantidades de amostra biológica limitaram o número de dosagens bioquímicas a serem realizadas, não tendo sido possível realizar a avaliação proteômica experimental. É importante destacar que as proteínas diferencialmente expressas, identificadas pela análise dos bancos de dados de proteômica, não foram validadas no presente estudo. Nesse sentido, a discussão dos resultados levanta hipóteses acerca de um possível mecanismo reunindo os achados experimentais e os resultados evidenciados pela bioinformática, uma vez que a patogênese da DM é altamente complexa e envolve alterações em vários processos biológicos.

É importante citar que, de forma geral, os experimentos com modelo animal avaliam fatores associados à uma janela temporal. Em contrapartida, a DM apresenta características desenvolvimentais que são moduladas em diferentes estágios da vida, frente aos mais diversos estímulos, além do componente genético associado à herdabilidade da doença. Por esse motivo, torna-se difícil mimetizar ao mesmo tempo todos os possíveis gatilhos envolvidos com o desenvolvimento da DM. Desse modo, mais investigações são necessárias para elucidar a relação entre a resposta ao estresse e os mecanismos biológicos compensatórios envolvendo o enfrentamento ao estresse crônico.

6 PERSPECTIVAS

Considerando as variações do protocolo experimental recentemente descritas na literatura, pretendemos adaptar o modelo, a fim de promover diariamente a exposição a dois estressores, com duração de 12 horas cada, durante as seis semanas de protocolo. Sugere-se que uma adaptação do protocolo de CUMS possa promover a ativação exacerbada do eixo HPA, além de induzir um estado pró-inflamatório, com comprometimento neurotrófico e alterações do estado redox em animais suscetíveis. Nesse contexto será possível estudar os mecanismos envolvidos com essas alterações já descritas e classicamente associadas à patogênese da DM, relacionando-as com o comportamento anedônico.

Ainda, pretendemos realizar a avaliação proteômica de animais submetidos ao CUMS e validar, via *Western blott*, as proteínas diferencialmente expressas nos processos biológicos enriquecidos que forem identificados. Dessa forma, além de avaliar parâmetros neurotróficos, oxidativos e inflamatórios, temos a perspectiva de estudar possíveis alterações na neurotransmissão que possam estar envolvidas com a manifestação do comportamento anedônico, incluindo o sistema serotoninérgico, glutamatérgico e seus receptores.

7 REFERÊNCIAS

1. Buckner JD, Lewis EM, Tucker RP. Mental Health Problems and Suicide Risk: The Impact of Acute Suicidal Affective Disturbance. *Arch Suicide Res.* 8 de fevereiro de 2019;1–20.
2. Cameron C, Habert J, Anand L, Furtado M. Optimizing the management of depression: primary care experience. *Psychiatry Res.* dezembro de 2014;220 Suppl 1:S45–57.
3. Lépine J-P, Briley M. The increasing burden of depression. *Neuropsychiatr Dis Treat.* 2011;7(Suppl 1):3–7.
4. Ferrari AJ, Charlson FJ, Norman RE, Patten SB, Freedman G, Murray CJL, et al. Burden of depressive disorders by country, sex, age, and year: findings from the global burden of disease study 2010. *PLoS Med.* novembro de 2013;10(11):e1001547.
5. Luppia M, Heinrich S, Angermeyer MC, König H-H, Riedel-Heller SG. Cost-of-illness studies of depression: a systematic review. *J Affect Disord.* fevereiro de 2007;98(1–2):29–43.
6. Berton O, Nestler EJ. New approaches to antidepressant drug discovery: beyond monoamines. *Nat Rev Neurosci.* fevereiro de 2006;7(2):137–51.
7. Labaka A, Goñi-Balentziaga O, Lebeña A, Pérez-Tejada J. Biological Sex Differences in Depression: A Systematic Review. *Biol Res Nurs.* 2018;20(4):383–92.
8. Riecher-Rössler A. Prospects for the classification of mental disorders in women. *Eur Psychiatry.* maio de 2010;25(4):189–96.
9. Otte C, Gold SM, Penninx BW, Pariante CM, Etkin A, Fava M, et al. Major depressive disorder. *Nat Rev Dis Primers.* 15 de 2016;2:16065.
10. Kessler RC, Berglund P, Demler O, Jin R, Koretz D, Merikangas KR, et al. The epidemiology of major depressive disorder: results from the National Comorbidity Survey Replication (NCS-R). *JAMA.* 18 de junho de 2003;289(23):3095–105.
11. Vigo D, Thornicroft G, Atun R. Estimating the true global burden of mental illness. *Lancet Psychiatry.* fevereiro de 2016;3(2):171–8.

12. Lee Y, Rosenblat JD, Lee J, Carmona NE, Subramaniapillai M, Shekotikhina M, et al. Efficacy of antidepressants on measures of workplace functioning in major depressive disorder: A systematic review. *J Affect Disord.* 2018;227:406–15.
13. World Health Organization. Depression and other common mental disorders: global health estimates [Internet]. 2017 [citado 7 de março de 2019]. Disponível em: <https://apps.who.int/iris/handle/10665/254610>
14. Sinyor M, Schaffer A, Levitt A. The Sequenced Treatment Alternatives to Relieve Depression (STAR*D) Trial: A Review. *The Canadian Journal of Psychiatry.* março de 2010;55(3):126–35.
15. Fleck MP, Horwath E. Pharmacologic management of difficult-to-treat depression in clinical practice. *Psychiatr Serv.* agosto de 2005;56(8):1005–11.
16. Gibson TB, Jing Y, Smith Carls G, Kim E, Bagalman JE, Burton WN, et al. Cost burden of treatment resistance in patients with depression. *Am J Manag Care.* maio de 2010;16(5):370–7.
17. Ivanova JI, Birnbaum HG, Kidolezi Y, Subramanian G, Khan SA, Stensland MD. Direct and indirect costs of employees with treatment-resistant and non-treatment-resistant major depressive disorder. *Curr Med Res Opin.* outubro de 2010;26(10):2475–84.
18. Olchanski N, McInnis Myers M, Halseth M, Cyr PL, Bockstedt L, Goss TF, et al. The economic burden of treatment-resistant depression. *Clin Ther.* abril de 2013;35(4):512–22.
19. Johnston KM, Powell LC, Anderson IM, Szabo S, Cline S. The burden of treatment-resistant depression: A systematic review of the economic and quality of life literature. *J Affect Disord.* 01 de 2019;242:195–210.
20. Bergfeld IO, Mantione M, Figuee M, Schuurman PR, Lok A, Denys D. Treatment-resistant depression and suicidality. *J Affect Disord.* 01 de 2018;235:362–7.
21. Braun C, Bschor T, Franklin J, Baethge C. Suicides and Suicide Attempts during Long-Term Treatment with Antidepressants: A Meta-Analysis of 29 Placebo-Controlled

- Studies Including 6,934 Patients with Major Depressive Disorder. *Psychother Psychosom.* 2016;85(3):171–9.
22. Esperidião-Antonio V, Majeski-Colombo M, Toledo-Monteverde D, Moraes-Martins G, Fernandes JJ, Assis MB de, et al. Neurobiology of the emotions. *Archives of Clinical Psychiatry (São Paulo).* 2008;35(2):55–65.
 23. Kuhn M, Popovic A, Pezawas L. Neuroplasticity and memory formation in major depressive disorder: an imaging genetics perspective on serotonin and BDNF. *Restor Neurol Neurosci.* 2014;32(1):25–49.
 24. Furczyk K, Schutová B, Michel TM, Thome J, Büttner A. The neurobiology of suicide - A Review of post-mortem studies. *J Mol Psychiatry [Internet].* 23 de abril de 2013 [citado 12 de março de 2019];1(1). Disponível em: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4223890/>
 25. Manji HK, Drevets WC, Charney DS. The cellular neurobiology of depression. *Nat Med.* maio de 2001;7(5):541–7.
 26. Nestler EJ, Barrot M, DiLeone RJ, Eisch AJ, Gold SJ, Monteggia LM. Neurobiology of depression. *Neuron.* 28 de março de 2002;34(1):13–25.
 27. Campbell S, Marriott M, Nahmias C, MacQueen GM. Lower hippocampal volume in patients suffering from depression: a meta-analysis. *Am J Psychiatry.* abril de 2004;161(4):598–607.
 28. Cotter D, Mackay D, Landau S, Kerwin R, Everall I. Reduced glial cell density and neuronal size in the anterior cingulate cortex in major depressive disorder. *Arch Gen Psychiatry.* junho de 2001;58(6):545–53.
 29. Cotter D, Mackay D, Chana G, Beasley C, Landau S, Everall IP. Reduced neuronal size and glial cell density in area 9 of the dorsolateral prefrontal cortex in subjects with major depressive disorder. *Cereb Cortex.* abril de 2002;12(4):386–94.
 30. Geng H, Wu F, Kong L, Tang Y, Zhou Q, Chang M, et al. Disrupted Structural and Functional Connectivity in Prefrontal-Hippocampus Circuitry in First-Episode

- Medication-Naïve Adolescent Depression. PLOS ONE. 10 de fevereiro de 2016;11(2):e0148345.
31. Liu W, Ge T, Leng Y, Pan Z, Fan J, Yang W, et al. The Role of Neural Plasticity in Depression: From Hippocampus to Prefrontal Cortex. *Neural Plast [Internet]*. 2017 [citado 12 de março de 2019];2017. Disponível em: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5299163/>
 32. Saltiel PF, Silvershein DI. Major depressive disorder: mechanism-based prescribing for personalized medicine. *Neuropsychiatr Dis Treat*. 2015;11:875–88.
 33. Davis JM. Theories of biological etiology of affective disorders. *Int Rev Neurobiol*. 1970;12:145–75.
 34. Coppen A. The biochemistry of affective disorders. *Br J Psychiatry*. novembro de 1967;113(504):1237–64.
 35. Morissette DA, Stahl SM. Modulating the serotonin system in the treatment of major depressive disorder. *CNS Spectr*. dezembro de 2014;19 Suppl 1:57–67; quiz 54–7, 68.
 36. Lapin IP, Oxenkrug GF. Intensification of the central serotonergic processes as a possible determinant of the thymoleptic effect. *Lancet*. 18 de janeiro de 1969;1(7586):132–6.
 37. Rush AJ, Trivedi MH, Wisniewski SR, Nierenberg AA, Stewart JW, Warden D, et al. Acute and longer-term outcomes in depressed outpatients requiring one or several treatment steps: a STAR*D report. *Am J Psychiatry*. novembro de 2006;163(11):1905–17.
 38. Trivedi MH, Fava M, Wisniewski SR, Thase ME, Quitkin F, Warden D, et al. Medication augmentation after the failure of SSRIs for depression. *N Engl J Med*. 23 de março de 2006;354(12):1243–52.
 39. Bădescu S, Tătaru C, Kobylinska L, Georgescu E, Zahiu D, Zăgrean A, et al. The association between Diabetes mellitus and Depression. *J Med Life*. 2016;9(2):120–5.

40. Luppino FS, Wit LM de, Bouvy PF, Stijnen T, Cuijpers P, Penninx BWJH, et al. Overweight, Obesity, and Depression: A Systematic Review and Meta-analysis of Longitudinal Studies. *Arch Gen Psychiatry*. 1º de março de 2010;67(3):220–9.
41. Varan Ö, Babaoğlu H, Göker B. Associations between Depressive Disorders and Inflammatory Rheumatic Diseases. *Curr Top Med Chem*. 2018;18(16):1395–401.
42. Raedler TJ. Inflammatory mechanisms in major depressive disorder. *Curr Opin Psychiatry*. novembro de 2011;24(6):519–25.
43. Raison CL, Lowry CA, Rook GAW. Inflammation, sanitation, and consternation: loss of contact with coevolved, tolerogenic microorganisms and the pathophysiology and treatment of major depression. *Arch Gen Psychiatry*. dezembro de 2010;67(12):1211–24.
44. Krishnadas R, Cavanagh J. Depression: an inflammatory illness? *J Neurol Neurosurg Psychiatry*. maio de 2012;83(5):495–502.
45. Lichtblau N, Schmidt FM, Schumann R, Kirkby KC, Himmerich H. Cytokines as biomarkers in depressive disorder: current standing and prospects. *Int Rev Psychiatry*. outubro de 2013;25(5):592–603.
46. Dowlati Y, Herrmann N, Swardfager W, Liu H, Sham L, Reim EK, et al. A meta-analysis of cytokines in major depression. *Biol Psychiatry*. 1º de março de 2010;67(5):446–57.
47. Howren MB, Lamkin DM, Suls J. Associations of depression with C-reactive protein, IL-1, and IL-6: a meta-analysis. *Psychosom Med*. fevereiro de 2009;71(2):171–86.
48. Gimeno D, Kivimäki M, Brunner EJ, Elovainio M, De Vogli R, Steptoe A, et al. Associations of C-reactive protein and interleukin-6 with cognitive symptoms of depression: 12-year follow-up of the Whitehall II study. *Psychol Med*. março de 2009;39(3):413–23.
49. Banks WA. Blood–Brain Barrier Transport of Cytokines. In: *NeuroImmune Biology* [Internet]. Elsevier; 2008 [citado 12 de março de 2019]. p. 93–107. (Cytokines and the

Brain; vol. 6). Disponível em:
<http://www.sciencedirect.com/science/article/pii/S1567744307100065>

50. Steinberg BE, Silverman HA, Robbiati S, Gunasekaran MK, Tsaava T, Battinelli E, et al. Cytokine-specific Neurograms in the Sensory Vagus Nerve. *Bioelectron Med.* 2016;3:7–17.
51. Quevedo FK| II| J. Bases Biológicas dos Transtornos Psiquiátricos: Uma Abordagem Translacional. Artmed Editora; 2009. 332 p.
52. Heneka MT, Kummer MP, Latz E. Innate immune activation in neurodegenerative disease. *Nat Rev Immunol.* julho de 2014;14(7):463–77.
53. Moon ML, McNeil LK, Freund GG. Macrophages make me sick: How macrophage activation states influence sickness behavior. *Psychoneuroendocrinology.* novembro de 2011;36(10):1431–40.
54. Felger JC, Lotrich FE. Inflammatory cytokines in depression: neurobiological mechanisms and therapeutic implications. *Neuroscience.* 29 de agosto de 2013;246:199–229.
55. Miller AH, Raison CL. The role of inflammation in depression: from evolutionary imperative to modern treatment target. *Nat Rev Immunol.* janeiro de 2016;16(1):22–34.
56. Cooper JA, Arulpragasam AR, Treadway MT. Anhedonia in depression: biological mechanisms and computational models. *Curr Opin Behav Sci.* agosto de 2018;22:128–35.
57. Müller N, Schwarz MJ. A psychoneuroimmunological perspective to Emil Kraepelins dichotomy: schizophrenia and major depression as inflammatory CNS disorders. *Eur Arch Psychiatry Clin Neurosci.* junho de 2008;258 Suppl 2:97–106.
58. Schwieler L, Erhardt S, Nilsson L, Linderholm K, Engberg G. Effects of COX-1 and COX-2 inhibitors on the firing of rat midbrain dopaminergic neurons--possible involvement of endogenous kynurenic acid. *Synapse.* abril de 2006;59(5):290–8.
59. Banasr M, Dwyer JM, Duman RS. Cell atrophy and loss in depression: reversal by antidepressant treatment. *Curr Opin Cell Biol.* dezembro de 2011;23(6):730–7.

60. Walker AK, Budac DP, Bisulco S, Lee AW, Smith RA, Beenders B, et al. NMDA receptor blockade by ketamine abrogates lipopolysaccharide-induced depressive-like behavior in C57BL/6J mice. *Neuropsychopharmacology*. agosto de 2013;38(9):1609–16.
61. O'Connor JC, Lawson MA, André C, Moreau M, Lestage J, Castanon N, et al. Lipopolysaccharide-induced depressive-like behavior is mediated by indoleamine 2,3-dioxygenase activation in mice. *Mol Psychiatry*. maio de 2009;14(5):511–22.
62. Miller AH, Maletic V, Raison CL. Inflammation and its discontents: the role of cytokines in the pathophysiology of major depression. *Biol Psychiatry*. 1º de maio de 2009;65(9):732–41.
63. Belmaker RH, Agam G. Major depressive disorder. *N Engl J Med*. 3 de janeiro de 2008;358(1):55–68.
64. Liu Y-Z, Wang Y-X, Jiang C-L. Inflammation: The Common Pathway of Stress-Related Diseases. *Front Hum Neurosci*. 2017;11:316.
65. Pariante CM, Miller AH. Glucocorticoid receptors in major depression: relevance to pathophysiology and treatment. *Biol Psychiatry*. 1º de março de 2001;49(5):391–404.
66. Nicolaides NC, Kyratzi E, Lamprokostopoulou A, Chrousos GP, Charmandari E. Stress, the stress system and the role of glucocorticoids. *Neuroimmunomodulation*. 2015;22(1–2):6–19.
67. Hasler G, Drevets WC, Manji HK, Charney DS. Discovering endophenotypes for major depression. *Neuropsychopharmacology*. outubro de 2004;29(10):1765–81.
68. Ising M, Horstmann S, Kloiber S, Lucae S, Binder EB, Kern N, et al. Combined dexamethasone/corticotropin releasing hormone test predicts treatment response in major depression - a potential biomarker? *Biol Psychiatry*. 1º de julho de 2007;62(1):47–54.
69. Pariante CM, Lightman SL. The HPA axis in major depression: classical theories and new developments. *Trends Neurosci*. setembro de 2008;31(9):464–8.

70. Cohen S, Janicki-Deverts D, Doyle WJ, Miller GE, Frank E, Rabin BS, et al. Chronic stress, glucocorticoid receptor resistance, inflammation, and disease risk. *Proc Natl Acad Sci U S A.* 17 de abril de 2012;109(16):5995–9.
71. Miller GE, Chen E, Sze J, Marin T, Arevalo JMG, Doll R, et al. A functional genomic fingerprint of chronic stress in humans: blunted glucocorticoid and increased NF-kappaB signaling. *Biol Psychiatry.* 15 de agosto de 2008;64(4):266–72.
72. Pace TWW, Miller AH. Cytokines and glucocorticoid receptor signaling. Relevance to major depression. *Ann N Y Acad Sci.* outubro de 2009;1179:86–105.
73. Carvalho AL, Caldeira MV, Santos SD, Duarte CB. Role of the brain-derived neurotrophic factor at glutamatergic synapses. *Br J Pharmacol.* março de 2008;153(Suppl 1):S310–24.
74. Carroll BJ. Clomipramine and glucocorticoid receptor function. *Neuropsychopharmacology.* agosto de 2009;34(9):2192–3; author reply 2194-2195.
75. Halliwell B. Free radicals and antioxidants: updating a personal view. *Nutr Rev.* maio de 2012;70(5):257–65.
76. Halliwell B. Cell culture, oxidative stress, and antioxidants: avoiding pitfalls. *Biomed J.* junho de 2014;37(3):99–105.
77. Vaváková M, Ďuračková Z, Trebatická J. Markers of Oxidative Stress and Neuroprogression in Depression Disorder. *Oxid Med Cell Longev.* 2015;2015:898393.
78. Maes M, Mihaylova I, Kubera M, Uytterhoeven M, Vrydaghs N, Bosmans E. Lower whole blood glutathione peroxidase (GPX) activity in depression, but not in myalgic encephalomyelitis / chronic fatigue syndrome: another pathway that may be associated with coronary artery disease and neuroprogression in depression. *Neuro Endocrinol Lett.* 2011;32(2):133–40.
79. Dalle-Donne I, Aldini G, Carini M, Colombo R, Rossi R, Milzani A. Protein carbonylation, cellular dysfunction, and disease progression. *J Cell Mol Med.* junho de 2006;10(2):389–406.

80. Salim S. Oxidative Stress and Psychological Disorders. *Curr Neuropharmacol.* março de 2014;12(2):140–7.
81. Gaschler MM, Stockwell BR. Lipid peroxidation in cell death. *Biochem Biophys Res Commun.* 15 de 2017;482(3):419–25.
82. Islam MR, Islam MR, Ahmed I, Moktadir AA, Nahar Z, Islam MS, et al. Elevated serum levels of malondialdehyde and cortisol are associated with major depressive disorder: A case-control study. *SAGE Open Med.* 2018;6:2050312118773953.
83. Khajehnasiri F, Mortazavi SB, Allameh A, Akhondzadeh S, Hashemi H. Total antioxidant capacity and malondialdehyde in depressive rotational shift workers. *J Environ Public Health.* 2013;2013:150693.
84. Mazereeuw G, Herrmann N, Andreazza AC, Khan MM, Lanctôt KL. A meta-analysis of lipid peroxidation markers in major depression. *Neuropsychiatr Dis Treat.* 2015;11:2479–91.
85. Black CN, Bot M, Scheffer PG, Cuijpers P, Penninx BWJH. Is depression associated with increased oxidative stress? A systematic review and meta-analysis. *Psychoneuroendocrinology.* janeiro de 2015;51:164–75.
86. Ighodaro OM, Akinloye OA. First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): Their fundamental role in the entire antioxidant defence grid. *Alexandria Journal of Medicine.* 1º de dezembro de 2018;54(4):287–93.
87. Chelikani P, Fita I, Loewen PC. Diversity of structures and properties among catalases. *Cell Mol Life Sci.* janeiro de 2004;61(2):192–208.
88. Maurya PK, Noto C, Rizzo LB, Rios AC, Nunes SOV, Barbosa DS, et al. The role of oxidative and nitrosative stress in accelerated aging and major depressive disorder. *Prog Neuropsychopharmacol Biol Psychiatry.* 4 de fevereiro de 2016;65:134–44.
89. Lopresti AL, Maker GL, Hood SD, Drummond PD. A review of peripheral biomarkers in major depression: the potential of inflammatory and oxidative stress biomarkers. *Prog Neuropsychopharmacol Biol Psychiatry.* 3 de janeiro de 2014;48:102–11.

90. Stefanescu C, Ciobica A. The relevance of oxidative stress status in first episode and recurrent depression. *J Affect Disord.* 20 de dezembro de 2012;143(1–3):34–8.
91. Rybka J, Kędziora-Kornatowska K, Banaś-Leżańska P, Majsterek I, Carvalho LA, Cattaneo A, et al. Interplay between the pro-oxidant and antioxidant systems and proinflammatory cytokine levels, in relation to iron metabolism and the erythron in depression. *Free Radic Biol Med.* outubro de 2013;63:187–94.
92. Ozcan ME, Gulec M, Ozerol E, Polat R, Akyol O. Antioxidant enzyme activities and oxidative stress in affective disorders. *Int Clin Psychopharmacol.* março de 2004;19(2):89–95.
93. Gałecki P, Szemraj J, Bieńkiewicz M, Florkowski A, Gałecka E. Lipid peroxidation and antioxidant protection in patients during acute depressive episodes and in remission after fluoxetine treatment. *Pharmacol Rep.* junho de 2009;61(3):436–47.
94. Kodydková J, Vávrová L, Zeman M, Jirák R, Macásek J, Stanková B, et al. Antioxidative enzymes and increased oxidative stress in depressive women. *Clin Biochem.* setembro de 2009;42(13–14):1368–74.
95. Tsai M-C, Huang T-L. Increased activities of both superoxide dismutase and catalase were indicators of acute depressive episodes in patients with major depressive disorder. *Psychiatry Res.* 30 de janeiro de 2016;235:38–42.
96. Antunes F, Brito PM. Quantitative biology of hydrogen peroxide signaling. *Redox Biology.* outubro de 2017;13:1.
97. Sies H, Berndt C, Jones DP. Oxidative Stress. *Annual Review of Biochemistry.* 2017;86(1):715–48.
98. Won H, Lim S, Jang M, Kim Y, Rashid MA, Jyothi KR, et al. Peroxiredoxin-2 upregulated by NF-κB attenuates oxidative stress during the differentiation of muscle-derived C2C12 cells. *Antioxid Redox Signal.* 1º de fevereiro de 2012;16(3):245–61.
99. Kim S-U, Hwang CN, Sun H-N, Jin M-H, Han Y-H, Lee H, et al. Peroxiredoxin I is an indicator of microglia activation and protects against hydrogen peroxide-mediated microglial death. *Biol Pharm Bull.* maio de 2008;31(5):820–5.

100. Skynner HA, Amos DP, Murray F, Salim K, Knowles MR, Munoz-Sanjuan I, et al. Proteomic analysis identifies alterations in cellular morphology and cell death pathways in mouse brain after chronic corticosterone treatment. *Brain Res.* 2 de agosto de 2006;1102(1):12–26.
101. Gutteridge JMC, Halliwell B. Antioxidants: Molecules, medicines, and myths. *Biochem Biophys Res Commun.* 19 de março de 2010;393(4):561–4.
102. Dringen R. Metabolism and functions of glutathione in brain. *Prog Neurobiol.* dezembro de 2000;62(6):649–71.
103. Gu F, Chauhan V, Chauhan A. Glutathione redox imbalance in brain disorders. *Curr Opin Clin Nutr Metab Care.* janeiro de 2015;18(1):89–95.
104. Gawryluk JW, Wang J-F, Andreazza AC, Shao L, Young LT. Decreased levels of glutathione, the major brain antioxidant, in post-mortem prefrontal cortex from patients with psychiatric disorders. *Int J Neuropsychopharmacol.* fevereiro de 2011;14(1):123–30.
105. Erel O. A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. *Clin Biochem.* abril de 2004;37(4):277–85.
106. Bartosz G. Non-enzymatic antioxidant capacity assays: Limitations of use in biomedicine. *Free Radic Res.* julho de 2010;44(7):711–20.
107. Cumurcu BE, Ozyurt H, Etikan I, Demir S, Karlidag R. Total antioxidant capacity and total oxidant status in patients with major depression: Impact of antidepressant treatment. *Psychiatry and Clinical Neurosciences.* 2009;63(5):639–45.
108. Liu T, Zhong S, Liao X, Chen J, He T, Lai S, et al. A Meta-Analysis of Oxidative Stress Markers in Depression. *PLoS ONE.* 2015;10(10):e0138904.
109. Michel TM, Pülschen D, Thome J. The role of oxidative stress in depressive disorders. *Curr Pharm Des.* 2012;18(36):5890–9.

110. Sheline YI, Disabato BM, Hranilovich J, Morris C, D'Angelo G, Pieper C, et al. Treatment course with antidepressant therapy in late-life depression. *Am J Psychiatry*. novembro de 2012;169(11):1185–93.
111. Lindqvist D, Mueller S, Mellon SH, Su Y, Epel ES, Reus VI, et al. Peripheral antioxidant markers are associated with total hippocampal and CA3/dentate gyrus volume in MDD and healthy controls-preliminary findings. *Psychiatry Res*. 30 de dezembro de 2014;224(3):168–74.
112. Lindqvist D, Dhabhar FS, James SJ, Hough CM, Jain FA, Bersani FS, et al. Oxidative stress, inflammation and treatment response in major depression. *Psychoneuroendocrinology*. 2017;76:197–205.
113. Xu Y, Wang C, Klabnik JJ, O'Donnell JM. Novel therapeutic targets in depression and anxiety: antioxidants as a candidate treatment. *Curr Neuropharmacol*. março de 2014;12(2):108–19.
114. Cimen B, Gumus CB, Cetin I, Ozsoy S, Aydin M, Cimen L. The Effects of Escitalopram Treatment on Oxidative/ Antioxidative Parameters in Patients with Depression. *Klinik Psikofarmakoloji Bülteni-Bulletin of Clinical Psychopharmacology*. setembro de 2015;25(3):272–9.
115. Zafir A, Ara A, Banu N. Invivo antioxidant status: a putative target of antidepressant action. *Prog Neuropsychopharmacol Biol Psychiatry*. 17 de março de 2009;33(2):220–8.
116. Abdel-Wahab BA, Salama RH. Venlafaxine protects against stress-induced oxidative DNA damage in hippocampus during antidepressant testing in mice. *Pharmacol Biochem Behav*. novembro de 2011;100(1):59–65.
117. Kolla N, Wei Z, Richardson JS, Li X-M. Amitriptyline and fluoxetine protect PC12 cells from cell death induced by hydrogen peroxide. *J Psychiatry Neurosci*. maio de 2005;30(3):196–201.
118. Chen Y, Ouyang J, Liu S, Zhang S, Chen P, Jiang T. The Role of Cytokines in the Peripheral Blood of Major Depressive Patients. *Clin Lab*. 1º de julho de 2017;63(7):1207–12.

119. Bakunina N, Pariante CM, Zunszain PA. Immune mechanisms linked to depression via oxidative stress and neuroprogression. *Immunology*. 2015;144(3):365–73.
120. Mellon SH, Wolkowitz OM, Schonemann MD, Epel ES, Rosser R, Burke HB, et al. Alterations in leukocyte transcriptional control pathway activity associated with major depressive disorder and antidepressant treatment. *Transl Psychiatry*. 24 de 2016;6:e821.
121. Bouvier E, Brouillard F, Molet J, Claverie D, Cabungcal J-H, Cresto N, et al. Nrf2-dependent persistent oxidative stress results in stress-induced vulnerability to depression. *Mol Psychiatry*. dezembro de 2017;22(12):1701–13.
122. Shin H-M, Kim M-H, Kim BH, Jung S-H, Kim YS, Park HJ, et al. Inhibitory action of novel aromatic diamine compound on lipopolysaccharide-induced nuclear translocation of NF-kappaB without affecting IkappaB degradation. *FEBS Lett.* 30 de julho de 2004;571(1–3):50–4.
123. Kishore N, Sommers C, Mathialagan S, Guzova J, Yao M, Hauser S, et al. A selective IKK-2 inhibitor blocks NF-kappa B-dependent gene expression in interleukin-1 beta-stimulated synovial fibroblasts. *J Biol Chem*. 29 de agosto de 2003;278(35):32861–71.
124. LaPlant Q, Chakravarty S, Vialou V, Mukherjee S, Koo JW, Kalahasti G, et al. Role of nuclear factor kappaB in ovarian hormone-mediated stress hypersensitivity in female mice. *Biol Psychiatry*. 15 de maio de 2009;65(10):874–80.
125. Munhoz CD, Lepsch LB, Kawamoto EM, Malta MB, Lima L de S, Avellar MCW, et al. Chronic unpredictable stress exacerbates lipopolysaccharide-induced activation of nuclear factor-kappaB in the frontal cortex and hippocampus via glucocorticoid secretion. *J Neurosci*. 5 de abril de 2006;26(14):3813–20.
126. Madrigal JL, Moro MA, Lizasoain I, Lorenzo P, Castrillo A, Boscá L, et al. Inducible nitric oxide synthase expression in brain cortex after acute restraint stress is regulated by nuclear factor kappaB-mediated mechanisms. *J Neurochem*. janeiro de 2001;76(2):532–8.
127. Koo JW, Russo SJ, Ferguson D, Nestler EJ, Duman RS. Nuclear factor- κ B is a critical mediator of stress-impaired neurogenesis and depressive behavior. *Proc Natl Acad Sci U S A*. 9 de fevereiro de 2010;107(6):2669–74.

128. Strawbridge R, Arnone D, Danese A, Papadopoulos A, Herane Vives A, Cleare AJ. Inflammation and clinical response to treatment in depression: A meta-analysis. *Eur Neuropsychopharmacol.* outubro de 2015;25(10):1532–43.
129. Carvalho LA, Torre JP, Papadopoulos AS, Poon L, Juruena MF, Markopoulou K, et al. Lack of clinical therapeutic benefit of antidepressants is associated overall activation of the inflammatory system. *J Affect Disord.* 15 de maio de 2013;148(1):136–40.
130. Kowiański P, Lietzau G, Czuba E, Waśkow M, Steliga A, Moryś J. BDNF: A Key Factor with Multipotent Impact on Brain Signaling and Synaptic Plasticity. *Cell Mol Neurobiol.* abril de 2018;38(3):579–93.
131. Kunugi H, Hori H, Adachi N, Numakawa T. Interface between hypothalamic-pituitary-adrenal axis and brain-derived neurotrophic factor in depression. *Psychiatry and Clinical Neurosciences.* 2010;64(5):447–59.
132. Post RM. Role of BDNF in bipolar and unipolar disorder: clinical and theoretical implications. *J Psychiatr Res.* dezembro de 2007;41(12):979–90.
133. Caviedes A, Lafourcade C, Soto C, Wyneken U. BDNF/NF-κB Signaling in the Neurobiology of Depression. *Curr Pharm Des.* 2017;23(21):3154–63.
134. Chen P, Jiang T, Ouyang J, Cui Y, Chen Y. Epigenetic programming of diverse glucocorticoid response and inflammatory/immune-mediated disease. *Med Hypotheses.* novembro de 2009;73(5):657–8.
135. Hjemdal O, Vogel PA, Solem S, Hagen K, Stiles TC. The relationship between resilience and levels of anxiety, depression, and obsessive-compulsive symptoms in adolescents. *Clin Psychol Psychother.* agosto de 2011;18(4):314–21.
136. Wang M, Perova Z, Arenkiel BR, Li B. Synaptic modifications in the medial prefrontal cortex in susceptibility and resilience to stress. *J Neurosci.* 28 de maio de 2014;34(22):7485–92.
137. García-León MÁ, Pérez-Mármol JM, González-Pérez R, García-Ríos MDC, Peralta-Ramírez MI. Relationship between resilience and stress: Perceived stress, stressful life

- events, HPA axis response during a stressful task and hair cortisol. *Physiol Behav.* 1º de abril de 2019;202:87–93.
138. Krishnan V, Nestler EJ. The molecular neurobiology of depression. *Nature.* 16 de outubro de 2008;455(7215):894–902.
 139. Henningsen K, Palmfeldt J, Christiansen S, Baiges I, Bak S, Jensen ON, et al. Candidate hippocampal biomarkers of susceptibility and resilience to stress in a rat model of depression. *Mol Cell Proteomics.* julho de 2012;11(7):M111.016428.
 140. Der-Avakian A, Barnes SA, Markou A, Pizzagalli DA. Translational Assessment of Reward and Motivational Deficits in Psychiatric Disorders. *Curr Top Behav Neurosci.* 2016;28:231–62.
 141. Scheggi S, De Montis MG, Gambarana C. Making Sense of Rodent Models of Anhedonia. *Int J Neuropsychopharmacol.* 1º de novembro de 2018;21(11):1049–65.
 142. Akimoto H, Oshima S, Sugiyama T, Negishi A, Nemoto T, Kobayashi D. Changes in brain metabolites related to stress resilience: Metabolomic analysis of the hippocampus in a rat model of depression. *Behav Brain Res.* 01 de 2019;359:342–52.
 143. Rossetti AC, Papp M, Gruca P, Paladini MS, Racagni G, Riva MA, et al. Stress-induced anhedonia is associated with the activation of the inflammatory system in the rat brain: Restorative effect of pharmacological intervention. *Pharmacol Res.* janeiro de 2016;103:1–12.
 144. Strekalova T, Couch Y, Kholod N, Boyks M, Malin D, Leprince P, et al. Update in the methodology of the chronic stress paradigm: internal control matters. *Behav Brain Funct.* 27 de abril de 2011;7:9.
 145. Kolasa M, Faron-Górecka A, Kuśmider M, Szafran-Pilch K, Solich J, Żurawek D, et al. Differential stress response in rats subjected to chronic mild stress is accompanied by changes in CRH-family gene expression at the pituitary level. *Peptides.* novembro de 2014;61:98–106.

146. Bergström A, Jayatissa MN, Thykjaer T, Wiborg O. Molecular pathways associated with stress resilience and drug resistance in the chronic mild stress rat model of depression: a gene expression study. *J Mol Neurosci.* 2007;33(2):201–15.
147. Henningsen K, Andreasen JT, Bouzinova EV, Jayatissa MN, Jensen MS, Redrobe JP, et al. Cognitive deficits in the rat chronic mild stress model for depression: relation to anhedonic-like responses. *Behav Brain Res.* 2 de março de 2009;198(1):136–41.
148. Wang C, Wu H-M, Jing X-R, Meng Q, Liu B, Zhang H, et al. Oxidative parameters in the rat brain of chronic mild stress model for depression: relation to anhedonia-like responses. *J Membr Biol.* novembro de 2012;245(11):675–81.
149. Nestler EJ, Hyman SE. Animal Models of Neuropsychiatric Disorders. *Nat Neurosci.* outubro de 2010;13(10):1161–9.
150. Belzung C, Lemoine M. Criteria of validity for animal models of psychiatric disorders: focus on anxiety disorders and depression. *Biol Mood Anxiety Disord.* 7 de novembro de 2011;1:9.
151. Katz RJ. Animal model of depression: pharmacological sensitivity of a hedonic deficit. *Pharmacol Biochem Behav.* junho de 1982;16(6):965–8.
152. Willner P, Towell A, Sampson D, Sophokleous S, Muscat R. Reduction of sucrose preference by chronic unpredictable mild stress, and its restoration by a tricyclic antidepressant. *Psychopharmacology (Berl).* 1987;93(3):358–64.
153. Willner P. Reliability of the chronic mild stress model of depression: A user survey. *Neurobiol Stress.* 22 de agosto de 2016;6:68–77.
154. Willner P. The chronic mild stress (CMS) model of depression: History, evaluation and usage. *Neurobiol Stress.* fevereiro de 2017;6:78–93.
155. Yang X-H, Song S-Q, Xu Y. Resveratrol ameliorates chronic unpredictable mild stress-induced depression-like behavior: involvement of the HPA axis, inflammatory markers, BDNF, and Wnt/β-catenin pathway in rats. *Neuropsychiatr Dis Treat.* 2017;13:2727–36.

156. Liu W, Sheng H, Xu Y, Liu Y, Lu J, Ni X. Swimming exercise ameliorates depression-like behavior in chronically stressed rats: relevant to proinflammatory cytokines and IDO activation. *Behav Brain Res.* 1º de abril de 2013;242:110–6.
157. Xie Z-M, Wang X-M, Xu N, Wang J, Pan W, Tang X-H, et al. Alterations in the inflammatory cytokines and brain-derived neurotrophic factor contribute to depression-like phenotype after spared nerve injury: improvement by ketamine. *Sci Rep.* 09 de 2017;7(1):3124.
158. Koo JW, Duman RS. IL-1beta is an essential mediator of the antineurogenic and anhedonic effects of stress. *Proc Natl Acad Sci USA.* 15 de janeiro de 2008;105(2):751–6.
159. Lucca G, Comim CM, Valvassori SS, Réus GZ, Vuolo F, Petronilho F, et al. Effects of chronic mild stress on the oxidative parameters in the rat brain. *Neurochem Int.* junho de 2009;54(5–6):358–62.
160. Maluach AM, Misquitta KA, Prevot TD, Fee C, Sibille E, Banasr M, et al. Increased Neuronal DNA/RNA Oxidation in the Frontal Cortex of Mice Subjected to Unpredictable Chronic Mild Stress. *Chronic Stress (Thousand Oaks).* dezembro de 2017;1.
161. Moretti M, Colla A, de Oliveira Balen G, dos Santos DB, Budni J, de Freitas AE, et al. Ascorbic acid treatment, similarly to fluoxetine, reverses depressive-like behavior and brain oxidative damage induced by chronic unpredictable stress. *Journal of Psychiatric Research.* 1º de março de 2012;46(3):331–40.
162. Xu H-B, Zhang R-F, Luo D, Zhou Y, Wang Y, Fang L, et al. Comparative proteomic analysis of plasma from major depressive patients: identification of proteins associated with lipid metabolism and immunoregulation. *Int J Neuropsychopharmacol.* 1º de novembro de 2012;15(10):1413–25.
163. Yang Y, Yang D, Tang G, Zhou C, Cheng K, Zhou J, et al. Proteomics reveals energy and glutathione metabolic dysregulation in the prefrontal cortex of a rat model of depression. *Neuroscience.* 5 de setembro de 2013;247:191–200.

164. Zhang Y, Yuan S, Pu J, Yang L, Zhou X, Liu L, et al. Integrated Metabolomics and Proteomics Analysis of Hippocampus in a Rat Model of Depression. *Neuroscience*. 10 de 2018;371:207–20.
165. Mu J, Xie P, Yang Z-S, Yang D-L, Lv F-J, Luo T-Y, et al. Neurogenesis and major depression: implications from proteomic analyses of hippocampal proteins in a rat depression model. *Neurosci Lett*. 18 de abril de 2007;416(3):252–6.
166. Zhou M, Liu Z, Yu J, Li S, Tang M, Zeng L, et al. Quantitative Proteomic Analysis Reveals Synaptic Dysfunction in the Amygdala of Rats Susceptible to Chronic Mild Stress. *Neuroscience*. 15 de 2018;376:24–39.
167. Horowitz MA, Zunszain PA. Neuroimmune and neuroendocrine abnormalities in depression: two sides of the same coin. *Ann N Y Acad Sci*. setembro de 2015;1351:68–79.
168. Ichim G, Tauszig-Delamasure S, Mehlen P. Neurotrophins and cell death. *Exp Cell Res*. 1º de julho de 2012;318(11):1221–8.
169. Bruna B, Lobos P, Herrera-Molina R, Hidalgo C, Paula-Lima A, Adasme T. The signaling pathways underlying BDNF-induced Nrf2 hippocampal nuclear translocation involve ROS, RyR-Mediated Ca²⁺ signals, ERK and PI3K. *Biochem Biophys Res Commun*. 20 de outubro de 2018;505(1):201–7.
170. Ogura Y, Sato K, Kawashima K-I, Kobayashi N, Imura S, Fujino K, et al. Subtoxic levels of hydrogen peroxide induce brain-derived neurotrophic factor expression to protect PC12 cells. *BMC Res Notes*. 25 de novembro de 2014;7(1):840.

ANEXO A

Carta de aprovação do projeto sob número 150353 pela Comissão de Ética no Uso de Animais do Hospital de Clínicas de Porto Alegre (CEUA/HCPA).



**GRUPO DE PESQUISA E PÓS GRADUAÇÃO
COMISSÃO DE ÉTICA NO USO DE ANIMAIS**



Certificamos que o projeto abaixo, que envolve a produção, manutenção ou utilização de animais pertencentes ao filo Chordata, subfilo Vertebrata (exceto humanos), para fins de pesquisa científica, encontra-se de acordo com os preceitos da Lei nº 11.794, de 8 de outubro de 2008, do Decreto nº 6.899, de 15 de julho de 2009, e com as normas editadas pelo Conselho Nacional de Controle de Experimentação Animal (CONCEA), e foi aprovada pela COMISSÃO DE ÉTICA NO USO DE ANIMAIS (CEUA) e pelas áreas de apoio indicadas pelo pesquisador.

Projeto: 150353

Data de Aprovação do Projeto: 21/10/2016

Título: AVALIAÇÃO PROTEÔMICA NO MODELO EXPERIMENTAL DE DEPRESSÃO UNIPOLAR INDUZIDO POR ESTRESSE CRÔNICO MODERADO E IMPREVISÍVEL **Data de Término:** 01/03/2018

Pesquisador Responsável: ADRIANE RIBEIRO ROSA

Equipe de pesquisa:

BRUNA MARIA ASCOLI	ELLEN SCOTTON	LUIZA PAUL GÉA	RAFAEL COLOMBO
Submissão	Documento	Espécie/Linhagem	Sexo/Idade
24/07/2015	APROVAÇÃO	RATO - WISTAR	M/2meses
22/12/2016	EMENDA	N/A	-/-
15/04/2018	EMENDA		/
Total de Animais:			95


ANA HELENA DA ROSA PAZ
 Coordenadora

Comissão de Ética no Uso de Animais

- Os membros da CEUA/HCPA não participaram do processo de avaliação onde constam como pesquisadores.

- Toda e qualquer alteração do Projeto deverá ser comunicada à CEUA/HCPA.

- O pesquisador deverá apresentar relatórios semestrais de acompanhamento e relatório final ao CEUA/HCPA.

ANEXO B

Instruções para a submissão de manuscritos na revista Brain, Behavior, and Immunity.



TABLE OF CONTENTS

● Description	p.1
● Audience	p.2
● Impact Factor	p.2
● Abstracting and Indexing	p.2
● Editorial Board	p.2
● Guide for Authors	p.5



ISSN: 0889-1591

DESCRIPTION

Brain, Behavior, and Immunity, founded in 1987, is the official journal of the [Psychoneuroimmunology Research Society](#) (PNIRS). This innovative journal publishes peer-reviewed basic, experimental, and clinical studies dealing with **behavioral**, **neural**, **endocrine**, and **immune system** interactions in humans and animals. It is an international, interdisciplinary journal devoted to original research in neuroscience, immunology, integrative physiology, behavioral biology, psychiatry, psychology, and clinical medicine and is inclusive of research at the molecular, cellular, social, and whole organism level. The journal features online [submission](#) and review. Manuscripts are typically peer-reviewed and returned to authors within 30 days of submission, leading to timely publication of experimental results. There are no submission fees or page charges for *Brain, Behavior, and Immunity*, which is published eight times a year. Detailed instructions for authors can be found at <http://ees.elsevier.com/bbi/>.

Research areas include: Physiological mechanisms that convey messages between the immune and nervous systems and regulate their functions Stress and immunity, including the role of stress-related hormones and neurotransmitters on the immune system. Actions of cytokines, growth factors and PAMP activation on neuronal and glial cells that regulate behavior, learning, memory and neurogenesis Role of hormones, growth factors and cytokines in the immune and central or peripheral nervous systems Interactions between the immune system and brain that are involved in development of neurological, psychiatric, and mental health disorders Role of immunological processes in neurodegenerative disorders The effects of psychotropic medications on immunological mechanisms and their potential relevance to therapeutic interventions Neuroimaging studies examining how immunological mechanisms affect brain structure and function Clinical trials and experimental studies testing the effects on both immune stimulation and immune suppression on brain and behavior The role of microglia in pain, psychological processes and in psychiatric disorders Immunological mechanisms involved in traumatic brain injury and its resolution Immunologic disorders, infection and behavior Role of the immune system in development and maintenance of inflammatory and chronic pain Immune mechanisms that regulate the blood-brain-interface (BBI) Immune factors that affect health psychology Sleep, exercise, immunity and health Immune system interactions that affect behavior following use of psychotropic drugs, alcohol and other drugs of abuse Healthy aging of the immune system and brain Role of inflammation and stress during perinatal development Cancer and its treatment, stem cells and their effects on brain behavior and immunity Reciprocal communication between the microbiome, immune and nervous systems Regulation of nerve injury and repair by the immune system Psychosocial, behavioral, and neuroendocrine influences on immunity and on the development and progression of immunologically-mediated diseases Nutrition, inflammation, obesity and behavior Genomics of behavior and immunity

AUDIENCE

Neuroscientists, Immunologists, Endocrinologists, Physiologists, Psychiatrists, Rheumatologists, Clinicians

IMPACT FACTOR

2017: 6.306 © Clarivate Analytics Journal Citation Reports 2018

ABSTRACTING AND INDEXING

Scopus

EDITORIAL BOARD

Editor-in-Chief

C.M. Pariante, Institute of Psychiatry, Psychology and Neuroscience, The Maurice Wohl Clinical Neuroscience Institute, King's College London, Cutcombe Road, SE5 9RT, London, England, UK

Associate Editors

N. Harrison, Brighton and Sussex Medical School, Brighton, England, UK

M. R. Hutchinson, University of Adelaide, Adelaide, South Australia, Australia

M. Lynch, Trinity College, Dublin 2, Ireland

S.F. Maier, University of Colorado, Boulder, Colorado, USA

A.H. Miller, Emory University, Atlanta, Georgia, USA

V. Mondelli, King's College London, London, UK

Q.J. Pittman, University of Calgary, Calgary, Alberta, Canada

T.M. Reyes, University of Cincinnati College of Medicine, Cincinnati, Ohio, USA

S. Spencer, RMIT University, Bundoora Melbourne, Victoria, Australia

K.P. Su, China Medical University, Taichung, Taiwan

L.R. Watkins, University of Colorado, Boulder, Colorado, USA

Editorial Board

S. Allan, University of Manchester, Manchester, England, UK

P. Ashwood, University of California, Davis, Medical Center, Davis, California, USA

M.T. Bailey, The Ohio State University, Columbus, Ohio, USA

W.A. Banks, University of Washington, Seattle, Washington, USA

R. Barrientos, University of Colorado Boulder, Boulder, Colorado, USA

M.E. Bauer, Pontifical Catholic University of Rio Grande do Sul (PUCRS), Porto Alegre, Brazil

S. Ben-Eliyahu, Tel Aviv University, Tel Aviv, Israel

S.D. Bilbo, Harvard Medical School, Boston, Massachusetts, USA

A. Borsini, King's College London, London, England, UK

J. E. Bower, University of California at Los Angeles (UCLA), Los Angeles, California, USA

E. Brietzke, Universidade Federal de São Paulo, São Paulo, Brazil

L. Brundin, Van Andel Research Institute, Grand Rapids, Missouri, USA

L. Capuron, INRA UMR 1286 - University Bordeaux, Bordeaux, France

M.J. Carson, University of California, Riverside, Riverside, California, USA

L. Carvalho, Queen Mary University of London, London, England, UK

A. Cattaneo, King's College London, London, England, UK

J. Cavanagh, University of Glasgow, Glasgow, Scotland, UK

L.M. Christian, The Ohio State University Wexner Medical Center, 614 Columbus

C. Coe, University of Wisconsin at Madison, Madison, Wisconsin, USA

B. Conti, The Scripps Research Institute, La Jolla, California, USA

E. S. Costanzo, University of Wisconsin, Madison, Wisconsin, USA

J.F. Cryan, University College Cork, Cork, Ireland

C. Cunningham, Trinity College, Dublin 2, Ireland

C. D'Mello, University of Calgary, Calgary, Alberta, Canada

A. Danese, King's College London, London, England, UK

T. Deak, State University of New York (SUNY) at Binghamton, Binghamton, New York, USA

K. Dev, Trinity College, Dublin, Ireland

A.C. DeVries, The Ohio State University, Columbus, Ohio, USA

B.N. Dittel, BloodCenter of Wisconsin, Milwaukee, Wisconsin, USA

N. Eijkamp, Universitair Medisch Centrum Utrecht (UMC Utrecht), Utrecht, Netherlands

D. Engblom, Linköping Universitet, Linköping, Sweden
C. Engelard, The Pennsylvania State University, University Park, Pennsylvania, USA
J. Felger, Emory University School of Medicine, Atlanta, USA
R. Fernandez-Botran, University of Louisville, Louisville, Kentucky, USA
L. K. Fonken, The University of Texas at Austin, Austin, Texas, USA
J.A. Foster, McMaster University, Hamilton, Ontario, Canada
M.G. Frank, University of Colorado, Boulder, Colorado, USA
G. Freund, University of Illinois College of Medicine, Urbana, Illinois, USA
D. Ganea, Temple University, Philadelphia, Pennsylvania, USA
A. Gaultier, University of Virginia, Charlottesville, Virginia, USA
J.P. Godbout, The Ohio State University, Columbus, Ohio, USA
D. Goldsmith, Emory University School of Medicine, Atlanta, Georgia, USA
R.M. Gorczynski, University of Toronto, Toronto, Ontario, Canada
P. Grace, University of Colorado, Boulder, Colorado, USA
L. Harden, University of the Witwatersrand, Johannesburg, South Africa
A. Harkin, Trinity College, Dublin, Ireland
E. Haroon, Emory University, Atlanta, Georgia, USA
K. Hashimoto, Chiba University, Chiba, Japan
S. Hong, University of California at San Diego (UCSD), La Jolla, California, USA
M.R. Irwin, UCLA Semel Institute for Neuroscience & Human Behavior, Los Angeles, California, USA
L. Janusek, Loyola University Chicago, Maywood, Illinois, USA
D.S. Jessop, University of Bristol, Bristol, England, UK
C. Jiang, Second Military Medical University, Shanghai, China
J. D. Johnson, Kent State University, Kent, Ohio, USA
R.W. Johnson, University of Illinois at Urbana-Champaign, Urbana, Illinois, USA
I. Johnston, The University of Sydney, Sydney, New South Wales, Australia
A. Kavelaars, University of Texas M.D. Anderson Cancer Center, Houston, Texas, USA
A. Kentner, MCPHS University, Boston, Massachusetts, USA
G. Khandaker, University of Cambridge, Cambridge, England, UK
E. Kouassi, University of Montreal, Montreal, Quebec, Canada
A.W. Kusnecov, Rutgers University, New Brunswick, New Jersey, USA
J. Lasselins, Stockholms Universitet, Stockholm, Sweden
D.A. Lawrence, Wadsworth Centre, Albany, New York, USA
S. Layé, Université Victor Segalen Bordeaux 2, Bordeaux, France
Y. Li, Shanghai Jiao Tong University, Shanghai, China
Q. Liu, Dalian University of Technology, Dalian, China
D.J. Loane, University of Maryland School of Medicine, Baltimore, Maryland, USA
F.E. Lotrich, University of Pittsburgh Medical Center (UPMC), Pittsburgh, Pennsylvania, USA
A. Lovett-Racke, The Ohio State University, Columbus, Ohio, USA
C. Lowry, University of Colorado Boulder, Boulder, Colorado, USA
J.R. Lukens, University of Virginia, Charlottesville, Virginia, USA
S.K. Lutgendorf, University of Iowa, Iowa City, Iowa, USA
K. Madden, University of Rochester, Rochester, New York, USA
A.L. Marsland, University of Pittsburgh, Pittsburgh, Pennsylvania, USA
H. Mathews, Loyola University Chicago, MAYWOOD, Illinois, USA
U. Meyer, Universität Zürich, Zürich, Switzerland
G.E. Miller, Northwestern University, Evanston, Illinois, USA
P.J. Mills, University of California at San Diego (UCSD), La Jolla, California, USA
D.M. Nance, University of California at Irvine (UCI) College of Medicine, Orange, California, USA
Y. Nolan, University College Cork, Cork, Ireland
M.R. Opp, University of Colorado Boulder, Boulder, Colorado, USA
B.K. Ormerod, University of Florida, Gainesville, Florida, USA
T. Pace, University of Arizona, Tucson, Arizona, USA
C. Pae, Bucheon St. Mary's Hospital, The Republic of Korea
M.O. Parat, University of Queensland, Woolloongabba, Queensland, Australia
R. Pekelmann Markus, Universidade de São Paulo (USP), São Paulo, Brazil
Y. Peng, Nantong University, Nantong Jiangsu Province, China
A.R. Prossin, University of Texas, Houston, Texas, USA
L.M. Pyter, Ohio State University Medical Center, Columbus, Ohio, USA
N. Quan, The Ohio State University, Columbus, Ohio, USA
C.L. Raison, Emory University, Atlanta, Georgia, USA
A. Reaux Le Goazigo, INSERM, Paris, France
L Redwine, University of California at San Diego (UCSD), La Jolla, California, USA
J.S. Rhodes, University of Illinois, Urbana, Illinois, USA
N. Rohleder, Brandeis University, Waltham, Massachusetts, USA
A. Rolls, Technion - Israel Institute of Technology, Haifa, Israel
C. Rummel, Justus-Liebig-Universität Gießen, Giessen, Germany

J. Savitz, Laureate Institute for Brain Research, Tulsa, Oklahoma, USA
P.E. Sawchenko, The Salk Institute for Biological Studies, La Jolla, California, USA
M. Schedlowski, Institute for Behavioral Sciences, Zurich, Switzerland
S.J. Schleifer, UMDNJ, Newark, New Jersey, USA
S.C. Segerstrom, University of Kentucky, Lexington, Kentucky, USA
J.F. Sheridan, Ohio State University Medical Center, Columbus, Ohio, USA
R.J. Simpson, University of Arizona, Tucson, Arizona, USA
G.M. Slavich, University of California at Los Angeles (UCLA), Los Angeles, California, USA
C. Song, Guangdong Ocean University (GDOU), Zhanjiang City, China
H. Su, University of Macau, Taipa, Macao
J.L. Teeling, Southampton General Hospital, Southampton, England, UK
F. Turkheimer, King's College London, London, England, UK
A-M. van Dam, Vrije Universiteit Medisch Centrum (VUMC), Amsterdam, Netherlands
J. Van De Water, University of California, Davis, Davis, California, USA
R. von Bernhardi, Pontificia Universidad Católica de Chile, Santiago, Chile
C.V. Vorhees, Cincinnati Children's Hospital Medical Center, Cincinnati, Ohio, USA
J. Wang, Johns Hopkins University, Baltimore, Maryland, USA
Z.M. Weil, Wexner Medical Center, Columbus, Ohio, USA
E. Wohleb, University of Cincinnati, Cincinnati, Ohio, USA
J. Woods, University of Illinois at Urbana-Champaign, Urbana, USA
R. Yirmiya, Hebrew University of Jerusalem, Jerusalem, Israel
T. Yuan, Shanghai JiaoTong University School of Medicine, Shanghai, China
P.A. Zunszain, King's College London, London, England, UK

Editor-in-Chief Emeritus:

Robert Ader (1987–2002), University of Rochester
Keith W. Kelley (2003–2017), University of Illinois

GUIDE FOR AUTHORS

Your Paper Your Way

We now differentiate between the requirements for new and revised submissions. You may choose to submit your manuscript as a single Word or PDF file to be used in the refereeing process. Only when your paper is at the revision stage, will you be requested to put your paper in to a 'correct format' for acceptance and provide the items required for the publication of your article.

To find out more, please visit the Preparation section below.

INTRODUCTION

Brain, Behavior, and Immunity, founded in 1987, is the official journal of the Psychoneuroimmunology Research Society (PNIRS). This innovative journal publishes peer-reviewed basic, experimental, and clinical studies dealing with behavioral, neural, endocrine, and immune system interactions in humans and animals. It is an international, interdisciplinary journal devoted to original research in neuroscience, immunology, integrative physiology, behavioral biology, psychiatry, psychology, and clinical medicine and is inclusive of research at the molecular, cellular, social, and whole organism level. The journal features online submission and review. Manuscripts are typically peer-reviewed and returned to authors within 30 days of submission, leading to timely publication of experimental results. There are no submission fees or page charges for *Brain, Behavior, and Immunity*, which is published eight times a year. Detailed instructions for authors can be found at <http://ees.elsevier.com/bbi/>.

Research areas include:

- Physiological mechanisms that convey messages between the immune and nervous systems and regulate their functions
- Stress and immunity, including the role of stress-related hormones and neurotransmitters on the immune system
- Actions of cytokines, growth factors and PAMP activation on neuronal and glial cells that regulate behavior, learning, memory and neurogenesis
- Role of hormones, growth factors and cytokines in the immune and central or peripheral nervous systems
- Interactions between the immune system and brain that are involved in development of neurological, psychiatric and mental health disorders
- Role of immunological processes in neurodegenerative disorders
- The effects of psychotropic medications on immunological mechanisms and their potential relevance to therapeutic interventions
- Neuroimaging studies examining how immunological mechanisms affect brain structure and function
- Clinical trials and experimental studies testing the effects on both immune stimulation and immune suppression on brain and behavior
- The role of microglia in pain, psychological processes and in psychiatric disorders
- Immunological mechanisms involved in traumatic brain injury and its resolution
- Immunologic disorders, infection and behavior
- Role of the immune system in development and maintenance of inflammatory and chronic pain
- Immune mechanisms that regulate the blood-brain-interface (BBI)
- Immune factors that affect health psychology
- Sleep, exercise, immunity and health
- Immune system interactions that affect behavior following use of psychotropic drugs, alcohol and other drugs of abuse
- Healthy aging of the immune system and brain
- Role of inflammation and stress during perinatal development
- Cancer and its treatment, stem cells and their effects on brain behavior and immunity
- Reciprocal communication between the microbiome, immune and nervous systems
- Regulation of nerve injury and repair by the immune system
- Psychosocial, behavioral, and neuroendocrine influences on immunity and on the development and progression of immunologically-mediated diseases
- Nutrition, inflammation, obesity and behavior
- Genomics of behavior and immunity

Types of article

Original full-length research reports, full-length review articles, short communications, brief commentaries, and letters to the editor will be considered for publication.

Full-length research reports: The chief criteria for the acceptance of submitted papers are the quality, originality, and clarity of the work reported, addressing one or more of the research areas reported above. There is no word limit on full length research reports, but papers should be concisely written and most should be able to articulate their findings within approximately 6,000 words.

Reviews: The journal publishes invited or unsolicited reviews on a contemporary topic, discussed authoritatively with the aim of providing a solid, and often novel, interpretation of research evidence, and of integrating a mechanistic model when applicable. Reviews consist of approximately 6,000 words of text and no more than 100 scientific references. Reviews must contain at least one figure highlighting the key aspects of the article, complete with explanatory figure legends. If appropriate,

a color version of the figure can be published in the online publication, with a black-and-white figure in the print version. If the author chooses this option, the figure legend must be self-explanatory in the absence of color-coding.

Short communications: Manuscripts published as short communications are, primarily, reports of novel, solid, important findings on contemporary, fast-moving topics. Small replication studies or incomplete data that do not move the field forward, and descriptions of methods and techniques, are not appropriate for this format. Papers will be considered short communications if the text, references, and a maximum of two tables or figures (or one of each) are limited to 3,500 words. Authors may elect to include additional illustrations, but the limitation to 3,500 words will remain.

Commentaries: These are short pieces written to accompany the publication of impactful full-length research reports. Invited by the Editor, they are limited to 900-1000 words and 5-10 references (including a reference to the relevant published report).

Viewpoints: These are opinion pieces that provide a personal view on broad, contemporary topics relevant to the interaction between health, brain, behaviour and immunity. Invited by the Editor, they are limited to 900-1000 words and 5-10 references, and will generally be immediately 'open-access' at no costs to the authors.

Letters to the editor: These should be of high scientific quality, contain less than 500 words, and cite no more than 5 scientific references. If the letter is directed to a paper published in Brain, Behavior, and Immunity, the author of that paper will be provided an opportunity to respond. Both the letter to the editor and the author's response will be published simultaneously.

Announcements: *Brain, Behavior, and Immunity* will consider for publication announcements of interest to the readership such as notices of scientific meetings.

Language (usage and editing services)

Please write your text in good English (American or British usage is accepted, but not a mixture of these). Poor standard of grammar or spelling will lead to the paper being sent back to Authors without peer-review. Authors who feel their English language manuscript may require editing to eliminate possible grammatical or spelling errors and to conform to correct scientific English may wish to use the English Language Editing service available from Elsevier's WebShop.

Study design and statistical reporting

BBI aspires to publish papers with the highest standards of reporting and presentation of methodological details, including the study design and the statistics used.

Study Design: State whether: 1) samples/animals were assigned randomly to various experimental groups (and the specific method of randomization); 2) the data collected was processed randomly and appropriately blocked; 3) experimenters were blind to group assignment and outcome assignment; and 4) an appropriate sample size was computed when the study was being designed.

Data Handling: Clearly state the numbers of participants, animals, or samples included in the study. Provide detailed explanations of the reasons for any attrition in the study. Explain how outliers are defined and handled and any data removed before analysis must be reported. Report how often each experiment was performed and whether the results were substantiated by repetition under a range of conditions. Sufficient information about sample collection must be provided to distinguish between independent biological data points and technical replicates.

Statistical reporting: Authors should identify the precise statistical tests used. In addition, planned comparisons, details of controls and power analyses to determine sample sizes, if applicable, should be reported. Complete results of the statistical analyses, including p values (rather than ranges), degrees of freedom and any estimates of effects size, should be reported in full in the Results section, including all within- and between-subject factors. For multiple comparisons and multiple correlations, define measures taken to reduce Type 1 errors. For neuroimaging studies, methods for controlling for multiple comparisons and the cluster-forming statistical threshold used must be reported. For ANOVAs, and other multivariate analyses, define measures taken to control for violation of the sphericity assumption and how you report results of corrected degrees of freedom statistics. Finally, state the name and version of the statistical software that was used.

Addressing Sex as a Biological Variable: We ask all authors to ensure proper consideration of sex as a biological variable. For example, any papers utilizing subjects (cells, animals, humans) of only one sex must state the sex of the samples in the title and abstract of the paper, with the obvious exception of sex-specific issues (e.g., prostate or ovarian function). Authors must also state the rationale for using samples from one sex rather than from both. For cellular work, the sex of origin of cells used should be reported, or if cells or tissue from both sexes were used without regard to sex, this fact should be indicated. Finally, the inability for any reason to study sex differences where they may exist should be discussed as a study limitation.

Format

Manuscripts should be prepared using a 12-point font, double-spaced throughout (including tables, footnotes, references, and figure captions) with 1-in. margins on all sides. Unusual typeface is acceptable only if it is clear and legible. For initial submission, all manuscripts must be prepared and submitted in one of the following formats: Microsoft Word (.doc), WordPerfect (.wps), or Rich Text Format (.rtf). All figures and tables should be clearly labeled at the top.

Revised manuscripts should not be marked using underlined or bolded words to indicate changes from the original submission. Instead, changes in the revised manuscript must be explained in a rebuttal letter. Submission of all revised manuscripts requires both figures and tables to be submitted separately from the manuscript text: do not insert figures and tables at the end of the text for revised manuscripts. Instead, the electronic submission system requires identification and submission of figures and tables separate from the text of revised manuscripts (see information below for graphs, scans, and illustrations). For more information, please also see the Author Gateway Web page for *Brain, Behavior, and Immunity* available through the journal home page at <https://www.elsevier.com/locate/ybrbi>.

Contact details for submission

Manuscripts must be written in English and submitted electronically at <http://ees.elsevier.com/bbi/>. New contributors should first register at this site and then log into the Elsevier Editorial System (EES) with their user name and password. There are eight steps that must be completed to submit a manuscript: Enter Article Title; Select Article Type; Add/Edit Remove Author (corresponding author does not need to be the person who submits the paper); Submit Abstract; Enter Key Words; Select Document Classification; Enter Comments (recommend expert reviewers); Attach Files. All sections except the last one can be 'copied and pasted' into text boxes from existing files. The files that must be attached separately are: cover letter to the Editor-in-Chief, manuscript, figures, and tables. An introductory cover letter must outline the most important research findings and their significance. Complete legends (captions) for both figures and tables should be placed at the end of the manuscript. Figures must be attached as separate files or as a single file. Tables must also be attached as either individual tables or a single file with all the tables. All files containing figures or tables must clearly identify each figure or part of figure by adding, at the top of each figure or table, the name of the first author and abbreviated title of the manuscript. Authors can also upload supplementary material such as video, audio, movie and other files (which will be available as a link in the PDF file that the system generates). After the files are attached, the EES system will create a PDF file, which may require a few minutes. You will then be asked to approve the PDF file, a step that must be completed before the new submission is sent to the Editor-in-Chief who will initiate the review process.

Submission checklist

You can use this list to carry out a final check of your submission before you send it to the journal for review. Please check the relevant section in this Guide for Authors for more details.

Ensure that the following items are present:

One author has been designated as the corresponding author with contact details:

- E-mail address
- Full postal address

All necessary files have been uploaded:

Manuscript:

- Include keywords
- All figures (include relevant captions)
- All tables (including titles, description, footnotes)
- Ensure all figure and table citations in the text match the files provided
- Indicate clearly if color should be used for any figures in print

Graphical Abstracts / Highlights files (where applicable)

Supplemental files (where applicable)

Further considerations

- Manuscript has been 'spell checked' and 'grammar checked'
- All references mentioned in the Reference List are cited in the text, and vice versa
- Permission has been obtained for use of copyrighted material from other sources (including the Internet)
- A competing interests statement is provided, even if the authors have no competing interests to declare
- Journal policies detailed in this guide have been reviewed
- Referee suggestions and contact details provided, based on journal requirements

For further information, visit our [Support Center](#).

BEFORE YOU BEGIN

Ethics in publishing

Please see our information pages on [Ethics in publishing](#) and [Ethical guidelines for journal publication](#).

Studies in humans and animals

If the work involves the use of human subjects, the author should ensure that the work described has been carried out in accordance with [The Code of Ethics of the World Medical Association \(Declaration of Helsinki\)](#) for experiments involving humans. The manuscript should be in line with the [Recommendations for the Conduct, Reporting, Editing and Publication of Scholarly Work in Medical Journals](#) and aim for the inclusion of representative human populations (sex, age and ethnicity) as per those recommendations. The terms [sex and gender](#) should be used correctly.

Authors should include a statement in the manuscript that informed consent was obtained for experimentation with human subjects. The privacy rights of human subjects must always be observed.

All animal experiments should comply with the [ARRIVE guidelines](#) and should be carried out in accordance with the U.K. Animals (Scientific Procedures) Act, 1986 and associated guidelines, [EU Directive 2010/63/EU for animal experiments](#), or the National Institutes of Health guide for the care and use of Laboratory animals (NIH Publications No. 8023, revised 1978) and the authors should clearly indicate in the manuscript that such guidelines have been followed. The sex of animals must be indicated, and where appropriate, the influence (or association) of sex on the results of the study.

Declaration of interest

All authors must disclose any financial and personal relationships with other people or organizations that could inappropriately influence (bias) their work. Examples of potential competing interests include employment, consultancies, stock ownership, honoraria, paid expert testimony, patent applications/registrations, and grants or other funding. Authors must disclose any interests in two places: 1. A summary declaration of interest statement in the title page file (if double-blind) or the manuscript file (if single-blind). If there are no interests to declare then please state this: 'Declarations

of interest: none'. This summary statement will be ultimately published if the article is accepted. 2. Detailed disclosures as part of a separate Declaration of Interest form, which forms part of the journal's official records. It is important for potential interests to be declared in both places and that the information matches. [More information](#).

Submission declaration and verification

Submission of an article implies that the work described has not been published previously (except in the form of an abstract, a published lecture or academic thesis, see '[Multiple, redundant or concurrent publication](#)' for more information), that it is not under consideration for publication elsewhere, that its publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere in the same form, in English or in any other language, including electronically without the written consent of the copyright-holder. To verify originality, your article may be checked by the originality detection service [Crossref Similarity Check](#).

Use of inclusive language

Inclusive language acknowledges diversity, conveys respect to all people, is sensitive to differences, and promotes equal opportunities. Articles should make no assumptions about the beliefs or commitments of any reader, should contain nothing which might imply that one individual is superior to another on the grounds of race, sex, culture or any other characteristic, and should use inclusive language throughout. Authors should ensure that writing is free from bias, for instance by using 'he or she', 'his/her' instead of 'he' or 'his', and by making use of job titles that are free of stereotyping (e.g. 'chairperson' instead of 'chairman' and 'flight attendant' instead of 'stewardess').

Authorship

While the journal does not request details of authors contribution, in accordance with the Consensus Statement on Surgery Journals Authorship (2005) we expect that all authors meet all three of the following conditions: 1) Authors make substantial contributions to conception and design, and/or acquisition of data, and/or analysis and interpretation of data; 2) Authors participate in drafting the article or revising it critically for important intellectual content; and 3) Authors give final approval of the version to be submitted and any revised version.

Authors are expected to consider carefully the list and order of authors **before** submitting their manuscript and provide the definitive list of authors at the time of the original submission. Any addition, deletion or rearrangement of author names in the authorship list should be made only **before** the manuscript has been accepted and only if approved by the journal Editor. To request such a change, the Editor must receive the following from the **corresponding author**: (a) the reason for the change in author list and (b) written confirmation (e-mail, letter) from all authors that they agree with the addition, removal or rearrangement. In the case of addition or removal of authors, this includes confirmation from the author being added or removed.

Only in exceptional circumstances will the Editor consider the addition, deletion or rearrangement of authors **after** the manuscript has been accepted. While the Editor considers the request, publication of the manuscript will be suspended. If the manuscript has already been published in an online issue, any requests approved by the Editor will result in a corrigendum.

Reporting clinical trials

Randomized controlled trials should be presented according to the CONSORT guidelines. At manuscript submission, authors must provide the CONSORT checklist accompanied by a flow diagram that illustrates the progress of patients through the trial, including recruitment, enrollment, randomization, withdrawal and completion, and a detailed description of the randomization procedure. The [CONSORT checklist and template flow diagram](#) are available online.

Registration of clinical trials

Registration in a public trials registry is a condition for publication of clinical trials in this journal in accordance with [International Committee of Medical Journal Editors](#) recommendations. Trials must register at or before the onset of patient enrolment. The clinical trial registration number should be included at the end of the abstract of the article. A clinical trial is defined as any research study that prospectively assigns human participants or groups of humans to one or more

health-related interventions to evaluate the effects of health outcomes. Health-related interventions include any intervention used to modify a biomedical or health-related outcome (for example drugs, surgical procedures, devices, behavioural treatments, dietary interventions, and process-of-care changes). Health outcomes include any biomedical or health-related measures obtained in patients or participants, including pharmacokinetic measures and adverse events. Purely observational studies (those in which the assignment of the medical intervention is not at the discretion of the investigator) will not require registration.

Article transfer service

This journal is part of our Article Transfer Service. This means that if the Editor feels your article is more suitable in one of our other participating journals, then you may be asked to consider transferring the article to one of those. If you agree, your article will be transferred automatically on your behalf with no need to reformat. Please note that your article will be reviewed again by the new journal.

[More information](#).

Copyright

Upon acceptance of an article, authors will be asked to complete a 'Journal Publishing Agreement' (see [more information](#) on this). An e-mail will be sent to the corresponding author confirming receipt of the manuscript together with a 'Journal Publishing Agreement' form or a link to the online version of this agreement.

Subscribers may reproduce tables of contents or prepare lists of articles including abstracts for internal circulation within their institutions. [Permission](#) of the Publisher is required for resale or distribution outside the institution and for all other derivative works, including compilations and translations. If excerpts from other copyrighted works are included, the author(s) must obtain written permission from the copyright owners and credit the source(s) in the article. Elsevier has [preprinted forms](#) for use by authors in these cases.

For gold open access articles: Upon acceptance of an article, authors will be asked to complete an 'Exclusive License Agreement' ([more information](#)). Permitted third party reuse of gold open access articles is determined by the author's choice of [user license](#).

Author rights

As an author you (or your employer or institution) have certain rights to reuse your work. [More information](#).

Elsevier supports responsible sharing

Find out how you can [share your research](#) published in Elsevier journals.

Role of the funding source

You are requested to identify who provided financial support for the conduct of the research and/or preparation of the article and to briefly describe the role of the sponsor(s), if any, in study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the article for publication. If the funding source(s) had no such involvement then this should be stated.

Funding body agreements and policies

Elsevier has established a number of agreements with funding bodies which allow authors to comply with their funder's open access policies. Some funding bodies will reimburse the author for the gold open access publication fee. Details of [existing agreements](#) are available online.

Open access

This journal offers authors a choice in publishing their research:

Subscription

- Articles are made available to subscribers as well as developing countries and patient groups through our [universal access programs](#).
- No open access publication fee payable by authors.
- The Author is entitled to post the [accepted manuscript](#) in their institution's repository and make this public after an embargo period (known as green Open Access). The [published journal article](#) cannot be shared publicly, for example on ResearchGate or Academia.edu, to ensure the sustainability of peer-reviewed research in journal publications. The embargo period for this journal can be found below.

Gold open access

- Articles are freely available to both subscribers and the wider public with permitted reuse.
- A gold open access publication fee is payable by authors or on their behalf, e.g. by their research funder or institution.

Regardless of how you choose to publish your article, the journal will apply the same peer review criteria and acceptance standards.

For gold open access articles, permitted third party (re)use is defined by the following [Creative Commons user licenses](#):

Creative Commons Attribution (CC BY)

Lets others distribute and copy the article, create extracts, abstracts, and other revised versions, adaptations or derivative works of or from an article (such as a translation), include in a collective work (such as an anthology), text or data mine the article, even for commercial purposes, as long as they credit the author(s), do not represent the author as endorsing their adaptation of the article, and do not modify the article in such a way as to damage the author's honor or reputation.

Creative Commons Attribution-NonCommercial-NoDerivs (CC BY-NC-ND)

For non-commercial purposes, lets others distribute and copy the article, and to include in a collective work (such as an anthology), as long as they credit the author(s) and provided they do not alter or modify the article.

The gold open access publication fee for this journal is **USD 3150**, excluding taxes. Learn more about Elsevier's pricing policy: <https://www.elsevier.com/openaccesspricing>.

Green open access

Authors can share their research in a variety of different ways and Elsevier has a number of green open access options available. We recommend authors see our [open access page](#) for further information. Authors can also self-archive their manuscripts immediately and enable public access from their institution's repository after an embargo period. This is the version that has been accepted for publication and which typically includes author-incorporated changes suggested during submission, peer review and in editor-author communications. Embargo period: For subscription articles, an appropriate amount of time is needed for journals to deliver value to subscribing customers before an article becomes freely available to the public. This is the embargo period and it begins from the date the article is formally published online in its final and fully citable form. [Find out more](#).

This journal has an embargo period of 12 months.

Submit your article

Please submit your article via <http://ees.elsevier.com/bbi>

Referees

Please submit the names and institutional e-mail addresses of several potential referees. For more details, visit our [Support site](#). Note that the editor retains the sole right to decide whether or not the suggested reviewers are used.

Additional information

PREPARATION

NEW SUBMISSIONS

Submission to this journal proceeds totally online and you will be guided stepwise through the creation and uploading of your files. The system automatically converts your files to a single PDF file, which is used in the peer-review process.

As part of the Your Paper Your Way service, you may choose to submit your manuscript as a single file to be used in the refereeing process. This can be a PDF file or a Word document, in any format or layout that can be used by referees to evaluate your manuscript. It should contain high enough quality figures for refereeing. If you prefer to do so, you may still provide all or some of the source files at the initial submission. Please note that individual figure files larger than 10 MB must be uploaded separately.

References

There are no strict requirements on reference formatting at submission. References can be in any style or format as long as the style is consistent. Where applicable, author(s) name(s), journal title/book title, chapter title/article title, year of publication, volume number/book chapter and the article

number or pagination must be present. Use of DOI is highly encouraged. The reference style used by the journal will be applied to the accepted article by Elsevier at the proof stage. Note that missing data will be highlighted at proof stage for the author to correct.

Formatting requirements

There are no strict formatting requirements but all manuscripts must contain the essential elements needed to convey your manuscript, for example Abstract, Keywords, Introduction, Materials and Methods, Results, Conclusions, Artwork and Tables with Captions.

If your article includes any Videos and/or other Supplementary material, this should be included in your initial submission for peer review purposes.

Divide the article into clearly defined sections.

Figures and tables embedded in text

Please ensure the figures and the tables included in the single file are placed next to the relevant text in the manuscript, rather than at the bottom or the top of the file. The corresponding caption should be placed directly below the figure or table.

REVISED SUBMISSIONS

Use of word processing software

Regardless of the file format of the original submission, at revision you must provide us with an editable file of the entire article. Keep the layout of the text as simple as possible. Most formatting codes will be removed and replaced on processing the article. The electronic text should be prepared in a way very similar to that of conventional manuscripts (see also the [Guide to Publishing with Elsevier](#)). See also the section on Electronic artwork.

To avoid unnecessary errors you are strongly advised to use the 'spell-check' and 'grammar-check' functions of your word processor.

Article structure

Subdivision - numbered sections

Divide your article into clearly defined and numbered sections. Subsections should be numbered 1.1 (then 1.1.1, 1.1.2, ...), 1.2, etc. (the abstract is not included in section numbering). Use this numbering also for internal cross-referencing: do not just refer to 'the text'. Any subsection may be given a brief heading. Each heading should appear on its own separate line.

Introduction

State the objectives of the work and provide an adequate background, avoiding a detailed literature survey or a summary of the results.

Material and methods

Provide sufficient details to allow the work to be reproduced by an independent researcher. Methods that are already published should be summarized, and indicated by a reference. If quoting directly from a previously published method, use quotation marks and also cite the source. Any modifications to existing methods should also be described.

Results

Results should be clear and concise.

Discussion

This should explore the significance of the results of the work, not repeat them. Avoid extensive citations and discussion of published literature.

Conclusions

The main conclusions of the study may be presented in a short Conclusions section, which may stand alone or form a subsection of a Discussion.

Appendices

If there is more than one appendix, they should be identified as A, B, etc. Formulae and equations in appendices should be given separate numbering: Eq. (A.1), Eq. (A.2), etc.; in a subsequent appendix, Eq. (B.1) and so on. Similarly for tables and figures: Table A.1; Fig. A.1, etc.

Essential title page information

Title. Concise and informative. Titles are often used in information-retrieval systems. Avoid abbreviations and formulae where possible. **Author names and affiliations.** Please clearly indicate the given name(s) and family name(s) of each author and check that all names are accurately spelled. You can add your name between parentheses in your own script behind the English transliteration. Present the authors' affiliation addresses (where the actual work was done) below the names. Indicate all affiliations with a lower-case superscript letter immediately after the author's name and in front of the appropriate address. Provide the full postal address of each affiliation, including the country name and, if available, the e-mail address of each author. **Corresponding author.** Clearly indicate who will handle correspondence at all stages of refereeing and publication, also post-publication. This responsibility includes answering any future queries about Methodology and Materials. Ensure that the e-mail address is given and that contact details are kept up to date by the corresponding author. **Present/permanent address.** If an author has moved since the work described in the article was done, or was visiting at the time, a 'Present address' (or 'Permanent address') may be indicated as a footnote to that author's name. The address at which the author actually did the work must be retained as the main, affiliation address. Superscript Arabic numerals are used for such footnotes. **Word count.** Please include a word count, excluding references and tables.

Highlights

Highlights are mandatory for this journal. They consist of a short collection of bullet points that convey the core findings of the article and should be submitted in a separate editable file in the online submission system. Please use 'Highlights' in the file name and include 3 to 5 bullet points (maximum 85 characters, including spaces, per bullet point). You can view [example Highlights](#) on our information site.

Abstract

A concise and factual abstract is required. The abstract should state briefly the purpose of the research, the principal results and major conclusions. An abstract is often presented separately from the article, so it must be able to stand alone. For this reason, References should be avoided, but if essential, then cite the author(s) and year(s). Also, non-standard or uncommon abbreviations should be avoided, but if essential they must be defined at their first mention in the abstract itself.

Keywords

A list of up to 10 **keywords** or phrases suitable for indexing should be provided.

Abbreviations

Do not use periods after abbreviations of measure (cm, s, kg, mA, etc.) in text or tables, except for "in." (inch). The American Chemical Society *Style Guide* should be used as a reference for proper abbreviations.

Acknowledgements

Collate acknowledgements in a separate section at the end of the article before the references and do not, therefore, include them on the title page, as a footnote to the title or otherwise. List here those individuals who provided help during the research (e.g., providing language help, writing assistance or proof reading the article, etc.).

Formatting of funding sources

List funding sources in this standard way to facilitate compliance to funder's requirements:

Funding: This work was supported by the National Institutes of Health [grant numbers xxxx, yyyy]; the Bill & Melinda Gates Foundation, Seattle, WA [grant number zzzz]; and the United States Institutes of Peace [grant number aaaa].

It is not necessary to include detailed descriptions on the program or type of grants and awards. When funding is from a block grant or other resources available to a university, college, or other research institution, submit the name of the institute or organization that provided the funding.

If no funding has been provided for the research, please include the following sentence:

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Units

Follow internationally accepted rules and conventions: use the international system of units (SI). If other units are mentioned, please give their equivalent in SI.

Math formulae

Please submit math equations as editable text and not as images. Present simple formulae in line with normal text where possible and use the solidus (/) instead of a horizontal line for small fractional terms, e.g., X/Y. In principle, variables are to be presented in italics. Powers of e are often more conveniently denoted by exp. Number consecutively any equations that have to be displayed separately from the text (if referred to explicitly in the text).

Footnotes

Footnotes should be used sparingly. Number them consecutively throughout the article. Many word processors build footnotes into the text, and this feature may be used. Should this not be the case, indicate the position of footnotes in the text and present the footnotes themselves separately at the end of the article.

Electronic artwork

General points

- Make sure you use uniform lettering and sizing of your original artwork.
- Preferred fonts: Arial (or Helvetica), Times New Roman (or Times), Symbol, Courier.
- Number the illustrations according to their sequence in the text.
- Use a logical naming convention for your artwork files.
- Indicate per figure if it is a single, 1.5 or 2-column fitting image.
- For Word submissions only, you may still provide figures and their captions, and tables within a single file at the revision stage.
- Please note that individual figure files larger than 10 MB must be provided in separate source files. A detailed [guide on electronic artwork](#) is available.

You are urged to visit this site; some excerpts from the detailed information are given here.

Formats

Regardless of the application used, when your electronic artwork is finalized, please 'save as' or convert the images to one of the following formats (note the resolution requirements for line drawings, halftones, and line/halftone combinations given below):

EPS (or PDF): Vector drawings. Embed the font or save the text as 'graphics'.

TIFF (or JPG): Color or grayscale photographs (halftones): always use a minimum of 300 dpi.

TIFF (or JPG): Bitmapped line drawings: use a minimum of 1000 dpi.

TIFF (or JPG): Combinations bitmapped line/half-tone (color or grayscale): a minimum of 500 dpi is required.

Please do not:

- Supply files that are optimized for screen use (e.g., GIF, BMP, PICT, WPG); the resolution is too low.
- Supply files that are too low in resolution.
- Submit graphics that are disproportionately large for the content.

Color artwork

Please make sure that artwork files are in an acceptable format (TIFF (or JPEG), EPS (or PDF), or MS Office files) and with the correct resolution. If, together with your accepted article, you submit usable color figures then Elsevier will ensure, at no additional charge, that these figures will appear in color online (e.g., ScienceDirect and other sites) regardless of whether or not these illustrations are reproduced in color in the printed version. **For color reproduction in print, you will receive information regarding the costs from Elsevier after receipt of your accepted article.** Please indicate your preference for color: in print or online only. [Further information on the preparation of electronic artwork](#).

Illustration services

Elsevier's WebShop offers Illustration Services to authors preparing to submit a manuscript but concerned about the quality of the images accompanying their article. Elsevier's expert illustrators can produce scientific, technical and medical-style images, as well as a full range of charts, tables and graphs. Image 'polishing' is also available, where our illustrators take your image(s) and improve them to a professional standard. Please visit the website to find out more.

Figure captions

Ensure that each illustration has a caption. A caption should comprise a brief title (**not** on the figure itself) and a description of the illustration. Keep text in the illustrations themselves to a minimum but explain all symbols and abbreviations used.

Tables

Please submit tables as editable text and not as images. Tables can be placed either next to the relevant text in the article, or on separate page(s) at the end. Number tables consecutively in accordance with their appearance in the text and place any table notes below the table body. Be sparing in the use of tables and ensure that the data presented in them do not duplicate results described elsewhere in the article. Please avoid using vertical rules and shading in table cells.

References

Citation in text

Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. Unpublished results and personal communications are not recommended in the reference list, but may be mentioned in the text. If these references are included in the reference list they should follow the standard reference style of the journal and should include a substitution of the publication date with either 'Unpublished results' or 'Personal communication'. Citation of a reference as 'in press' implies that the item has been accepted for publication.

Web references

As a minimum, the full URL should be given and the date when the reference was last accessed. Any further information, if known (DOI, author names, dates, reference to a source publication, etc.), should also be given. Web references can be listed separately (e.g., after the reference list) under a different heading if desired, or can be included in the reference list.

Data references

This journal encourages you to cite underlying or relevant datasets in your manuscript by citing them in your text and including a data reference in your Reference List. Data references should include the following elements: author name(s), dataset title, data repository, version (where available), year, and global persistent identifier. Add [dataset] immediately before the reference so we can properly identify it as a data reference. The [dataset] identifier will not appear in your published article.

References in a special issue

Please ensure that the words 'this issue' are added to any references in the list (and any citations in the text) to other articles in the same Special Issue.

Reference management software

Most Elsevier journals have their reference template available in many of the most popular reference management software products. These include all products that support [Citation Style Language styles](#), such as [Mendeley](#). Using citation plug-ins from these products, authors only need to select the appropriate journal template when preparing their article, after which citations and bibliographies will be automatically formatted in the journal's style. If no template is yet available for this journal, please follow the format of the sample references and citations as shown in this Guide. If you use reference management software, please ensure that you remove all field codes before submitting the electronic manuscript. [More information on how to remove field codes from different reference management software](#).

Users of Mendeley Desktop can easily install the reference style for this journal by clicking the following link:

<http://open.mendeley.com/use-citation-style/brain-behavior-and-immunity>

When preparing your manuscript, you will then be able to select this style using the Mendeley plug-ins for Microsoft Word or LibreOffice.

Reference formatting

There are no strict requirements on reference formatting at submission. References can be in any style or format as long as the style is consistent. Where applicable, author(s) name(s), journal title/book title, chapter title/article title, year of publication, volume number/book chapter and the article number or pagination must be present. Use of DOI is highly encouraged. The reference style used by the journal will be applied to the accepted article by Elsevier at the proof stage. Note that missing data will be highlighted at proof stage for the author to correct. If you do wish to format the references yourself they should be arranged according to the following examples:

Reference style

Text: All citations in the text should refer to:

1. **Single author:** the author's name (without initials, unless there is ambiguity) and the year of publication;
2. **Two authors:** both authors' names and the year of publication;
3. **Three or more authors:** first author's name followed by 'et al.' and the year of publication.

Citations may be made directly (or parenthetically). Groups of references can be listed either first alphabetically, then chronologically, or vice versa.

Examples: 'as demonstrated (Allan, 2000a, 2000b, 1999; Allan and Jones, 1999).... Or, as demonstrated (Jones, 1999; Allan, 2000)... Kramer et al. (2010) have recently shown ...'

List: References should be arranged first alphabetically and then further sorted chronologically if necessary. More than one reference from the same author(s) in the same year must be identified by the letters 'a', 'b', 'c', etc., placed after the year of publication.

Examples:

Reference to a journal publication:

Van der Geer, J., Hanraads, J.A.J., Lupton, R.A., 2010. The art of writing a scientific article. *J. Sci. Commun.* 163, 51–59. <https://doi.org/10.1016/j.Sc.2010.00372>.

Reference to a journal publication with an article number:

Van der Geer, J., Hanraads, J.A.J., Lupton, R.A., 2018. The art of writing a scientific article. *Heliyon*. 19, e00205. <https://doi.org/10.1016/j.heliyon.2018.e00205>.

Reference to a book:

Strunk Jr., W., White, E.B., 2000. *The Elements of Style*, fourth ed. Longman, New York.

Reference to a chapter in an edited book:

Mettam, G.R., Adams, L.B., 2009. How to prepare an electronic version of your article, in: Jones, B.S., Smith , R.Z. (Eds.), *Introduction to the Electronic Age*. E-Publishing Inc., New York, pp. 281–304.

Reference to a website:

Cancer Research UK, 1975. Cancer statistics reports for the UK. <http://www.cancerresearchuk.org/aboutcancer/statistics/cancerstatsreport/> (accessed 13 March 2003).

Reference to a dataset:

[dataset] Oguro, M., Imahiro, S., Saito, S., Nakashizuka, T., 2015. Mortality data for Japanese oak wilt disease and surrounding forest compositions. Mendeley Data, v1. <https://doi.org/10.17632/xwj98nb39r.1>.

Journal abbreviations source

Journal names should be abbreviated according to the [List of Title Word Abbreviations](#).

Video

Elsevier accepts video material and animation sequences to support and enhance your scientific research. Authors who have video or animation files that they wish to submit with their article are strongly encouraged to include links to these within the body of the article. This can be done in the same way as a figure or table by referring to the video or animation content and noting in the body text where it should be placed. All submitted files should be properly labeled so that they directly relate to the video file's content. In order to ensure that your video or animation material is directly usable, please provide the file in one of our recommended file formats with a preferred maximum size of 150 MB per file, 1 GB in total. Video and animation files supplied will be published online in the electronic version of your article in Elsevier Web products, including [ScienceDirect](#). Please supply 'stills' with your files: you can choose any frame from the video or animation or make a separate image. These will be used instead of standard icons and will personalize the link to your video data. For more detailed instructions please visit our [video instruction pages](#). Note: since video and animation cannot be embedded in the print version of the journal, please provide text for both the electronic and the print version for the portions of the article that refer to this content.

Data visualization

Include interactive data visualizations in your publication and let your readers interact and engage more closely with your research. Follow the instructions [here](#) to find out about available data visualization options and how to include them with your article.

Supplementary material

Supplementary material such as applications, images and sound clips, can be published with your article to enhance it. Submitted supplementary items are published exactly as they are received (Excel or PowerPoint files will appear as such online). Please submit your material together with the article and supply a concise, descriptive caption for each supplementary file. If you wish to make changes to

supplementary material during any stage of the process, please make sure to provide an updated file. Do not annotate any corrections on a previous version. Please switch off the 'Track Changes' option in Microsoft Office files as these will appear in the published version.

Research data

This journal encourages and enables you to share data that supports your research publication where appropriate, and enables you to interlink the data with your published articles. Research data refers to the results of observations or experimentation that validate research findings. To facilitate reproducibility and data reuse, this journal also encourages you to share your software, code, models, algorithms, protocols, methods and other useful materials related to the project.

Below are a number of ways in which you can associate data with your article or make a statement about the availability of your data when submitting your manuscript. If you are sharing data in one of these ways, you are encouraged to cite the data in your manuscript and reference list. Please refer to the "References" section for more information about data citation. For more information on depositing, sharing and using research data and other relevant research materials, visit the [research data](#) page.

Data linking

If you have made your research data available in a data repository, you can link your article directly to the dataset. Elsevier collaborates with a number of repositories to link articles on ScienceDirect with relevant repositories, giving readers access to underlying data that gives them a better understanding of the research described.

There are different ways to link your datasets to your article. When available, you can directly link your dataset to your article by providing the relevant information in the submission system. For more information, visit the [database linking page](#).

For [supported data repositories](#) a repository banner will automatically appear next to your published article on ScienceDirect.

In addition, you can link to relevant data or entities through identifiers within the text of your manuscript, using the following format: Database: xxxx (e.g., TAIR: AT1G01020; CCDC: 734053; PDB: 1XFN).

Mendeley Data

This journal supports Mendeley Data, enabling you to deposit any research data (including raw and processed data, video, code, software, algorithms, protocols, and methods) associated with your manuscript in a free-to-use, open access repository. During the submission process, after uploading your manuscript, you will have the opportunity to upload your relevant datasets directly to *Mendeley Data*. The datasets will be listed and directly accessible to readers next to your published article online.

For more information, visit the [Mendeley Data for journals page](#).

Data in Brief

You have the option of converting any or all parts of your supplementary or additional raw data into one or multiple data articles, a new kind of article that houses and describes your data. Data articles ensure that your data is actively reviewed, curated, formatted, indexed, given a DOI and publicly available to all upon publication. You are encouraged to submit your article for *Data in Brief* as an additional item directly alongside the revised version of your manuscript. If your research article is accepted, your data article will automatically be transferred over to *Data in Brief* where it will be editorially reviewed and published in the open access data journal, *Data in Brief*. Please note an open access fee of 500 USD is payable for publication in *Data in Brief*. Full details can be found on the [Data in Brief website](#). Please use [this template](#) to write your Data in Brief.

Data statement

To foster transparency, we encourage you to state the availability of your data in your submission. This may be a requirement of your funding body or institution. If your data is unavailable to access or unsuitable to post, you will have the opportunity to indicate why during the submission process, for example by stating that the research data is confidential. The statement will appear with your published article on ScienceDirect. For more information, visit the [Data Statement page](#).

Additional information

AFTER ACCEPTANCE

Online proof correction

Corresponding authors will receive an e-mail with a link to our online proofing system, allowing annotation and correction of proofs online. The environment is similar to MS Word: in addition to editing text, you can also comment on figures/tables and answer questions from the Copy Editor. Web-based proofing provides a faster and less error-prone process by allowing you to directly type your corrections, eliminating the potential introduction of errors.

If preferred, you can still choose to annotate and upload your edits on the PDF version. All instructions for proofing will be given in the e-mail we send to authors, including alternative methods to the online version and PDF.

We will do everything possible to get your article published quickly and accurately. Please use this proof only for checking the typesetting, editing, completeness and correctness of the text, tables and figures. Significant changes to the article as accepted for publication will only be considered at this stage with permission from the Editor. It is important to ensure that all corrections are sent back to us in one communication. Please check carefully before replying, as inclusion of any subsequent corrections cannot be guaranteed. Proofreading is solely your responsibility.

Offprints

The corresponding author will, at no cost, receive a customized [Share Link](#) providing 50 days free access to the final published version of the article on [ScienceDirect](#). The Share Link can be used for sharing the article via any communication channel, including email and social media. For an extra charge, paper offprints can be ordered via the offprint order form which is sent once the article is accepted for publication. Both corresponding and co-authors may order offprints at any time via Elsevier's [Webshop](#). Corresponding authors who have published their article gold open access do not receive a Share Link as their final published version of the article is available open access on ScienceDirect and can be shared through the article DOI link.

Additional information

AUTHOR INQUIRIES

Visit the [Elsevier Support Center](#) to find the answers you need. Here you will find everything from Frequently Asked Questions to ways to get in touch.

You can also [check the status of your submitted article](#) or find out [when your accepted article will be published](#).

© Copyright 2018 Elsevier | <https://www.elsevier.com>