



UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL

FACULDADE DE MEDICINA

**PROGRAMA DE PÓS-GRADUAÇÃO EM PSIQUIATRIA E CIÊNCIAS DO
COMPORTAMENTO**

DISSERTAÇÃO DE MESTRADO

**FATOR NEUROTRÓFICO DERIVADO DO CÉREBRO NOS TRANSTORNOS POR
USO DE SUBSTÂNCIAS – UMA META-ANÁLISE**

FELIPE ORNELL

Orientador: Prof. Dra. Lisia von Diemen

Co-orientador: Prof. Dr. Felix Kessler

Porto Alegre

Abril de 2017



UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL
FACULDADE DE MEDICINA
**PROGRAMA DE PÓS-GRADUAÇÃO EM PSIQUIATRIA E CIÊNCIAS DO
COMPORTAMENTO**

DISSERTAÇÃO DE MESTRADO

**FATOR NEUROTRÓFICO DERIVADO DO CÉREBRO (BDNF) NOS
TRANSTORNOS POR USO DE SUBSTÂNCIAS – UMA META-ANÁLISE**

Dissertação de Mestrado a ser apresentada ao Programa de Pós-Graduação em Psiquiatria e Ciências do Comportamento como requisito parcial à obtenção do título de Mestre em Psiquiatria e Ciências do Comportamento.

FELIPE ORNELL

Orientadora: Profa. Dra. Lisia von Diemen

Co-orientador: Prof. Dr. Felix Kessler

Porto Alegre

Abril de 2017

CATALOGAÇÃO

CIP - Catalogação na Publicação

Ornell, Felipe
FATOR NEUROTRÓFICO DERIVADO DO CÉREBRO NOS
TRANSTORNOS POR USO DE SUBSTÂNCIAS – UMA META-ANÁLISE
/ Felipe Ornella. -- 2017.
93 f.

Orientadora: Lisia von Diemen.
Coorientador: Felix Kessler.

Dissertação (Mestrado) -- Universidade Federal do
Rio Grande do Sul, Faculdade de Medicina, Programa
de Pós-Graduação em Ciências Médicas: Psiquiatria,
Porto Alegre, BR-RS, 2017.

1. Transtornos por uso de substâncias. 2. BDNF. 3.
Biomarcadores. 4. Meta-análise. I. von Diemen, Lisia
, orient. II. Kessler, Felix, coorient. III. Título.

“Ostra feliz não faz pérola”. A ostra, para fazer uma pérola, precisa ter dentro de si um grão de areia que a faça sofrer. Sofrendo a ostra diz para si mesma:

“Preciso envolver essa areia pontuda que me machuca com uma esfera lisa que lhe tire as pontas...” Ostras felizes não fazem pérolas... Pessoas felizes não sentem a necessidade de criar. O ato criador, seja na ciência ou na arte, surge sempre de uma dor. Não é preciso que seja uma dor doída... Por vezes a dor aparece como aquela coceira que tem o nome de curiosidade (...)

Rubem Alves

À Laís Taffarel.

AGRADECIMENTOS

Aos meus orientadores Prof^a. Dra Lisia von Diemen e Prof. Dr Felix Kessler, exemplos de profissionais e de seres humanos. A minha gratidão por me possibilitarem “viver” um mestrado. Obrigado pelo aprendizado diário, pela paciência, por me estimularem a crescer dentro e fora da academia e por me guiarem neste processo.

Ao Prof. Dr Flavio Pechansky por ter aberto as portas do grupo de pesquisa, por atuar como orientador não oficial, por instigar a pensar sempre além e buscar a excelência mesmo nas coisas mais simples.

À Juliana Nicterwitz Scherer, colega e amiga que surgiu na academia e passou a ser parte indispensável da minha vida. Obrigado pela parceria diária dos últimos 4 anos, obrigado por partilhar a tua capacidade ímpar de síntese, e emprestar a tua genialidade e os teus ouvidos. Obrigado por me ensinar que pode haver equilíbrio entre profissional e pessoal.

Ao Prof. Dr. João Quevedo, meu orientador de Iniciação Científica, a pessoa que me possibilitou iniciar uma carreira acadêmica sólida e que me indicou dar sequência a ela dentro do CPAD.

À Professora Nerilza Volpato por ser um modelo de profissional e de ser humano. Obrigado por ter estado presente nos momentos mais difíceis.

À Professora Yara Lamen Lhanos que me fez ter vontade de ser professor.

À Dra. Fernanda Hansen e ao Dr. Felipe Schuch meus coorientadores não oficiais, obrigado. Obrigado por todo o auxílio e por toda a paciência, sem vocês este trabalho não teria acontecido.

Ao Fernando Pezzini Rebelatto, à Ana Laura Tavares e ao Andrei Garziera pelo auxílio durante todo o processo de desenvolvimento deste estudo, sem vocês o estudo também não teria ocorrido.

À Dra Anne Sordi, primeira pessoa com quem trabalhei diretamente dentro do grupo. Obrigado por todo o espaço que me deste desde o início, pela confiança e por estimular meu raciocínio operacional e metodológico.

À Dra Roberta Silvestrin e à Dra. Joana Narvaez, parceiras de trabalho e de vida. Obrigado pela disponibilidade, por todos os aprendizados e principalmente pelo afeto.

À Dra. Carla Dal Bosco, Dra Tanara Sousa e Silvia Halpern pelo apoio incondicional e por todo o aprendizado.

À Melissa e à Cleide, peças fundamentais no funcionamento do grupo de pesquisa.

Aos alunos de Iniciação Científica, Vaness Loss, Vanessa Dal Cin, Vanessa Assunção, Vanessa Eggres, Roseana Paes, Rafaela Ornell, Guilherme, Luana, Luciana, Bruna Ferlin, e aos colegas Yeger Telles, Letícia Fara e Gerson Rossi por toda a dedicação, e comprometimento, se não fossem vocês nenhuma pesquisa do CPAD aconteceria.

À Ingrid Francke, ao Edgar Klein, ao Marcelo Slomka e à Ilan Ramos que fizeram surgir meu interesse pela dependência química.

À professora Mestre Tânia Fiorentin que mesmo com as dificuldades de lecionar em uma escola estadual de uma cidade cujo Índice de Desenvolvimento Humano era um dos 20 piores do Rio Grande do Sul me fez despertar o interesse pela ciência.

À Dra Amanda Steckert, por me ensinar toda a minha base teórica e prática sobre biomarcadores.

Ao André por ter sido meu suporte em todos os sentidos durante estes últimos anos, por ter mantido a paciência quando a minha já havia esgotado.

À minha mãe Maria Elena por ter aberto mão de muitas coisas e se esforçado além dos limites para que eu pudesse estudar.

Às outras 3 mães que a vida me deu Zélia Cavalini a “mãe preta”, minha tia Ana Fátima Rech e minha madrinha Geni Zanella Zanotelli.

RESUMO

Os transtornos por Uso de Substâncias (TUS) estão entre os transtornos psiquiátricos mais prevalentes no mundo. De acordo com a Organização Mundial de Saúde (OMS), cerca de 10% da população dos centros urbanos de todo o planeta usa, abusivamente, alguma substância psicoativa. As causas dos TUS são complexas e multifatoriais, englobando aspectos genéticos, psicossociais, comorbidades psiquiátricas, sensibilidade/resiliência ao estresse, desenvolvimento cerebral, efeitos na neuroplasticidade e no sistema cerebral de recompensa (SCR). Nos últimos anos tem crescido a proposição dos TUS como uma doença do cérebro, caracterizada por alterações disfuncionais no cérebro. Nesse sentido, a identificação de biomarcadores, que indiquem o estado patológico e a gravidade dos TUS, pode auxiliar no direcionamento e no aumento da efetividade do tratamento. O fator neurotrófico derivado do cérebro – BDNF – principal NT cerebral, parece estar implicado na base fisiopatológica de diversas doenças neurodegenerativas e psiquiátricas. Nos TUS, diversos estudos têm avaliado o BDNF em dependentes de diferentes tipos e classes de drogas, todavia os resultados são contraditórios e o papel do BDNF continua sendo um campo pouco compreendido. Nesse sentido, o objetivo deste estudo foi realizar uma meta-análise dos níveis periféricos de BDNF em usuários de substâncias psicoativas, com exceção do tabaco, comparados a controles. Para isso, realizou-se uma busca sistemática no PubMed / MEDLINE, EMBASE e PsycINFO. A seleção dos estudos elegíveis e a extração dos dados foram realizadas por dois revisores independentes; os desacordos foram resolvidos por consenso e com a opinião de um terceiro revisor. As listas de referência de todos os documentos recuperados foram avaliadas, a fim de identificar estudos elegíveis adicionais não detectados na busca inicial. Ao final, trinta estudos foram incluídos na meta-análise totalizando 1698 casos e 1363 controles. As análises foram realizadas levando em consideração o tipo de droga, a matriz de análise e o status do uso no momento da dosagem (uso ativo e abstinência); as meta-análises foram realizadas por subgrupos de acordo com o tipo de droga consumida e, nas meta-regressões, foram avaliados fatores

potencialmente relacionados à alteração do BDNF. Na análise geral não foram encontradas diferenças significativas entre casos e controles. Após ajuste para viés de publicação, encontrou-se níveis menores de BDNF sérico em usuários ativos em relação aos controles; em contrapartida, em usuários abstinentes, os níveis de BDNF foram semelhantes aos controles. A análise por droga mostrou níveis mais baixos de BDNF em dependentes de álcool, em comparação com os controles, e em dependentes de crack/cocaína durante o uso ativo. Nas análises de meta-regressão, os níveis séricos de BDNF foram associados aos anos de uso de drogas, idade, sexo e tempo de abstinência. Em geral, as análises sérica e plasmática apresentaram resultados discrepantes. Essas evidências sugerem que os níveis de BDNF sérico estão consistentemente mais baixos durante o uso ativo de drogas, mas se equiparam aos controles durante a abstinência. Mais do que isso, o BDNF parece diminuir com a idade, com o tempo de doença e, também, sofrer influência do tempo de abstinência e do gênero. Estes resultados indicam que, menores níveis de BDNF podem estar relacionados com a gravidade e com a progressão da dependência de drogas. Assim, é possível que o BDNF seja um biomarcador discriminatório de estadiamento nos TUS, como já descrito para outros transtornos psiquiátricos e doenças neurodegenerativas.

ABSTRACT

Substance Use Disorders (SUDS) are among the most prevalent psychiatric disorders worldwide. According to the World Health Organization (WHO), about 10% of the population in urban centers abuse some kind of psychoactive substance. SUDS causes are complex, multifactorial, and encompass genetic and psychosocial aspects, psychiatric comorbidities, sensitivity/resilience to stress, cerebral development, effects on the reward center and on the cerebral neuroplasticity. Over the last years, the proposition of SUDS as a brain disease is growing, as it is being associated to dysfunctional alterations on the brain. In this regard, looking for biomarkers that are able to indicate the pathological status and to identify SUDS severity could be helpful directing treatment and increasing its effectiveness. Brain-derived neurotrophic factor (BDNF), the main neurotransmitter in the brain, appears to be implicated in the pathophysiological basis of various neurodegenerative and psychiatric disorders. In SUDS, several studies have been evaluating BDNF in dependents of different types and classes of drugs. However the results are contradictory and the role of BDNF still remains poorly understood. In this sense, the aim of the present study was to perform a meta-analysis of peripheral levels of BDNF in users of psychoactive substances, with the exception of tobacco, compared to healthy controls. Hence, a systematic search was conducted through PubMed/MEDLINE, EMBASE and PsycINFO. Both the selection of eligible studies and data extraction were performed by two independent reviewers. Disagreements were resolved by consensus and with the opinion of a third reviewer. The reference list of all retrieved papers were evaluated in order to identify additional eligible studies not identified by the initial search. At the end, thirty studies were included in the meta-analysis, with a total of 1698 cases and 1363 controls. The analysis were performed taking into consideration the type of drug used, the biological matrices analysed and the status of use at the moment of the dosage (active use or withdrawn). Meta-analysis were performed by subgroups according to the type of drug used. In the meta-regressions, factors potentially related to BDNF alteration were evaluated. In the general analysis, no significative differences were found between cases and controls. After adjusting for publication bias, we found lower

levels of serum BDNF in active users compared to controls; in contrast, in withdrawn users, BDNF levels were similar to controls. Analysis by type of drug showed lower levels of BDNF in alcohol-dependent patients compared to controls, and in crack/cocaine dependents during active use. In meta-regression analysis, serum BDNF levels were associated with years of drug use, age, sex, and withdrawal time. In general, serum and plasma analysis presented divergent results. These evidences suggest that serum BDNF levels are consistently lower during active drug use, but are matched to controls during withdrawal. More than that, BDNF seems to decrease with age and with years of drug use, and also undergo the influence of withdrawal time and gender. These results indicate that lower levels of BDNF may be related to severity and progression of drug dependence. Thus, it is possible that BDNF is a discriminant staging biomarker in SUDS, as already described for other psychiatric disorders and neurodegenerative diseases.

LISTA DE ILUSTRAÇÕES

Table 1. Characteristics of the studies included in the meta-analysis of serum or plasma BDNF levels in Substance Use Disorder.....	64
Table 2. Brain-derived neurotrophic factor (BDNF) levels from serum samples.....	67
Table 3. Brain-derived neurotrophic factor (BDNF) levels from plasma samples.....	68
Table 4. Meta-regression of potential moderators brain-derived neurotrophic factor (BDNF) in substance use disorders.....	69
Figure 1. Flow diagram of the meta-analysis of brain-derived neurotrophic factor (BDNF) in substance use disorders.....	70
Figure 2. Overall, plasma and serum Brain-derived neurotrophic factor (BDNF) levels in substance use disorders.	71
Figure 3. Brain-derived neurotrophic factor (BDNF) levels from serum in alcohol users.....	72
Figure 4. Brain-derived neurotrophic factor (BDNF) levels from serum in opiate users.....	73
Figure 5. Brain-derived neurotrophic factor (BDNF) levels from serum in crack/cocaine users.....	74
Figure 6. Brain-derived neurotrophic factor (BDNF) levels from serum in methamphetamine users.....	74
Supplemental information 1. Quality assessment of the eligible studies using the Newcastle-Ottawa Scale.	75
Supplemental information 2. Quality assessment of the eligible studies using the Cochrane Collaboration.....	76

LISTA DE ABREVIATURAS E SÍMBOLOS

AIDS – Síndrome da imunodeficiência adquirida, do inglês *acquired immunodeficiency syndrome*

ATP – Adenosina trifosfato

ATV – Área tegumentar ventral

BDNF – Fator neurotrófico derivado do cérebro, do inglês *Brain-derived neurotrophic factor*

CID – Código internacional de doenças

COF – Côrtex órbito–frontal medial

CPF – Côrtex pré-frontal

CPFdI – Côrtex pré-frontal dorsolateral

DA – DA

DC – Dependente de cocaína

DH – Dependente de heroína

DM – Depressão Maior

GABA – Ácido gama-aminobutirico

DSM – Manual diagnóstico e estatístico dos transtornos mentais, do inglês *Diagnostic and statistical manual of mental disorders*

GCA – Girus cingulado anterior

GPx – Glutationa peroxidase

GSH – Glutationa

HIV – Vírus da imunodeficiência humana, do inglês *Human immunodeficiency virus*

MET – Metanfetamina

NAc – Núcleo accumbens

NGF – Fator de crescimento neural, do inglês *Nerve growth factor*

NIDA – Instituto nacional sobre drogas de abuso, do inglês *National institute of drug abuse*

NT – NT

OMS – Organização mundial da saúde

PUBMED – Website da biblioteca nacional de medicina dos Estados Unidos

RC – Circuito de recompensa cerebral, do inglês *Reward circuit*

SNC – Sistema nervoso central

SPA – Substância psicoativa

TB – Transtorno Bipolar

TUS – Transtornos por uso de substâncias

UNODC – Escritório das Nações Unidas sobre Drogas e Crime, do inglês *United Nations Office on Drugs and Crime*

SUMÁRIO

1 Introdução	16
1.1 Transtornos por Uso de Substâncias – Epidemiologia.....	16
1.2 Diagnóstico e características clínicas dos Transtornos por Uso de Substâncias	17
1.3 Neurobiologia dos Transtornos por Uso de Substâncias	19
1.4 Biomarcadores	22
1.5 BDNF	23
1.6 BDNF nos Transtornos por uso de substâncias.....	24
2 Objetivo geral:.....	29
2.1 Objetivos específicos:	29
3 Artigo.....	30
4 Conclusões e considerações finais	78
Referências.....	80
Anexo 1 – Projetos em andamento.....	89
Anexo 2. Artigos em desenvolvimento:	90
Anexo 3 – Artigos publicados durante o período do Mestrado.....	91
Anexo 4 – Artigos submetidos durante o período do Mestrado	92
Anexo 5 – Capítulos de livros escritos durante o período do Mestrado	93
Anexo 6 – Prêmios recebidos durante o período do Mestrado	94

1 Introdução

1.1 Transtornos por uso de substâncias – Epidemiologia

Os transtornos por uso de substâncias (TUS) estão entre os transtornos psiquiátricos mais prevalentes no mundo (1-4). A Organização Mundial de Saúde estima que 10% da população dos centros urbanos de todo o planeta usa abusivamente alguma substância psicoativa, independente de sexo, idade, nível social e de instrução (5).

O tabaco e o álcool são as drogas lícitas mais consumidas no mundo. Estima-se que existam cerca de 1 bilhão e 200 milhões de pessoas fumantes no mundo, fazendo com que o tabaco seja um dos quatro principais fatores de risco para doenças não transmissíveis e a principal causa global de morte evitável, gerando 6 milhões de mortes diretas ou indiretas a cada ano(6). O álcool é consumido por 38,3% da população mundial, havendo diferenças nas prevalências entre as regiões. Em 2010, o consumo de álcool entre pessoas com 15 anos ou mais foi estimado em 6.2 litros de álcool puro por ano (13,5 gramas de álcool puro por dia), um quarto deste consumo (24,8%) refere-se a álcool produzido ilegalmente. O consumo de álcool aumenta o risco de desenvolver mais de 200 doenças, incluindo cirrose hepática e alguns tipos de câncer, além de estar associado à violência e a acidentes. Em 2012, estima-se que tenham ocorrido 3,3 milhões de mortes devido ao uso nocivo do álcool representando 5,9% de todas as mortes ocorridas no mundo (7).

No Brasil, estima-se que entre 14,7% (8) e 16,9% da população é tabagista e 50% consome bebidas alcoólicas, dos quais 17% (11,7 milhões) são abusadores ou dependentes (9). Além disso, o estudo sinaliza para a precocidade do início do consumo de ambas as substâncias que, frequentemente, são apontadas como porta de entrada para drogas ilícitas (10-12).

Dados recentes, publicados pela UNODC, sugerem que, cerca de 5% da população adulta do mundo, ou 247 milhões de pessoas entre 15 e 64 anos, consumiram drogas ilícitas em 2014. Além disso, estima-se que 11,74% desta população (29 milhões de pessoas) apresente critérios para dependência; entre as

substâncias mais utilizadas destacam-se cannabis, opioides, opiáceos, cocaínicos, anfetaminas e êxtase (13). Ressalta-se que, apesar deste relatório ter sinalizado o aumento no número de usuários recreativos de drogas no mundo, se comparado aos relatórios publicados nos anos anteriores, esta estimativa se manteve proporcionalmente estável ao crescimento da população global. Apesar disso, o percentual de pessoas que apresentam critérios para dependência aumentou desproporcionalmente, pela primeira, vez em seis anos (13).

No Brasil, a maconha é a substância ilícita mais utilizada, tanto na população adulta, quanto adolescente; 6,8% dos adultos e 4,3% dos adolescentes referiram uso na vida e 2,5% e 3,4%, respectivamente, relataram uso no último ano. A cocaína inalada ocupa a segunda posição, o uso na vida foi relatado por 3,8% dos adultos e 2,3% dos adolescentes, dos quais 1,7% e 1,6%, respectivamente, relataram uso no último ano. O uso de crack na vida foi relatado por 1,3% dos adultos e 0,8% dos adolescentes, sendo que 0,7% e 0,1% fizeram uso no último ano (14). Outro dado relevante é que, 41,4% da população que relatou uso no ano anterior é dependente (9). Apesar dos índices de consumo de crack serem inferiores, se comparados a outras drogas, observa-se que é a droga ilícita que mais conduz a internações em hospitais psiquiátricos (15) e que mais provoca demanda por atendimento (16), gerando custo expressivo para o sistema público de saúde.

1.2 Diagnóstico e características clínicas dos Transtornos por Uso de Substâncias

Historicamente, observa-se que, os TUS foram mal compreendidos, considerados como uma falha moral, mau hábito ou mesmo hedonismo e não como uma condição de saúde, o que contribuiu para a desassistência à esta população, evidenciada durante muito tempo (17). Atualmente, a OMS define droga como qualquer substância química, lícita ou ilícita, medicinal ou nociva, não produzida pelo organismo, com capacidade de modificar o seu funcionamento (18),

Os TUS constituem transtornos crônicos caracterizados, sobretudo, por um conjunto de sintomas de ordem cognitiva, comportamental e fisiológica, que indicam

um padrão patológico e mal adaptativo de uso que acarreta a continuidade do consumo, apesar das consequências negativas (19, 20).

A avaliação apropriada é essencial, tanto no planejamento do processo terapêutico quanto na pesquisa clínica (23,24). O diagnóstico dos TUS é fundamentalmente clínico, apesar das dosagens de algumas enzimas auxiliarem na avaliação sobre o funcionamento hepático e cardiovascular, não existem análises bioquímicas específicas para o diagnóstico dos TUS. Atualmente, o diagnóstico dos TUS é realizado a partir de critérios estabelecidos no Código Internacional de Doenças (21) ou no Manual Diagnóstico e Estatístico dos Transtornos Mentais (22). Existem diversos instrumentos disponíveis para triar, rastrear, avaliar e diagnosticar, de forma estruturada, o uso problemático de substâncias - a indicação de cada um depende do objetivo a ser alcançado (23, 24).

Diferente do DSM IV e da CID 10, que diferenciam o diagnóstico de abuso ou de dependência, no DSM V o diagnóstico dos TUS enfatiza os prejuízos adjacentes do consumo de substâncias. Neste sentido, a classificação pode ser Leve (2 a 3 critérios são satisfeitos), Moderada (4 ou 5 critérios são satisfeitos) ou Grave (6 ou mais critérios são satisfeitos) e ser aplicada a 10 classes distintas de substâncias: álcool, cafeína, Cannabis, alucinógenos, inalantes, opioides, sedativos, hipnóticos e ansiolíticos, estimulantes (anfetamínicos, cocaínicos e outros), tabaco e outras substâncias (ou substâncias desconhecidas). O diagnóstico dos TUS pode ser aplicado a todas as 10 classes, exceto a cafeína, e é realizado a partir da satisfação de critérios comportamentais, fisiológicos e subjetivos, que podem variar na intensidade e na gravidade.

Evidências científicas, pré-clínicas e clínicas, sinalizam que a gravidade dos TUS pode estar relacionada à intensidade de alterações em circuitos cerebrais. Estas alterações podem persistir, mesmo após a desintoxicação, sobretudo em casos mais graves, o que poderia explicar as recaídas e a fissura intensa. Atualmente, acredita-se que quatro sistemas neuronais estejam particularmente envolvidos na fisiopatologia dos TUS (25):

- O sistema de recompensa cerebral (em especial o accumbens)
- O sistema de motivação (córtex orbitofrontal)
- O circuito de memória e aprendizagem (amígdala e hipocampo)

- A área de controle e planejamento (córtex pré-frontal e giro do cíngulo anterior)

Robinson e colaboradores (26, 27) propuseram uma hipótese biopsicológica da dependência, a “Teoria de Incentivo-Sensibilização”, que parte de um tripé de questionamentos fundamental para compreender o motivo de os TUS tornarem-se um problema de saúde pública em todo o mundo:

1. Por que os dependentes desejam drogas? Qual a base neurobiológica e psicológica da fissura por drogas?
2. Por que o desejo por drogas persiste mesmo depois de longos períodos de abstinência?
3. A fissura é atribuível a "gostar" de drogas (aos efeitos prazerosos subjetivos de drogas)?

Sob estes questionamentos discutiram quatro princípios explicativos:

1. Substâncias psicoativas com potencial dependógeno compartilham a capacidade de alterar a estrutura e o funcionamento cerebral;
2. Os principais sistemas cerebrais alterados na dependência são aqueles relacionados ao processo de incentivo motivação e recompensa;
3. As neuroadaptações tornam o sistema de recompensa cerebral hipersensíveis ("sensibilizados") a drogas e a estímulos associados às drogas;
4. Os sistemas cerebrais sensibilizados não são mediadores dos efeitos prazerosos ou eufóricos das drogas, mas sim do subcomponente de recompensa a “saliência de incentivo”.

1.3 Neurobiologia dos Transtornos por Uso de Substâncias

Avanços significativos referentes à psicobiologia dos TUS foram evidenciados nas últimas décadas, o que reforça a compreensão destes transtornos como uma doença crônica, caracterizada por alterações cerebrais (17, 28-31), e que, portanto, necessita de acompanhamento longitudinal. Compreender estas neuroadaptações pode auxiliar no estabelecimento de ferramentas analíticas que potencializem o entendimento da variabilidade interindividual na resposta a drogas de abuso e ao

tratamento (32-34). Identificar parâmetros biológicos, clínicos e neuronais, que refletem a gravidade dos TUS, pode acarretar implicações importantes na prática clínica, auxiliando na compreensão da fisiopatologia da doença, no rastreamento e identificação de casos mais graves, na avaliação da resposta ao tratamento e do potencial de recaída, possibilitando uma atenção mais individualizada e influenciando significativamente na elaboração de novas estratégias de prevenção e tratamento (30, 33).

Assim como em outras doenças, os pacientes com TUS apresentam sintomatologia heterogênea e respostas diferentes às intervenções terapêuticas (35). Identificar biomarcadores que apontem processos biológicos normais e processos patogênicos, relacionados à dependência, pode constituir instrumentos valiosos na clínica dos TUS, fornecendo medidas objetivas que permitam classificar pacientes, detectar subgrupos e prever a resposta ao tratamento, otimizando os resultados e gerando avanços clínicos significativos (35, 36).

Apesar de os estudos iniciais, sobre a neurobiologia dos TUS, terem enfatizado o impacto agudo das substâncias psicoativas (SPAs), atualmente, a compreensão dos TUS como uma doença crônica fez com que os esforços tenham sido direcionados ao entendimento das alterações neuroadaptativas agudas e de longo prazo. Isso objetiva compreender processos neuroquímicos que auxiliem no entendimento dos mecanismos genéticos, epigenéticos, celulares e moleculares, relacionados ao processo de transição do uso para a dependência e às recidivas crônicas que ocorrem mesmo após períodos longos de abstinência (25).

As drogas de abuso, em geral, compartilham uma característica comum: acarretam o aumento – direto ou indireto – da concentração de dopamina (DA) extracelular no sistema cerebral de recompensa, sobretudo no núcleo accumbens (NAC) estendendo-se, para a amígdala, induzindo assim o estado de excitação (25, 37-41). A DA é um neurotransmissor envolvido na modulação de diversas funções fisiológicas e no desempenho de comportamentos motivados dirigidos a metas. O aumento da sua concentração sináptica, em regiões específicas do cérebro, é associado à sensação de prazer e bem-estar desencadeada diante de certos comportamentos.

A DA também possui papel chave nos processos de aprendizagem, associada a recompensas e a definição de eventos de potencial gratificante. Diante de estímulos desencadeadores de prazer (alimentos, sexo, fuga em situações de perigo, entre outros), a ativação do sistema cerebral de recompensa é desencadeada pela expressão de DA, criando uma memória específica que estimula o organismo a buscá-la novamente (39, 42-45). Desta forma, a DA está fortemente relacionada ao reforço e à consolidação da memória, abrangendo, também, processos como: atenção, aprendizagem, cognição, humor, emoções, regulação do sono, alimentação, controle dos movimentos voluntários entre outros (39, 41, 46).

De acordo com Volkow e colaboradores, o uso de drogas provoca um efeito agudo de superliberação de DA, acarretando concentrações muito superiores aos níveis basais, com intensidade e duração que excedem aquelas desencadeadas por reforçadores naturais (44, 45, 47, 48). Postula-se que, este processo, ocasiona o desequilíbrio entre os circuitos dopaminérgicos relacionados à recompensa, prejudicando o condicionamento e as funções executivas de planejamento, regulação emocional e tomada de decisão (47, 49). Por outro lado, acredita-se que, as alterações na neurotransmissão cerebral de DA, adjacentes ao uso crônico de drogas, estariam relacionadas a um estado hipodopaminérgico que conduziria a desregulação dos circuitos de recompensa (50). Possivelmente, a repetição crônica da atividade da DA, no sistema cerebral de recompensa, pode gerar um desequilíbrio neuroquímico e conduzir a uma homeostase alterada, que desenvolverá neuroadaptações estruturais e funcionais permanentes na neuroplasticidade cerebral (51, 52), que por sua vez, estariam relacionadas ao processo de transição do uso recreativo para a dependência (53).

Volkow e colaboradores sugerem que, diferenças marcantes em circuitos cerebrais são evidenciadas em pacientes com dependência de substâncias se comparados com indivíduos sem dependência. De acordo com esse modelo, a dependência envolve outros circuitos além do sistema de recompensa (NAc, CPF, área tegmentar ventral-ATV), como: memória e condicionamento (amigdala, COF, hipocampo e estriado dorsal), controle executivo (córtex pré-frontal dorsolateral (CPFDL), giro cingulado anterior (GCA), córtex frontal inferior e COF lateral e motivação (COF medial para atribuição de saliência, GCA, ATV, estriado dorsal e

côrTEX motor). Durante o processo de adição, quando a saliência da droga nos circuitos de recompensa, motivação e memória superam os circuitos de controle, ocorre uma retroalimentação positiva - iniciada pelo consumo da droga e perpetuada pelo aumento da ativação dos circuitos de motivação e memória. Além da DA, diversos outros neurotransmissores estão envolvidos nessas neuroadaptações, incluindo glutamato, GABA, norepinefrina, fator liberador de corticotrofina e receptores opióides (49).

Mudanças na plasticidade encefálica podem ser adaptativas, quando associadas a um ganho de função, ou mal adaptativas, quando está associada à perda de funções e a consequências negativas – como é observado em dependentes de drogas. A liberação de DA, desencadeada por drogas de abuso, produz mudanças na forma como os neurônios integram a neurotransmissão excitatória e inibitória, dando início a uma cascata de sinalização complexa, que compromete a neuroplasticidade relacionada tanto a aprendizagem quanto ao desenvolvimento de comportamentos de procura pela droga (51). Este processo envolve neuroadaptações disfuncionais nos sistemas de: recompensa, saliência, motivação, controle inibitório, função executiva, memória e condicionamento, circuitos modulados por vias dopaminérgicas (25, 54, 55).

Evidências apontam para um papel importante das NTs cerebrais nas neuroadaptações induzidas pelo uso de substâncias, podendo estar associadas ao desenvolvimento da dependência, fissura, dano cognitivo e predisposição para recaída (56, 57). O estudo de biomarcadores acende a possibilidade de melhor compreender a fisiopatologia dos TUS, possibilitando prever a gravidade da doença e avaliar o prognóstico e a construção de métodos terapêuticos mais individualizados (58).

1.4 Biomarcadores

Embora existam instrumentos para mensurar se o indivíduo fez uso de substâncias psicoativas – os drogômetros –, esta mensuração limita-se à identificação aguda. Apesar dos avanços na pesquisa, evidenciados nas últimas décadas, ainda não existe nenhum biomarcador que possibilite estimar, de forma

precisa, a trajetória da dependência, os subtipos clínicos, nem a eficácia do tratamento; tampouco, que possa agregar alguma informação preditiva ou que reflita processos neurobiológicos associados com a gravidade da doença (36, 59).

Biomarcadores são características objetivamente observáveis, medidas e avaliadas como indicadores de processos biológicos normais, patogênicos ou respostas farmacológicas para uma intervenção terapêutica (60). A identificação e a utilização clínica de biomarcadores apropriados pode possibilitar uma compreensão mecanicista do processo patológico, constituindo uma ferramenta de medição clínica precisa que possibilite determinar a progressão da doença, os efeitos das intervenções e a resposta clínica, permitindo remodelar o tratamento e potencializar resultados terapêuticos (33, 60).

Existem diversos tipos de biomarcadores: diagnósticos, preditivos, mecanicistas, prognósticos, que podem ser quantificados de diferentes formas (fluidos corporais, soro, plasma, cabelo, urina, sangue, unhas, exames e neuroimagem). A quantificação de substâncias em fluidos corporais, como urina, saliva e sangue, são os biomarcadores mais precisos evidenciados na atualidade. Por outro lado, os dados sobre biomarcadores, com capacidade de prever desfechos clínicos ou processos que se relacionam com a neurobiologia da doença, ainda são incipientes. De acordo com a diretora do NIDA, Nora Volkow, há uma necessidade urgente de biomarcadores que reflitam a exposição crônica a fármacos, bem como, que prevejam ou se correlacionem com as trajetórias de doença e as respostas ao tratamento (36). Neste sentido, os fatores neurotróficos têm sido alvo de investigações na fisiopatologia de diversos transtornos, incluindo os TUS (61, 62).

1.5 BDNF

As NTs (NT) têm papel essencial na regulação de distintas atividades celulares no Sistema Nervoso Central (SNC), como a expressão gênica, o crescimento, a diferenciação e a sobrevivência celular, a apoptose e a plasticidade sináptica (63). O BDNF (64) é a NT com papel na plasticidade sináptica melhor estabelecido na literatura (65). Estima-se que esteja relacionado à mediação dos

principais processos dependentes de estímulo externo, como aprendizado, experiências e memórias, incluindo efeito das substâncias de abuso (66). As NTs, em especial o BDNF, parecem estar implicadas na base fisiopatológica de diversas doenças neurodegenerativas (67-70) e psiquiátricas (70-77)

O BDNF é considerado a principal neurotrofina cerebral, sua produção é realizada principalmente pela glia e pelos núcleos neuronais, sendo altamente expresso na amígdala, neocôrtex, hipocampo e cerebelo, áreas relacionadas com funções cognitivas (78, 79). Também, está relacionado à modulação de diversas funções como o crescimento celular, maturação, nutrição, diferenciação, conectividade sináptica, crescimento e integridade neuronal, modulação da transmissão, reparo neuronal, além de processos de neurogênese e neurodegeneração (71, 80-82). O BDNF periférico tem sido amplamente utilizado em pesquisas clínicas, uma vez que parece atravessar livremente a barreira hemato-encefálica e seus níveis no soro e no plasma estão fortemente correlacionados com as concentrações de BDNF encontradas no SNC (83-87). Apesar de estudos prévios apontarem para a associação entre níveis periféricos e cerebrais de BDNF (86-88), em humanos, é difícil estimar de quais áreas encefálicas o BDNF periférico é proveniente, o que torna a transposição dos achados pré-clínicos complexa, compondo uma limitação importante.

Além disso, as matrizes biológicas usados para dosagem do BDNF periférico tem mostrado heterogeneidade. Os níveis do BDNF no soro humano são de 20 - 50 vezes superiores aos níveis no plasma (89, 90). O BDNF do plasma é proveniente, principalmente, dos linfócitos, das células endoteliais, dos linfonócitos e das células periféricas não neuronais, incluindo as células do músculo liso vascular (91, 92). Já o BDNF sérico, mimetiza o total de BDNF armazenado e liberado pelas plaquetas durante o processo de coagulação (93, 94). Tendo em vista que as plaquetas contêm a maior parte do BDNF circulante no sangue, o soro parece refletir melhor a quantidade do BDNF no sangue total, constituindo um marcador mais fidedigno (76, 93).

1.6 BDNF nos Transtornos por uso de substâncias

Estudos pré-clínicos apontam o envolvimento do BDNF na neuroadaptação induzida por drogas, relacionada à transição do uso para a dependência, fissura, recaída e sensibilização à droga (95, 96); isso é evidenciado, principalmente, nos estudos que avaliaram o álcool e os psicoestimulantes (97, 98). Apesar disso, os resultados, de uma forma geral, são controversos. Enquanto a administração prolongada de nicotina, por exemplo, ocasionou o aumento da expressão encefálica do BDNF no cérebro de ratos (99), as anfetaminas diminuíram os níveis de BDNF no hipotálamo e nas regiões corticais (100), e a morfina gerou um aumento imediato na expressão da proteína BDNF no VTA, mas após a retirada, a longo prazo, verificou-se de uma diminuição(101).

Uma revisão verificou que os efeitos do álcool sobre o BDNF cerebral de ratos são controversos entre diferentes estudos (98). As pesquisas pré-clínicas, com cocaína, demonstraram que a administração de BDNF em distintas áreas cerebrais acarreta efeito direto no comportamento de busca por cocaína, todavia, pode ter efeitos opostos dependendo da região cerebral. Infusão de BDNF na ATV e no NAc, durante a abstinência precoce de cocaína, potencializou a procura pela droga e a recaída (102, 103). Em contraposição, a administração de BDNF no córtex pré-frontal de ratos, no último dia de autoadministração de cocaína (realizada por 10 dias) reduziu a procura por cocaína (104).

Acredita-se que o uso de SPA acarrete neuroadaptações e que elas estejam associadas com a reinstalação da busca pela droga, e ocorrem, principalmente, na via CPF-NAc, na qual o BDNF é expresso. Os neurônios piramidais corticais, que saem do CPF, são a principal fonte de BDNF dentro do estriado, incluindo o NAc. A administração aguda de cocaína induz a expressão de RNAm de BDNF no CPFm, córtex cingulado e estriado, que pode representar os estágios iniciais da plasticidade cortical induzidas pelos psicoestimulantes. Após a administração repetida de cocaína, o BDNF permanece aumentado nas estruturas prosencefálicas (105). Na abstinência precoce (22h), há diminuição do BDNF no CPF dorsomedial, o que não ocorre na abstinência tardia (21 dias) (96). Além disso, após um período de auto-administração de cocaína, o BDNF está aumentado nas estruturas mesolímbicas, incluindo ATV, NAc e amígdala em períodos prolongados de abstinência (106). Considerando que o BDNF é transportado nas projeções glutamatérgicas do CPF

para o NAc, e que esta via regula a recaída para a busca de drogas, a variação da expressão do BDNF nesta via pode constituir um componente crítico na neuroplasticidade induzida pela cocaína nos neurônios corticais e mesolímbicos (96).

Ao avaliar os níveis de BDNF em dependentes de álcool, em uso ativo, algumas investigações encontraram níveis semelhantes aos controles no *baseline* (107-109), todavia, também há relatos de aumento (61, 110) e redução (111, 112). Já nos dependentes de cocaína, dois estudos recentes avaliaram o BDNF pré-desintoxicação, tendo encontrado níveis diminuídos em relação aos controles (113, 114). Em relação às outras drogas, nos usuários de metanfetamina e de heroína, o nível de BDNF foi encontrado tanto aumentado (115, 116) quanto diminuído (117, 118) em relação a controles. Nos usuários crônicos de maconha, o BDNF não diferiu em relação aos controles (119) e nos usuários de ecstasy foi encontrado elevado (120). Analisados em conjunto, estes estudos revelam que os níveis de BDNF, em indivíduos dependentes de substâncias não abstinentes, são muito variados, tanto entre drogas diferentes quanto entre usuários da mesma substância. Cabe salientar que, a maioria dos estudos foi com tamanhos amostrais pequenos e não há uma boa descrição de como os controles foram selecionados, fatos que podem estar relacionados com achados divergentes.

Mais recentemente, os resultados da variação do BDNF durante a abstinência de cocaína/crack e de álcool têm sido associados à manutenção da abstinência, reforçando o possível papel do BDNF como marcador prognóstico na dependência química (108, 121). O BDNF também tem sido associado com a gravidade dos problemas com cocaína/crack e álcool (122, 123). Estes resultados reforçam a hipótese de que o BDNF é um potencial indicador para a gravidade do uso de drogas e do prognóstico. Portanto, o entendimento do comportamento desta NT pode auxiliar na elucidação do porquê os tratamentos disponíveis funcionam para alguns pacientes e não para outros.

Em dependentes de álcool e cocaína, o BDNF foi avaliado também na abstinência e como preditor de recaída. Durante a abstinência precoce de cocaína (14 dias), o BDNF aumentou durante a desintoxicação e o valor ao final das duas semanas foi correlacionado positivamente com sintomas de abstinência, de

depressão e de ansiedade e com fissura, tendo esse estudo avaliado apenas 23 pacientes e 46 controles (113). Esse achado sugere que níveis maiores de BDNF na abstinência precoce seriam encontrados em indivíduos mais sintomáticos e mais graves. Nessa mesma linha, o valor do BDNF, após 3 semanas de abstinência de cocaína, foi maior no grupo que recaiu em relação ao grupo que se manteve abstinente após 90 dias da alta hospitalar (61). Em dependentes de álcool, o BDNF, avaliado 6 meses após um período de desintoxicação, foi maior nos que estavam abstinentes em relação aos que recaíram, sendo a proporção do aumento também maior nos abstinentes, quando avaliados em relação ao valor do BDNF no primeiro dia da internação (108).

Investigações do nosso grupo de pesquisa apontaram que, a redução no BDNF sérico foi associada à gravidade do uso de crack, além disso, o nível de BDNF estava reduzido na admissão, mas na alta estava semelhante aos controles (122). Outra investigação revelou que, os níveis de BDNF aumentaram durante a abstinência precoce de crack, em uma correlação inversa com o número de pedras de crack usadas nos últimos 30 dias e aos anos de uso de crack (114).

Outro estudo, publicado recentemente, sugeriu uma associação entre níveis mais elevados de BDNF e melhores resultados clínicos, em usuários de crack-cocaína, após desintoxicação (124). Neste sentido, o aumento do BDNF durante a abstinência poderia estar associado à capacidade regenerativa do cérebro após a interrupção do consumo. Esses resultados encontrados são diferentes daqueles descritos por Corominas e D'As, para usuários de cocaína, que associaram o aumento do BDNF durante a abstinência precoce a maiores índices de recaída (61, 121).

Estes resultados mostram a existência de discrepâncias entre os estudos, havendo relatos da redução, do aumento e da não alteração do BDNF em dependentes de drogas. Se considerarmos o fato de estarem ou não abstinentes a controvérsia é ainda maior. Alguns estudos têm ponderado que o BDNF periférico prediz resposta ao tratamento e ao prognóstico, mas os resultados permanecem discrepantes. Ressalta-se que, as inconsistências nos resultados, podem ser decorrentes de diferenças nas características da população estudada ou da ausência de poder estatístico, devido ao reduzido

tamanho amostral. Assim, o papel do BDNF nos TUS permanece sendo pouco explorado.

Uma técnica reconhecida, usada para resolver discrepâncias entre estudos transversais, é a meta-análise; um método quantitativo de combinar os resultados de estudos independentes, visando potencializar o poder estatístico e permitir conclusões mais consistentes. Este método aumenta o poder para se distinguir entre pequenos tamanhos de efeito e ausência de efeito. Além disso, pode ajudar a determinar se a variação de efeito entre os estudos é devida, apenas, à flutuação estatística esperada ou a diferenças reais na amostra utilizada. Uma análise de meta-regressão, um método estatístico mais sofisticado, pode avaliar fatores moderadores associados a discrepâncias entre os estudos.

2 Objetivo geral:

Avaliar os níveis periféricos de BDNF em pacientes com Transtornos por uso de substâncias, lícitas e ilícitas, exceto tabaco, comparando-os com controles saudáveis a partir de uma revisão sistemática da literatura

2.1 Objetivos específicos:

Verificar se há diferenças nas concentrações séricas e plasmáticas de BDNF de acordo com o tipo e a classe da substância psicoativa utilizada;

Avaliar se a retirada da droga (abstinência) está relacionada com alterações nos níveis periféricos de BDNF;

Avaliar potenciais fatores moderadores (sexo, idade, idade de início do consumo da substância, tempo de uso, e tempo de abstinência);

Confirmação de submissão



Felipe Ornell <felipeornell@gmail.com>

Fwd: Submission Confirmation

1 mensagem

Lisia Von Diemen Idiemen <Idiemen@hcpa.edu.br>
Para: Felipe Ornell <felipeornell@gmail.com>

29 de março de 2017 11:02

----- Forwarded message -----

From: Biological Psychiatry <eesserver@eesmail.elsevier.com>
Date: 2017-03-29 10:57 GMT-03:00
Subject: Submission Confirmation
To: Idiemen@hcpa.edu.br, lisiavd@gmail.com

Dear Lisia,

We have received your article "Brain-derived neurotrophic factor in Substance Use Disorders: a systematic review and meta-analysis" for consideration for publication in Biological Psychiatry.

Your manuscript will be given a reference number once an editor has been assigned.

To track the status of your paper, please do the following:

1. Go to this URL: <https://ees.elsevier.com/bps/>

2. Enter these login details:

Your username is: Idiemen@hcpa.ufrgs.br

If you need to retrieve password details, please go to: http://ees.elsevier.com/bps/automail_query.asp

3. Click [Author Login]

This takes you to the Author Main Menu.

4. Click [Submissions Being Processed]

Thank you for submitting your work to Biological Psychiatry.

Kind regards,

Elsevier Editorial System
Biological Psychiatry

3 Artigo

Brain-derived neurotrophic factor in Substance Use Disorders: a systematic review and meta-analysis.

Felipe Ornell^{ab}, Fernanda Hansen^{ac}, Felipe Barreto Schuch^{cd}, Fernando Pezzini Rebelatto^a, Ana Laura Tavares^a, Juliana Scherer^{ab}, Andrei Garziera Valerio^{ab}, Flávio Pechansky^{ab}, Felix Henrique Paim Kessler^{ab}, Lisia von Diemen^{ab}

^aCenter for Drug and Alcohol Research and Collaborating Center on Alcohol and Drugs – HCPA/SENAD, Hospital de Clínicas de Porto Alegre, Federal University of Rio Grande do Sul, Rua Professor Álvaro Alvim, 400, 90420-020, Porto Alegre, RS, Brazil.

^bPostgraduate Program in Psychiatry and Behavioral Science, Federal University of Rio Grande do Sul, Porto Alegre, RS, Brazil.

^cDepartment of Biochemistry, Institute of Biological Sciences, Federal University of Rio Grande do Sul, Porto Alegre, RS, Brazil.

^cHospital de Clínicas de Porto Alegre, Porto Alegre, RS, Brazil.

^dCentro Universitário La Salle, Porto Alegre, RS, Brazil.

Corresponding Author: Felipe Ornell

Phone: +55 51 33596488

E-mail address: fornell@hcpa.edu.br'

Address: Rua Professor Álvaro Alvim, 400, 90420-020, Porto Alegre, RS, Brazil.

Funding: Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES)

Keywords: Biomarker, Substance Use Disorder, Brain-derived neurotrophic factor, Meta-analysis

Abstract: 231 words

Text: 3997

Figures: 6

Tables: 5

Supplemental information: 2

Abstract

Background: Changes in brain-derived neurotrophic factor (BDNF) are associated with several neurodegenerative and psychiatric disorders. It is not clear, however, whether BDNF levels are altered in substance use disorders (SUDS) **Methods:** We conducted a systematic search of electronic databases to identify studies comparing peripheral - plasma or serum - BDNF levels in adults with SUDS versus healthy controls. Thirty studies were included in the meta-analysis, accounting for 1698 participants with SUDS and 1363 controls. **Results:** The overall analysis found no significantly different BDNF levels of cases versus controls ($SMD=-0.11$, 95%CI -0.30 to 0.15, $p=0.39$, $I^2=94.02$). Subgroupanalysis revealed lower levels of BDNF in alcohol users compared to controls ($SMD=-0.45$, 95%CI -0.89 to -0.01, $p=0.04$, $I^2=90.54$). Studies using serum or plasma BDNF samples shown different, and sometimes, antagonistic results. After adjusting for publication bias, the resoults reveals that serum BDNF in current users was reduced ($SMD=-0.51$, 95%CI -1.12 to -0.08), but similar to controls in withdrawn. Metaregression analysis revealed that years of drug use, age, gender and length of withdrawn moderate the effects of drug use on peripheral BDNF levels. **Conclusions:** Peripheral BDNF is decreased in alcohol use disorders. However, this effect is not significant across difrent drugs types. Time using the drug, age, gender and the length of withdraw modifies the effects of drug abuse on BDNF levels, indicating that BDNF could be related with the severity and progression of drug dependence, and perhaps be a state biomarker, as already described for other psychiatric and neurodegenerative disorders.

Introduction

Drug addiction consists in a chronic disorder characterized by a set of cognitive, behavioral and physiological symptoms, by the continuity of use despite negative consequences and by high rates of relapse. Regardless of type of drug used, three main characteristics are observed among addicts: 1) Loss of control over use; 2) Tolerance; and 3) Withdrawal symptoms and adverse emotional states when use is interrupted^{1, 2}. This can derive from dysfunctional alterations in brain systems, mainly involving memory, reward, motivation and executive functions systems, resulting in compulsion, deficits in decision making, impulsivity and a reduction in inhibitory control^{3, 4}. Although it is postulated that chronic use of psychoactive-substances (PAS) entails changes in brain function and structure, leading to dysfunctional neuroadaptations – transitory or permanent –, the time of use and amount of drug necessary to provoke these alterations are unknown⁵. Grasping these neuroadaptations may allow the establishment of analytical tools that enhance the understanding of interindividual variability on drug abuse and treatment responses⁵⁻⁸.

Significant advances regarding the psychobiology of addictive disorders were evidenced in previous decades, reinforcing the comprehension of addiction as a chronic disease, characterized by structural and functional alterations in different brain systems⁸⁻¹². Brain-derived neurotrophic factor (BDNF) has been studied as a potential biomarker to understand the physiopathology of several psychiatric disorders, including Substance Use Disorder (SUDS)^{13, 14}. BDNF is the most abundant neurotrophin found in the brain and it is related to cell growth, differentiation, connectivity and synaptic neuroplasticity, neurotransmission

modulation and neuronal repair, besides neurogenic and neurodegenerative processes^{15, 16}, learning and memory¹⁷. Pre-clinical and clinical evidence indicate that neurotrophins are involved in the neuroadaptation process caused by PAS use as well as in different process related to mediation of behavioral effects, including drug sensitization, craving and relapse¹⁸⁻²². Peripheral BDNF has been widely assessed in clinical studies since it freely crosses the blood-brain barrier and its peripheral concentration is strongly correlated with concentrations found in central nervous system (CNS)^{23, 24}.

Previous studies that evaluated BDNF levels in drug addicts evidenced different and even controversial results, indicating that this variation may be related to the type of drug consumed, the pattern of use at the moment of the study and dependence stage²⁵⁻³³. However, published studies mostly used small and heterogeneous samples and evaluated, predominantly, only one type of drug, preventing the establishment of a concentration range of this biomarker in patients with a SUD of a different, or of multiple substances. There is not of authors' knowledge the existence of a systematic review that has meta analyzed the difference between BDNF levels in people with SUDS versus controls. Therefore, the present study aims to: 1) compare serum and plasma peripheral BDNF levels between subjects with SUDS and healthy controls; 2) evaluate whether this difference occurs in different drugs types, subtypes, withdrawn and blood fraction through subgroup analysis and 3) explore, using meta-regressions, potential moderating factors.

Methods and Materials

The protocol developed for this meta-analytic review followed the

recommendations of the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement³⁴. Each element of the search, selection of eligible studies (FO and AGV) and data extraction (FPR and ALT) were performed by two authors. Disagreements were resolved through consensus and with the opinion of a third author (FHPK).

Search strategy

We conducted a systematic search of all potentially eligible references without language restrictions, using PubMed/MEDLINE, EMBASE and PsycINFO computerized databases. The search strategy used was (addiction OR drug abuse OR dependence OR opioids OR heroin OR cocaine OR crack OR inhalants OR methamphetamine OR stimulants OR cannabis OR ecstasy OR club drugs OR alcohol OR alcohol* OR substance-related disorders OR SUDS) AND (BDNF OR brain derived neurotrophic factor OR neurogenesis) NOT (animal OR rat OR mouse OR mice). The last search was performed in June ninth 2016. This search strategy was augmented by tracking manually the reference list of all retrieved papers to identify additional eligible papers.

Study selection

The inclusion criteria were: (1) studies that evaluated individuals with diagnosis of SUDS of alcohol or illicit drugs meeting Diagnostic and Statistical Manual for Mental Disorders or International Classification of Diseases diagnostic; (2) studies comparing BDNF levels between people with SUDS and healthy volunteers or clinical trials using baseline data before drug treatment; (3) studies that measured peripheral BDNF levels; Exclusion criteria were (1) studies that evaluated samples

with baseline clinical comorbidities (e.g: depression, HIV, Parkinson's disease, alzheimer and dementia); (2) genetic studies without information on peripheral BDNF levels; and (3) papers written in the format of letter, poster, review, case reports, opinion or commentary. The authors consensually agreed on the final inclusion of references.

Data extraction

Two reviewers (FPR and ALT) independently extracted data from controls and SUDS group (sample size, mean and standard deviation) from all articles to prevent potential errors. Discrepancies in data entry were double-checked by the reviewers with the original published data and a consensus was reached. We extracted information regarding primary drug, psychiatric comorbidities, scales and/or questionnaires performed to evaluate psychiatric symptoms or disorders, BDNF levels, type of peripheral sample used for BDNF levels measurement and withdrawn length at time of BDNF levels measurement. Information about sex, age, age of first use, years of use, tobacco use, body mass index (BMI), and clinical diseases were also extracted. Furthermore, corresponding authors of included articles were contacted by e-mail in at least two different occasions whenever necessary data were unavailable, requesting the provision of data. In the cases multiple reports pertained to the same participants, we included only the largest data set. When data were available only in graphs, we extracted the data using the Plotdigitalizer software, web version³⁵.

Assessment of methodological quality

The quality of the 36 studies included in the systematic review was evaluated using the 9-item Newcastle-Ottawa Scale (NOS) for case-control studies³⁶. However, the item 3 ("Equivalent non-response rate between groups") was not used in quality assessment because of the proposal of this review. For comparability questions, age was set a priori as the most important matching or adjustment factor. Moreover, other 2 articles were evaluated by Cochrane Collaboration tool using the 7-bias risk assessment of Randomized Clinical Trials, using baseline data/information³⁷.

Meta-analysis

We conducted random effect meta-analyses due to the anticipated heterogeneity across studies. The analyses were performed with the Comprehensive Meta-Analysis software (CMA, Version 3). The meta-analysis were conducted in the following sequence. First, we conducted a comparative meta-analysis investigating differences on BDNF levels of drug users versus controls using the standardized mean deviation (SMD) together with the 95% confidence intervals (CI) of the effect size. Second, we calculated the subgroup differences according to the 1) drug subtype, 2) withdrawn (we considered in the withdrawn groups those with a minimum of 6 days), 3) drug classification as stimulant (cocaine, crack and methamphetamine), depressant (alcohol, heroine, opiates and ketamine) or hallucinogen (cannabis and amphetamine derivative (\pm)-3,4-methylenedioxymethamphetamine (MDMA, ecstasy)), 4) biological matrix used to measure BDNF (serum or plasma). Third, we investigated potential moderators of BDNF levels with meta-regression analyses. The potential moderators of interest were mean age, % of males, withdrawn length and years of drug use. We assessed heterogeneity using the I^2 statistics for each analysis. Publication bias was verified with the Begg-Mazumdar Kendall's tau and Egger tests.

Trim and fill analyses were conducted to remove the most extreme small studies from the positive side of the funnel plot, and recalculated the effect size at each interaction until the funnel plot was symmetric about the (new) effect size.

Results

Initially, 925 studies were localized as potentially relevant. Following the application of the eligibility criteria, 56 studies were included for review. After reading the preliminary included papers in full and tracking reference lists for possible missing papers, 38 articles reached the eligibility criteria and were included in this systematic review and 30 of them were also included in the meta-analysis (Figure 1).

Study and participants' characteristics

Across 47 arms of 30 unique studies, there were 2315 blood samples of 1698 participants with SUDS (79% males, mean age of 36.50 years) and 2049 blood samples of 1363 controls (75% males, mean age of 35.24 years) included. The sample size of drug users ranged from 9³⁸ to 179³⁹. Fourteen arms evaluated BDNF levels in alcohol (n=600), ten in cocaine (n=532), nine in heroin (n=472), two in cannabis (n=35), five in methamphetamine (n=355), two in ketamine (n=110), and one in MDMA (n=23) users. Twenty arms have blood samples in participants with current/active drug use (n=1119) and other 20 arms in withdrawn participants (n=1102). Twenty-nine arms evaluated participants using depressant drugs (n=1370), 15 in stimulant drugs (n=887) and three on hallucinogen drugs (n=58). Nine arms used data from plasma samples to measure BDNF levels (n=665) while 38 used serum samples (n=1650). Further details of the included studies are summarized in Table 1.

Quality of studies

Quality assessment of the eligible studies using the NOS (Supplemental information).

Four studies scored 8 of 8 possible points ^{25, 29, 40, 41}, 12 scored 7 of 8 possible points ^{27, 28, 30, 39, 42-49}, 13 scored 6 of 8 possible points ^{25, 50-61}, and 7 scored 5 of 8 possible points ^{26, 40, 42, 62-65}. The quality of the studies was most often compromised by the representativeness of the cases, following by the equivalence in ascertainment for cases and controls. The absence of information about how was the selection criteria of the sample and the validated instruments used to assess psychoactive SUDS without subsequent clinical evaluation was the main compromised points.

Quality assessment of the eligible studies using the Cochrane Collaboration tool (Supplemental information).

None of the 2 studies presented high risk of bias. Nonetheless, they have uncertain risk of bias for allocation concealment and selective reporting³⁸ and one of them also for random sequence generation⁶⁶.

Meta-analysis of overall comparative BDNF levels

Forty-seven studies, including 2315 blood samples of participants with drug use and 1698 controls were pooled and found no significant differences in BDNF levels ($SMD=-0.11$, $95\%CI -0.30$ to 0.15 , $p=0.39$, $I^2=94.02$) (Figure 2). No significant publication bias was identified in Begg and Mazumdar ($\tau=0.02$, $p=0.79$) or Egger tests ($intercept=1.03$, $p=0.60$). However, the Duval and Tweedie trim and fill technique adjusted the effect size for 0.22 ($95\%CI -0.07$ to 0.52) trimming nine

studies.

Subgroup analyses

We investigated BDNF levels on plasma or serum separately, as different subgroups (table 3). For each matrix, we investigated the specific role of current substance use status (current use/withdrawn), and the drug subtype – Alcohol (Figure 3), Heroin (Figure 4), Crack/cocaine (Figure 5), Cannabis, Ketamine, MDMA/Ecstasy or Methamphetamine (Figure 6). Following, we investigated the role of drug action and the drug subtype for people with current use or withdrawn, separately, for both serum and plasma BDNF levels. The subgroup analyses are detailed below.

Serum BDNF levels

Twenty-five studies, including 1650 blood samples of participants with SUDS and 1603 controls were pooled and no significant differences in BDNF levels were found ($SMD=-0.21$, $95\%CI -0.51$ to 0.08 , $p=0.16$, $I^2=93.46$). No significant publication bias was identified in Begg and Mazumdar ($\tau=-0.03$, $p=0.79$) or Egger tests ($intercept=-0.69$, $p=0.74$). However, the Duval and Tweedie trim and fill technique adjusted the effect size for 0.16 ($95\%CI -0.16$ to 0.49) trimming eight studies. The detailed analysis of BDNF levels can be seen in Table 2, but they will be briefly summarized below.

Analysis of subgroup of serum BDNF

Lower BDNF levels were found for users of depressant drugs ($SMD=-0.38$, $95\%CI -0.76$ to -0.03 , $p=0.04$, $I^2=93.02$) and for users of alcohol ($SMD=-0.45$, $95\%CI$

-0.89 to -0.01, $p=0.04$, $I^2=90.54$) and higher levels were found for users of MDMA/Ecstasy ($SMD=5.72$, 95%CI 4.35 to 7.09, $p>0.001$, $I^2=0$). After adjusting for publication bias, decreased levels were found for serum BDNF levels in people with current/actual use of drugs ($SMD=-0.51$, 95%CI -1.12 to -0.08), with one study trimmed, and the difference between users of alcohol and controls was shortened to -0.73 (95%CI -1.22 to -0.25) after trimming 3 studies (Table 2).

Serum BDNF levels in withdrawn participants

No significant effects were found on BDNF serum levels of withdrawn participants. The results remain not significant even analysing drugs with different actions (stimulant or depressant) or at a drug subtype level.

Serum BDNF levels in participants with active use

Lower BDNF levels were found in alcohol ($SMD=-0.64$, 95%CI -1.16 to -0.13, $p=0.01$, $I^2=86.84$) and crack/cocaine ($SMD=-0.96$, 95%CI -1.26 to -0.66, $p=0.001$, $I^2=0$) users, whilst increased BDNF levels were found for users of Ketamine ($SMD=1.37$, 95%CI 0.53 to 2.12, $p=0.001$, $I^2=0$). The publication bias adjusted effect size for people with alcohol use disorder and active use was of -0.97 (95%CI -1.57 to -0.32), after trimming two studies.

Plasma BDNF levels

Eight studies, including 665 blood samples of participants with drug use and 446 controls, were pooled and no significant difference was found in BDNF levels ($SMD=0.26$, 95%CI -0.36 to 0.90, $p=0.40$, $I^2=95.55$). Significant publication bias was identified at the Begg and Mazumdar ($\tau=0.52$, $p=0.04$) and the Egger tests

(intercept=-10.65, p=0.04). However, the Duval and Tweedie trim and fill technique was not adjusted. The detailed analysis of plasma BDNF levels can be seen in Table 3, but they follow briefly summarized below.

Analysis of subgroups of plasma BDNF

Increased BDNF levels were found for users of stimulant drugs (SMD=-0.59, 95%CI -0.12 to 1.05, p=0.01, I²=80.36) and for users of methphetamines (SMD=0.64, 95%CI 0.24 to 1.04, p=0.002, I²=0) and decreased levels were found for users of Heroin (SMD=-1.01, 95%CI -1.27 to -0.75, p>0.001, I²=0). After adjusting for publication bias, the effect size of stimulant drugs has slightly decreased to 0.47 (95%CI 0.62 to 0.88), after trimming one study (Table 3).

Plasma BDNF levels in withdrawn participants

Decreased BDNF levels were found for depressant drugs (SMD=-0.66, 95%CI -1.32 to --0.00, p=0.04, I²=66.34) and for alcohol (SMD=-0.96, 95%CI -1.26 to -0.66, p=0.001, I²=0), whilst increased BDNF levels were found for users of Methamphetamine (SMD=0.64, 95%CI 0.24 to 1.04, p=0.002, I²=0). No analysis of this subgroup was adjusted for publication bias.

Plasma BDNF levels in participants with active use

Plasma BDNF levels are increased in users of stimulant drugs (SMD=0.97, 95%CI 0.48 to 1.47, p>0.001, I²=0). This subgroup reflects based on the findings of one unique study in users of crack/cocaine. Lastly, users of Heroin presented decreased BDNF plasma levels (SMD=-1.01, 95% CI -1.27 to -0.75, p>0.001, I²=0). No analysis of this subgroup was adjusted for publication bias.

Meta-regressions of potential moderators

We have identified some moderators of the effect of drug use on BDNF serum levels, but not in plasma. In serum samples, sex ($k=38$, $\beta=-0.02$, 95%CI -0.04 to -0.01, $p=0.0008$, $R^2=0.07$), age ($k=38$, $\beta=-0.04$, 95%CI -0.91 to -0.00, $p=0.02$, $R^2=0.02$), years of use ($k=20$, $\beta=-0.06$, 95% CI -0.12 to -0.00, $p=0.04$, $R^2=0.03$) and the length of withdrawn ($k=15$, $\beta=-0.86$, 95%CI -1.71 to -0.02, $p=0.04$, $R^2=0.10$) moderated the difference of BDNF levels between drug users and healthy controls. The interactions between sex and age ($k=38$, intercept=3.11, 95%CI 1.24 to 4.98, $p=0.001$, $R^2=0.04$), sex, age and years of use ($k=20$, intercept=4.35, 95%CI 1.91 to 6.79, $p=0.0005$, $R^2=0.11$), but not the interaction between sex, age, years of use and length of withdrawn were significant ($k=8$, intercept=5.04, 95%CI -1.75 to 11.84, $p=0.14$, $R^2=0.27$). The full details of the models tested can be found in Table 4.

Discussion

This was the first meta-analysis to assess peripheral BDNF levels in a population with SUDS compared to healthy controls. The main findings of this study were: 1) Peripheral BDNF levels between serum and plasma are divergent; 2) Patients with active drug use show lower serum BDNF levels when compared with controls; 3) There is no difference between BDNF levels of withdrawn individuals and controls in any of the substance groups, except for alcohol at plasma sample; 4) There are differences in peripheral BDNF levels among types of PAS; 5) In depressant drugs, especially alcohol, results are more robust, both in serum and plasma analyses; 6) Meta-regressions showed that sex, age, length of withdrawn and years of drug use modifies the difference in BDNF levels between patients with

SUDS and controls. As a whole, our results suggest that BDNF could be a state-marker in SUDS.

One of the main results of this review and meta-analysis revealed differences in BDNF peripheral levels between serum and plasmatic biological matrices. Only two out of the thirty studies included in this investigation evaluated both biological matrices simultaneously and observed intra-individual and intra-group variations^{41, 52}. This difference has already been demonstrated in previous studies with other psychiatric disorders⁶⁷⁻⁶⁹. An anterior meta-analysis of subjects with Major Depression verified the reduction in BDNF serum levels and the absence of significant differences in the plasma, showing no correlation of BDNF levels between these biological matrices⁶⁸. Several hypotheses may help the comprehension of the discrepancy in peripheral dosages between the biological matrices. In general, the origin of BDNF in blood is not completely known – although the brain is believed to be the main source of circulating peripheral BDNF –, it is also produced in other regions⁷⁰⁻⁷⁶. While plasma BDNF derives mainly from lymphocytes, endothelial cells, monocytes and non-neural peripheral cells, including vascular smooth muscle cells^{75, 76}, serum BDNF is probably referred to total BDNF stored and released by platelets during blood clotting process^{77, 78}, generating a concentration of serum BDNF up to 50 times higher than the plasmatic one^{77, 79}. Besides that, while plasma BDNF has a half-life of less than one hour, platelets circulate for up to 11 days^{80, 81}. Considering that platelets contain the majority of the circulating BDNF in the blood, some investigations point out that serum levels seem to better reflect the amount of BDNF in the whole blood, constituting a more reliable marker^{77, 78, 82, 83}.

BDNF measurement in both plasma and serum BDNF is also subject to variability during detection due to biological, methodological and analytical factors⁸⁴.

The methodology applied to the collection and storage protocol may affect the samples stability^{68, 82, 85, 86}. In plasma, the time between puncturing and sample processing, centrifugation condition^{77, 85} and anticoagulant used can also interfere with the dosage. While Ethylenediaminetetraacetic acid (EDTA) would cause an increase in the detection of BDNF levels as a result of platelet release^{78, 87}, heparin would cause a decrease up to 60% in plasma detection⁸⁸.

The reduction of serum BDNF during active drug use found in our analyses corroborates with prior data demonstrating lower BDNF levels in manic and depressive episodes and in schizophrenia compared to euthymic subjects and healthy controls, showing that this neurotrophin may be a state-marker in psychiatric diseases.^{67, 89-93} The lack of significant difference in BDNF levels between cases and controls during withdrawal is also in accordance with previously described evidences in other psychiatric disorders where BDNF increased after withdrawn^{67, 90}. These results suggest that BDNF could vary between different stages of drug use and be associated with support the hypothesis that BDNF may be a stage biomarker of the disorder, capable of signaling the cumulative neurological damage of neuroprogressive nature of the psychiatric conditions⁹⁴⁻⁹⁷. In this sense, it is known that during the course of mental disorders the homeostatic functioning is disturbed and the CNS reorganizes itself; in the short term, this reorganization can indicate a mechanism of functional response, but in the long term it can cause dysfunctional and pathological alterations (allostatic load), resulting in inappropriate responses^{96, 98-103}. Moreover, in advanced stages of the disease, these inappropriate responses can persist even during remission¹⁰⁴. In SUDS, the chronic use of PAS generates repeated overactivation of the dopaminergic pathway, leading to neuroadaptive mechanisms that may cause dysregulation in the brain reward system. This would

contribute to the transition from use to dependence^{105, 106}. Pre-clinical evidence sustain this hypothesis; ethanol, for example, generates an increase in BDNF during acute administration and decrease when administration is continuous¹⁰⁷.

We have also verified that peripheral BDNF levels are different among users of different drugs, possibly due to factors such as the substance mechanism of action, pattern and duration of drug use, clinical and psychiatric comorbidities of users and polysubstance use. Tobacco, for instance, which is associated with increased peripheral BDNF, was not controlled in 39% of the studies^{108, 109}. In crack/cocaine users, for example, polysubstance use is very common: 77% refer alcohol consumption and 85% tobacco in the last year¹¹⁰. Considering the use of different drugs, Hilburn et al. have found differences in serum BDNF levels among alcohol, cocaine and methamphetamine users in relation to craving and duration of withdrawn¹¹¹.

Meta-regressions showed that reduction in serum BDNF was associated with sex, age, years of drug use and duration of withdrawn. This corroborates with previous investigations that verified the impact of aging and sex on stored and circulating levels of BDNF in peripheral blood¹¹². In psychiatric disorders, like bipolar disorder and schizophrenia, meta-analysis have already pointed out that age and duration of the disease are associated with the reduction of BDNF^{92, 93}. Moreover, the reduction of serum BDNF during aging was associated with hippocampal shrinkage and memory declining¹¹³. It should be noted that this process may be different between the sexes. There are reports of higher BDNF levels in elderly women⁸⁶, which may result from hormonal differences evidenced between sexes. Estrogen, for instance, interferes in the expression of BDNF, as well as variations in menstrual cycle^{73, 112, 114}. It is noteworthy that the only study with an exclusively female sample

found increased BDNF levels in crack users during early withdrawal in relation to controls, and, furthermore, this study showed that more than half of the women had some additional psychiatric comorbidity ⁴⁹, which may indicate a greater drug use severity.

Evidence suggests that reduced BDNF may be related to the dependence severity ^{25, 29, 30, 57}. Even in addicted individuals to the same substance, there are differences in BDNF according to severity of drug use and prognosis ^{29, 32, 58, 115}. In alcohol users, for example, significantly lower levels of BDNF were evidenced in individuals with delirium tremens when compared to those who did not have this condition and controls, which was maintained even after detoxification ²⁵. In another study, which followed patients for 180 days after detoxification, serum BDNF of patients who maintained withdrawn was higher than baseline and higher than the group that relapsed during this period ⁵⁷. Similarly, Scherer et al. found an association between higher levels of BDNF and better clinical outcomes in crack cocaine users during detoxification ¹¹⁵. In this sense, the increase of BDNF levels, approaching the levels of the controls, during the treatment, may be associated to better clinical outcomes ^{116, 117}.

On the other hand, in young ecstasy dependents, with fewer years of use and more occasional use, BDNF was increased at baseline ²⁶, which may signal the fact that they were in the initial phase of dependence. Another study with controlled administration of THC verified increased serum BDNF in healthy controls, but not in subjects who previously used cannabis ¹¹⁸, signaling that the effect of drugs on BDNF is distinct among recreational use and in early and advanced phases of dependence. In bipolar patients, for example, BDNF serum levels diverging between the early and late stages were previously reported ¹¹⁹. Beyond bipolar disorder, the

reduction in BDNF levels has also been related to the severity of depression and schizophrenia^{90, 93, 120}.

The present study has some limitations. First, we have encountered large heterogeneity (80% or higher) for all analysis. In this light, we have explored, using subgroup and moderators analysis, some of the potential sources of this heterogeneity in which age, gender, time using the drug and the drug type and subtype explained some of the heterogeneity. Second, we have encountered publication bias for some analysis. For those, we performed the Duval and Tweedie trim and fill technique and recalculated the effect size adjusting for publication bias.

Altogether, these evidences suggest that BDNF could be related with severity and progression of drug dependence, as already described for other psychiatric and neurodegenerative disorders^{92, 117, 121-129}. Moreover, when considering BDNF levels interpretation in drug users, factors such as protocol of analysis, drug type and pattern of drug use need to be considered. As shown in the present study, it seems that different drug classes affect BDNF levels in different ways – however, the mechanisms of action of these drugs in BDNF concentration are not fully understood yet. Nonetheless, great progress is being made in order to achieve a better understanding of neuropsychological factors of drug dependence.

Acknowledgments

We acknowledge the support of National Counsel of Technological and Scientific Development (CNPQ), Coordination for the Improvement of Higher Education Personnel (CAPES), State Foundation for Research Support(FAPERGS), Federal University of Rio Grande do Sul, Hospital de Clínicas de Porto Alegre and National Secretary for Drug Policies(SENAD)

Financial Disclosure

- Coordination for the Improvement of Higher Education Personnel (CAPES)

References

1. Wikler A. Dynamics of drug dependence. Implications of a conditioning theory for research and treatment. *Arch Gen Psychiatry* 1973; **28**(5): 611-616.
2. American Psychiatric Association. (2013). *Diagnostic and statistical manual of mental disorders* (5th ed.). Arlington, VA: American Psychiatric Publishing.
3. Volkow ND, Wang GJ, Fowler JS, Tomasi D, Telang F. Addiction: beyond dopamine reward circuitry. *Proc Natl Acad Sci U S A* 2011; **108**(37): 15037-15042.
4. George O, Koob GF. Individual differences in prefrontal cortex function and the transition from drug use to drug dependence. *Neurosci Biobehav Rev* 2010; **35**(2): 232-247.
5. Seger D. Cocaine, metamfetamine, and MDMA abuse: the role and clinical importance of neuroadaptation. *Clin Toxicol (Phila)* 2010; **48**(7): 695-708.
6. Mendelson J, Baggott MJ, Flower K, Galloway G. Developing biomarkers for methamphetamine addiction. *Curr Neuropharmacol* 2011; **9**(1): 100-103.
7. Büttner A. Review: The neuropathology of drug abuse. *Neuropathol Appl Neurobiol* 2011; **37**(2): 118-134.
8. Sinha R. New findings on biological factors predicting addiction relapse vulnerability. *Curr Psychiatry Rep* 2011; **13**(5): 398-405.
9. Leshner AI. Addiction is a brain disease, and it matters. *Science* 1997; **278**(5335): 45-47.
10. Koob GF. Neurobiology of addiction. Toward the development of new therapies. *Ann N Y Acad Sci* 2000; **909**: 170-185.
11. Volkow ND, Fowler JS, Wang GJ, Telang F, Logan J, Jayne M et al. Cognitive control of drug craving inhibits brain reward regions in cocaine abusers. *Neuroimage* 2010; **49**(3): 2536-2543.
12. Goldstein RZ, Volkow ND. Drug addiction and its underlying neurobiological basis: neuroimaging evidence for the involvement of the frontal cortex. *Am J Psychiatry* 2002; **159**(10): 1642-1652.

13. Autry AE, Monteggia LM. Brain-derived neurotrophic factor and neuropsychiatric disorders. *Pharmacol Rev* 2012; **64**(2): 238-258.
14. Nagahara AH, Tuszyński MH. Potential therapeutic uses of BDNF in neurological and psychiatric disorders. *Nat Rev Drug Discov* 2011; **10**(3): 209-219.
15. Huang EJ, Reichardt LF. Neurotrophins: roles in neuronal development and function. *Annu Rev Neurosci* 2001; **24**: 677-736.
16. Binder DK, Scharfman HE. Brain-derived neurotrophic factor. *Growth Factors* 2004; **22**(3): 123-131.
17. Bekinschtein P, Cammarota M, Izquierdo I, Medina JH. BDNF and memory formation and storage. *Neuroscientist* 2008; **14**(2): 147-156.
18. Grimm JW, Lu L, Hayashi T, Hope BT, Su TP, Shaham Y. Time-dependent increases in brain-derived neurotrophic factor protein levels within the mesolimbic dopamine system after withdrawal from cocaine: implications for incubation of cocaine craving. *J Neurosci* 2003; **23**(3): 742-747.
19. Castrén E. Neurotrophins as mediators of drug effects on mood, addiction, and neuroprotection. *Mol Neurobiol* 2004; **29**(3): 289-302.
20. Schmidt HD, Duman RS. Peripheral BDNF produces antidepressant-like effects in cellular and behavioral models. *Neuropsychopharmacology* 2010; **35**(12): 2378-2391.
21. Miguel-Hidalgo JJ. The role of glial cells in drug abuse. *Curr Drug Abuse Rev* 2009; **2**(1): 72-82.
22. Volkow ND, Koob G, Baler R. Biomarkers in substance use disorders. *ACS Chem Neurosci* 2015; **6**(4): 522-525.
23. Karege F, Schwald M, Cisse M. Postnatal developmental profile of brain-derived neurotrophic factor in rat brain and platelets. *Neurosci Lett* 2002; **328**(3): 261-264.
24. Pan W, Banks WA, Fasold MB, Bluth J, Kastin AJ. Transport of brain-derived neurotrophic factor across the blood-brain barrier. *Neuropharmacology* 1998; **37**(12): 1553-1561.
25. Huang MC, Chen CH, Liu HC, Chen CC, Ho CC, Leu SJ. Differential patterns

- of serum brain-derived neurotrophic factor levels in alcoholic patients with and without delirium tremens during acute withdrawal. *Alcohol Clin Exp Res* 2011; **35**(1): 126-131.
26. Angelucci F, Ricci V, Martinotti G, Palladino I, Spalletta G, Caltagirone C et al. Ecstasy (MDMA)-addicted subjects show increased serum levels of brain-derived neurotrophic factor, independently from a rise of drug-induced psychotic symptoms. *Addict Biol* 2010; **15**(3): 365-367.
 27. Cavus SY, Dilbaz N, Darcin AE, Eren F, Kaya H, Kaya O. Alterations in serum BDNF levels in early alcohol withdrawal and comparison with healthy controls. *Bulletin of Clinical Psychopharmacology*, vol. 222012, pp 210-215.
 28. Ke X, Ding Y, Xu K, He H, Zhang M, Wang D et al. Serum brain-derived neurotrophic factor and nerve growth factor decreased in chronic ketamine abusers. *Drug Alcohol Depend* 2014; **142**: 290-294.
 29. D'Sa C, Fox HC, Hong AK, Dileone RJ, Sinha R. Increased serum brain-derived neurotrophic factor is predictive of cocaine relapse outcomes: a prospective study. *Biol Psychiatry* 2011; **70**(8): 706-711.
 30. Sordi AO, Pechansky F, Kessler FH, Kapczinski F, Pfaffenseller B, Gubert C et al. Oxidative stress and BDNF as possible markers for the severity of crack cocaine use in early withdrawal. *Psychopharmacology (Berl)* 2014; **231**(20): 4031-4039.
 31. Pedraz M, Martín-Velasco AI, García-Marchena N, Araos P, Serrano A, Romero-Sanchiz P et al. Plasma concentrations of BDNF and IGF-1 in abstinent cocaine users with high prevalence of substance use disorders: relationship to psychiatric comorbidity. *PLoS One* 2015; **10**(3): e0118610.
 32. Corominas-Roso M, Roncero C, Daigre C, Grau-Lopez L, Ros-Cucurull E, Rodríguez-Cintas L et al. Changes in brain-derived neurotrophic factor (BDNF) during withdrawn could be associated with relapse in cocaine-dependent patients. *Psychiatry Res* 2015; **225**(3): 309-314.
 33. Roncero C, Palma-Álvarez RF, Ros-Cucurull E, Barral C, Gonzalvo B, Corominas-Roso M et al. Cocaine-induced Psychosis and Brain-derived Neurotrophic Factor in Patients with Cocaine Dependence: Report of Two Cases. *Clin Psychopharmacol Neurosci* 2016; **14**(1): 109-113.
 34. Moher D, Liberati A, Tetzlaff J, Altman DG, Group P. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *BMJ* 2009; **339**: b2535.

35. Plot Digitizer. <http://plotdigitizer.sourceforge.net/>.
36. Wells G, Shea B, O'Connell D, Peterson J, Welch V, Losos M et al. The Newcastle Ottawa Scale (NOS) for assessing the quality if nonrandomized studies in meta-analyses. Available from: URL: http://www.ohri.ca/programs/clinical_epidemiology/oxford.htm.
37. Higgins J, Green S, (editors). Cochrane Handbook for Systematic Reviews of Interventions Version 5.1.0 [updated March 2011]. The Cochrane Collaboration, 2011. Available from <http://handbook.cochrane.org>.
38. D'Souza DC, Pittman B, Perry E, Simen A. Preliminary evidence of cannabinoid effects on brain-derived neurotrophic factor (BDNF) levels in humans. *Psychopharmacology (Berl)* 2009; **202**(4): 569-578.
39. Ren W, Tao J, Wei Y, Su H, Zhang J, Xie Y et al. Time-Dependent Serum Brain-Derived Neurotrophic Factor Decline During Methamphetamine Withdrawal. *Medicine (Baltimore)* 2016; **95**(5): e2604.
40. Joe KH, Kim YK, Kim TS, Roh SW, Choi SW, Kim YB et al. Decreased plasma brain-derived neurotrophic factor levels in patients with alcohol dependence. *Alcohol Clin Exp Res* 2007; **31**(11): 1833-1838.
41. D'Sa C, Dileone RJ, Anderson GM, Sinha R. Serum and plasma brain-derived neurotrophic factor (BDNF) in abstinent alcoholics and social drinkers. *Alcohol* 2012; **46**(3): 253-259.
42. Chen SL, Lee SY, Chang YH, Wang TY, Chen SH, Chu CH et al. The BDNF Val66Met polymorphism and plasma brain-derived neurotrophic factor levels in Han Chinese heroin-dependent patients. *Sci Rep* 2015; **5**: 8148.
43. Kim DJ, Roh S, Kim Y, Yoon SJ, Lee HK, Han CS et al. High concentrations of plasma brain-derived neurotrophic factor in methamphetamine users. *Neurosci Lett* 2005; **388**(2): 112-115.
44. Narvaez JC, Magalhães PV, Fries GR, Colpo GD, Czepielewski LS, Vianna P et al. Peripheral toxicity in crack cocaine use disorders. *Neurosci Lett* 2013; **544**: 80-84.
45. von Diemen L, Kapczinski F, Sordi AO, de Magalhães Narvaez JC, Guimarães LS, Kessler FH et al. Increase in brain-derived neurotrophic factor expression in early crack cocaine withdrawal. *Int J Neuropsychopharmacol* 2014; **17**(1):

- 33-40.
46. Heberlein A, Büscher P, Schuster R, Kleimann A, Lichtinghagen R, Rhein M et al. Do changes in the BDNF promoter methylation indicate the risk of alcohol relapse? *Eur Neuropsychopharmacol* 2015; **25**(11): 1892-1897.
 47. Heberlein A, Käser M, Lichtinghagen R, Rhein M, Lenz B, Kornhuber J et al. TNF- α and IL-6 serum levels: neurobiological markers of alcohol consumption in alcohol-dependent patients? *Alcohol* 2014; **48**(7): 671-676.
 48. Heberlein A, Muschler M, Wilhelm J, Frieling H, Lenz B, Gröschl M et al. BDNF and GDNF serum levels in alcohol-dependent patients during withdrawal. *Prog Neuropsychopharmacol Biol Psychiatry* 2010; **34**(6): 1060-1064.
 49. Viola TW, Tractenberg SG, Levandowski ML, Pezzi JC, Bauer ME, Teixeira AL et al. Neurotrophic factors in women with crack cocaine dependence during early withdrawal: the role of early life stress. *J Psychiatry Neurosci* 2014; **39**(3): 206-214.
 50. Zhang J, Zhang X, Su H, Tao J, Xie Y, Han B et al. Increased serum brain-derived neurotrophic factor levels during opiate withdrawal. *Neurosci Lett* 2014; **571**: 61-65.
 51. Zhang K, Jiang H, Zhang Q, Du J, Wang Y, Zhao M. Brain-derived neurotrophic factor serum levels in heroin-dependent patients after 26 weeks of withdrawal. *Compr Psychiatry* 2016; **65**: 150-155.
 52. Zanardini R, Fontana A, Pagano R, Mazzaro E, Bergamasco F, Romagnosi G et al. Alterations of brain-derived neurotrophic factor serum levels in patients with alcohol dependence. *Alcohol Clin Exp Res* 2011; **35**(8): 1529-1533.
 53. Viola TW, Tractenberg SG, Kluwe-Schiavon B, Levandowski ML, Sanvicente-Vieira B, Wearick-Silva LE et al. Brain-Derived Neurotrophic Factor and Delayed Verbal Recall in Crack/Cocaine Dependents. *Eur Addict Res* 2015; **21**(5): 273-278.
 54. Schuster R, Kleimann A, Rehme MK, Taschner L, Glahn A, Groh A et al. Elevated methylation and decreased serum concentrations of BDNF in patients in levomethadone compared to diamorphine maintenance treatment. *Eur Arch Psychiatry Clin Neurosci* 2017; **267**(1): 33-40.
 55. Lee BC, Chul BL, Choi IG, Kim YK, Ham BJ, Yang BH et al. Relation between plasma brain-derived neurotrophic factor and nerve growth factor in the male

- patients with alcohol dependence. *Alcohol* 2009; **43**(4): 265-269.
56. Köhler S, Klimke S, Hellweg R, Lang UE. Serum brain-derived neurotrophic factor and nerve growth factor concentrations change after alcohol withdrawal: preliminary data of a case-control comparison. *Eur Addict Res* 2013; **19**(2): 98-104.
 57. Costa MA, Girard M, Dalmay F, Malauzat D. Brain-derived neurotrophic factor serum levels in alcohol-dependent subjects 6 months after alcohol withdrawal. *Alcohol Clin Exp Res* 2011; **35**(11): 1966-1973.
 58. Corominas-Roso M, Roncero C, Jose Eiroa-Orosa F, Gonzalvo B, Grau-Lopez L, Ribases M et al. Brain-derived neurotrophic factor serum levels in cocaine-dependent patients during early withdrawn. *Eur Neuropsychopharmacol* 2012.
 59. Corominas-Roso M, Roncero C, Eiroa-Orosa FJ, Ribasés M, Barral C, Daigre C et al. Serum brain-derived neurotrophic factor levels and cocaine-induced transient psychotic symptoms. *Neuropsychobiology* 2013; **68**(3): 146-155.
 60. Angelucci F, Ricci V, Pomponi M, Conte G, Mathé AA, Attilio Tonali P et al. Chronic heroin and cocaine abuse is associated with decreased serum concentrations of the nerve growth factor and brain-derived neurotrophic factor. *J Psychopharmacol* 2007; **21**(8): 820-825.
 61. Angelucci F, Ricci V, Spalletta G, Pomponi M, Tonioni F, Caltagirone C et al. Reduced serum concentrations of nerve growth factor, but not brain-derived neurotrophic factor, in chronic cannabis abusers. *Eur Neuropsychopharmacol* 2008; **18**(12): 882-887.
 62. Han B, Zhang XY, Wang DY, Ren WW, Gu YY, Zhu L et al. Serum brain-derived neurotrophic factor levels and psychotic symptoms in heroin dependence. *Compr Psychiatry* 2015; **62**: 80-85.
 63. Heberlein A, Dürsteler-MacFarland KM, Lenz B, Frieling H, Grösch M, Bönsch D et al. Serum levels of BDNF are associated with craving in opiate-dependent patients. *J Psychopharmacol* 2011; **25**(11): 1480-1484.
 64. Meng D, Wu T, Rao U, North CS, Xiao H, Javors MA et al. Serum NPY and BDNF response to a behavioral stressor in alcohol-dependent and healthy control participants. *Psychopharmacology (Berl)* 2011; **218**(1): 59-67.
 65. Ricci V, Martinotti G, Gelfo F, Tonioni F, Caltagirone C, Bria P et al. Chronic ketamine use increases serum levels of brain-derived neurotrophic factor. *Psychopharmacology (Berl)* 2011; **215**(1): 143-148.

66. Geisel O, Hellweg R, Müller CA. Serum levels of brain-derived neurotrophic factor in alcohol-dependent patients receiving high-dose baclofen. *Psychiatry Res* 2016; **240**: 177-180.
67. Polyakova M, Stuke K, Schuemberg K, Mueller K, Schoenknecht P, Schroeter ML. BDNF as a biomarker for successful treatment of mood disorders: a systematic & quantitative meta-analysis. *J Affect Disord* 2015; **174**: 432-440.
68. Bocchio-Chiavetto L, Bagnardi V, Zanardini R, Molteni R, Nielsen MG, Placentino A et al. Serum and plasma BDNF levels in major depression: a replication study and meta-analyses. *World J Biol Psychiatry* 2010; **11**(6): 763-773.
69. Piccinni A, Marazziti D, Catena M, Domenici L, Del Debbio A, Bianchi C et al. Plasma and serum brain-derived neurotrophic factor (BDNF) in depressed patients during 1 year of antidepressant treatments. *J Affect Disord* 2008; **105**(1-3): 279-283.
70. Rasmussen P, Brassard P, Adser H, Pedersen MV, Leick L, Hart E et al. Evidence for a release of brain-derived neurotrophic factor from the brain during exercise. *Exp Physiol* 2009; **94**(10): 1062-1069.
71. Kerschensteiner M, Gallmeier E, Behrens L, Leal VV, Misgeld T, Klinkert WE et al. Activated human T cells, B cells, and monocytes produce brain-derived neurotrophic factor in vitro and in inflammatory brain lesions: a neuroprotective role of inflammation? *J Exp Med* 1999; **189**(5): 865-870.
72. Azoulay D, Vachapova V, Shihman B, Miler A, Karni A. Lower brain-derived neurotrophic factor in serum of relapsing remitting MS: reversal by glatiramer acetate. *J Neuroimmunol* 2005; **167**(1-2): 215-218.
73. Sohrabji F, Lewis DK. Estrogen-BDNF interactions: implications for neurodegenerative diseases. *Front Neuroendocrinol* 2006; **27**(4): 404-414.
74. Yamamoto H, Gurney ME. Human platelets contain brain-derived neurotrophic factor. *J Neurosci* 1990; **10**(11): 3469-3478.
75. Nakahashi T, Fujimura H, Altar CA, Li J, Kambayashi J, Tandon NN et al. Vascular endothelial cells synthesize and secrete brain-derived neurotrophic factor. *FEBS Lett* 2000; **470**(2): 113-117.
76. Donovan MJ, Miranda RC, Kraemer R, McCaffrey TA, Tessarollo L, Mahadeo

- D et al. Neurotrophin and neurotrophin receptors in vascular smooth muscle cells. Regulation of expression in response to injury. *Am J Pathol* 1995; **147**(2): 309-324.
77. Fujimura H, Altar CA, Chen R, Nakamura T, Nakahashi T, Kambayashi J et al. Brain-derived neurotrophic factor is stored in human platelets and released by agonist stimulation. *Thromb Haemost* 2002; **87**(4): 728-734.
78. Lommatsch M, Schloetcke K, Klotz J, Schuhbaeck K, Zingler D, Zingler C et al. Brain-derived neurotrophic factor in platelets and airflow limitation in asthma. *Am J Respir Crit Care Med* 2005; **171**(2): 115-120.
79. Radka SF, Holst PA, Fritzsche M, Altar CA. Presence of brain-derived neurotrophic factor in brain and human and rat but not mouse serum detected by a sensitive and specific immunoassay. *Brain Res* 1996; **709**(1): 122-301.
80. Kishino A, Katayama N, Ishige Y, Yamamoto Y, Ogo H, Tatsuno T et al. Analysis of effects and pharmacokinetics of subcutaneously administered BDNF. *Neuroreport* 2001; **12**(5): 1067-1072.
81. Poduslo JF, Curran GL. Permeability at the blood-brain and blood-nerve barriers of the neurotrophic factors: NGF, CNTF, NT-3, BDNF. *Brain Res Mol Brain Res* 1996; **36**(2): 280-286.
82. Trajkovska V, Marcussen AB, Vinberg M, Hartvig P, Aznar S, Knudsen GM. Measurements of brain-derived neurotrophic factor: methodological aspects and demographical data. *Brain Res Bull* 2007; **73**(1-3): 143-149.
83. Serra-Millàs M. Are the changes in the peripheral brain-derived neurotrophic factor levels due to platelet activation? *World J Psychiatry* 2016; **6**(1): 84-101.
84. Yi J, Liu Z, Craft D, O'Mullan P, Ju G, Gelfand CA. Intrinsic peptidase activity causes a sequential multi-step reaction (SMSR) in digestion of human plasma peptides. *J Proteome Res* 2008; **7**(12): 5112-5118.
85. Zuccato C, Marullo M, Vitali B, Tarditi A, Mariotti C, Valenza M et al. Brain-derived neurotrophic factor in patients with Huntington's disease. *PLoS One* 2011; **6**(8): e22966.
86. Bus BA, Molendijk ML, Penninx BJ, Buitelaar JK, Kenis G, Prickaerts J et al. Determinants of serum brain-derived neurotrophic factor. *Psychoneuroendocrinology* 2011; **36**(2): 228-239.

87. Lommatzsch M, Virchow JC. Letter regarding article by Ejiri et al, "possible role of brain-derived neurotrophic factor in the pathogenesis of coronary artery disease". *Circulation* 2006; **113**(16): e724; author reply e724-725.
88. Begliuomini S, Casarosa E, Pluchino N, Lenzi E, Centofanti M, Freschi L et al. Influence of endogenous and exogenous sex hormones on plasma brain-derived neurotrophic factor. *Hum Reprod* 2007; **22**(4): 995-1002.
89. Munkholm K, Vinberg M, Kessing LV. Peripheral blood brain-derived neurotrophic factor in bipolar disorder: a comprehensive systematic review and meta-analysis. *Mol Psychiatry* 2016; **21**(2): 216-228.
90. Fernandes BS, Molendijk ML, Köhler CA, Soares JC, Leite CM, Machado-Vieira R et al. Peripheral brain-derived neurotrophic factor (BDNF) as a biomarker in bipolar disorder: a meta-analysis of 52 studies. *BMC Med* 2015; **13**: 289.
91. Fernandes BS, Berk M, Turck CW, Steiner J, Gonçalves CA. Decreased peripheral brain-derived neurotrophic factor levels are a biomarker of disease activity in major psychiatric disorders: a comparative meta-analysis. *Mol Psychiatry* 2014; **19**(7): 750-751.
92. Fernandes BS, Gama CS, Ceresér KM, Yatham LN, Fries GR, Colpo G et al. Brain-derived neurotrophic factor as a state-marker of mood episodes in bipolar disorders: a systematic review and meta-regression analysis. *J Psychiatr Res* 2011; **45**(8): 995-1004.
93. Fernandes BS, Steiner J, Berk M, Molendijk ML, Gonzalez-Pinto A, Turck CW et al. Peripheral brain-derived neurotrophic factor in schizophrenia and the role of antipsychotics: meta-analysis and implications. *Mol Psychiatry* 2015; **20**(9): 1108-1119.
94. Gama CS, Kunz M, Magalhães PV, Kapczinski F. Staging and neuroprogression in bipolar disorder: a systematic review of the literature. *Rev Bras Psiquiatr* 2013; **35**(1): 70-74.
95. Grande I, Magalhães PV, Kunz M, Vieta E, Kapczinski F. Mediators of allostatic load and systemic toxicity in bipolar disorder. *Physiol Behav* 2012; **106**(1): 46-50.
96. Kapczinski F, Vieta E, Andreazza AC, Frey BN, Gomes FA, Tramontina J et al. Allostatic load in bipolar disorder: implications for pathophysiology and treatment. *Neurosci Biobehav Rev* 2008; **32**(4): 675-692.

97. Kapczinski F, Magalhães PV, Balanzá-Martinez V, Dias VV, Frangou S, Gama CS et al. Staging systems in bipolar disorder: an International Society for Bipolar Disorders Task Force Report. *Acta Psychiatr Scand* 2014; **130**(5): 354-363.
98. McEwen BS. Protective and damaging effects of stress mediators. *N Engl J Med* 1998; **338**(3): 171-179.
99. McEwen BS. Stress, adaptation, and disease. Allostasis and allostatic load. *Ann N Y Acad Sci* 1998; **840**: 33-44.
100. McEwen BS. Allostasis and allostatic load: implications for neuropsychopharmacology. *Neuropsychopharmacology* 2000; **22**(2): 108-124.
101. McEwen BS. Sex, stress and the hippocampus: allostasis, allostatic load and the aging process. *Neurobiol Aging* 2002; **23**(5): 921-939.
102. McEwen BS. Mood disorders and allostatic load. *Biol Psychiatry* 2003; **54**(3): 200-207.
103. McEwen BS, Wingfield JC. The concept of allostasis in biology and biomedicine. *Horm Behav* 2003; **43**(1): 2-15.
104. Berk M, Kapczinski F, Andreazza AC, Dean OM, Giorlando F, Maes M et al. Pathways underlying neuroprogression in bipolar disorder: focus on inflammation, oxidative stress and neurotrophic factors. *Neurosci Biobehav Rev* 2011; **35**(3): 804-817.
105. Koob GF, Le Moal M. Drug addiction, dysregulation of reward, and allostasis. *Neuropsychopharmacology* 2001; **24**(2): 97-129.
106. George O, Le Moal M, Koob GF. Allostasis and addiction: role of the dopamine and corticotropin-releasing factor systems. *Physiol Behav* 2012; **106**(1): 58-64.
107. McGough NN, He DY, Logrip ML, Jeanblanc J, Phamluong K, Luong K et al. RACK1 and brain-derived neurotrophic factor: a homeostatic pathway that regulates alcohol addiction. *J Neurosci* 2004; **24**(46): 10542-10552.
108. Hashimoto K, Iwata Y, Nakamura K, Tsujii M, Tsuchiya KJ, Sekine Y et al. Reduced serum levels of brain-derived neurotrophic factor in adult male

- patients with autism. *Prog Neuropsychopharmacol Biol Psychiatry* 2006; **30**(8): 1529-1531.
109. Jamal M, Van der Does W, Elzinga BM, Molendijk ML, Penninx BW. Association between smoking, nicotine dependence, and BDNF Val66Met polymorphism with BDNF concentrations in serum. *Nicotine Tob Res* 2015; **17**(3): 323-329.
110. Bastos FI, Bertoni N. Pesquisa Nacional sobre o uso de crack: quem são os usuários de crack e/ou similares do Brasil? Quantos são nas capitais brasileiras? Editora Fiocruz: Rio de Janeiro, 2014 p224.
111. Hilburn C, Nejtek VA, Underwood WA, Singh M, Patel G, Gangwani P *et al.* Is serum brain-derived neurotrophic factor related to craving for or use of alcohol, cocaine, or methamphetamine? *Neuropsychiatr Dis Treat* 2011; **7**: 357-364.
112. Lommatsch M, Zingler D, Schuhbaeck K, Schloetcke K, Zingler C, Schuff-Werner P *et al.* The impact of age, weight and gender on BDNF levels in human platelets and plasma. *Neurobiol Aging* 2005; **26**(1): 115-123.
113. Erickson KI, Prakash RS, Voss MW, Chaddock L, Heo S, McLaren M *et al.* Brain-derived neurotrophic factor is associated with age-related decline in hippocampal volume. *J Neurosci* 2010; **30**(15): 5368-5375.
114. Sohrabji F, Miranda RC, Toran-Allerand CD. Identification of a putative estrogen response element in the gene encoding brain-derived neurotrophic factor. *Proc Natl Acad Sci U S A* 1995; **92**(24): 11110-11114.
115. Scherer JN, Schuch S, Ornell F, Sordi AO, Kessler FHP, von *et al.* Higher levels of BDNF are associated with inpatient treatment adherence of crack-cocaine users. *Drug and Alcohol Dependence* 2015.
116. Duman RS, Monteggia LM. A neurotrophic model for stress-related mood disorders. *Biol Psychiatry* 2006; **59**(12): 1116-1127.
117. Zuccato C, Cattaneo E. Brain-derived neurotrophic factor in neurodegenerative diseases. *Nat Rev Neurol* 2009; **5**(6): 311-322.
118. Angelucci F, Gruber SH, El Khoury A, Tonali PA, Mathé AA. Chronic amphetamine treatment reduces NGF and BDNF in the rat brain. *Eur Neuropsychopharmacol* 2007; **17**(12): 756-762.

119. Kauer-Sant'Anna M, Kapczinski F, Andreazza AC, Bond DJ, Lam RW, Young LT *et al.* Brain-derived neurotrophic factor and inflammatory markers in patients with early- vs. late-stage bipolar disorder. *Int J Neuropsychopharmacol* 2009; **12**(4): 447-458.
120. Bus BA, Molendijk ML, Tendolkar I, Penninx BW, Prickaerts J, Elzinga BM *et al.* Chronic depression is associated with a pronounced decrease in serum brain-derived neurotrophic factor over time. *Mol Psychiatry* 2015; **20**(5): 602-608.
121. Krabbe KS, Nielsen AR, Krogh-Madsen R, Plomgaard P, Rasmussen P, Erikstrup C *et al.* Brain-derived neurotrophic factor (BDNF) and type 2 diabetes. *Diabetologia* 2007; **50**(2): 431-438.
122. Connor B, Young D, Yan Q, Faull RL, Synek B, Dragunow M. Brain-derived neurotrophic factor is reduced in Alzheimer's disease. *Brain Res Mol Brain Res* 1997; **49**(1-2): 71-81.
123. Murer MG, Yan Q, Raisman-Vozari R. Brain-derived neurotrophic factor in the control human brain, and in Alzheimer's disease and Parkinson's disease. *Prog Neurobiol* 2001; **63**(1): 71-124.
124. Qin XY, Cao C, Cawley NX, Liu TT, Yuan J, Loh YP *et al.* Decreased peripheral brain-derived neurotrophic factor levels in Alzheimer's disease: a meta-analysis study (N=7277). *Mol Psychiatry* 2017; **22**(2): 312-320.
125. Ventriglia M, Zanardini R, Bonomini C, Zanetti O, Volpe D, Pasqualetti P *et al.* Serum brain-derived neurotrophic factor levels in different neurological diseases. *Biomed Res Int* 2013; **2013**: 901082.
126. Angelucci F, Spalletta G, di Iulio F, Ciaramella A, Salani F, Colontoni L *et al.* Alzheimer's disease (AD) and Mild Cognitive Impairment (MCI) patients are characterized by increased BDNF serum levels. *Curr Alzheimer Res* 2010; **7**(1): 15-20.
127. Ciammola A, Sassone J, Cannella M, Calza S, Poletti B, Frati L *et al.* Low brain-derived neurotrophic factor (BDNF) levels in serum of Huntington's disease patients. *Am J Med Genet B Neuropsychiatr Genet* 2007; **144B**(4): 574-577.
128. Lee JG, Shin BS, You YS, Kim JE, Yoon SW, Jeon DW *et al.* Decreased serum brain-derived neurotrophic factor levels in elderly korean with dementia. *Psychiatry Investig* 2009; **6**(4): 299-305.

129. Yasutake C, Kuroda K, Yanagawa T, Okamura T, Yoneda H. Serum BDNF, TNF-alpha and IL-1beta levels in dementia patients: comparison between Alzheimer's disease and vascular dementia. *Eur Arch Psychiatry Clin Neurosci* 2006; **256**(7): 402-406.

Tables and figures legends:

Table 1. Characteristics of the studies included in the meta-analysis of serum or plasma BDNF levels in Substance Use Disorder

Table 2. Brain-derived neurotrophic factor (BDNF) levels from serum samples.

Table 3. Brain-derived neurotrophic factor (BDNF) levels from plasma samples.

Table 4. Meta-regression of potential moderators brain-derived neurotrophic factor (BDNF) in substance use disorders

Figure 1. Flow diagram of the meta-analysis of brain-derived neurotrophic factor (BDNF) in substance use disorders

Figure 2. Overall, plasma and serum Brain-derived neurotrophic factor (BDNF) levels in substance use disorders

Figure 3. Brain-derived neurotrophic factor (BDNF) levels from serum in alcohol users

Figure 4. Brain-derived neurotrophic factor (BDNF) levels from serum in opiate users

Figure 5. Brain-derived neurotrophic factor (BDNF) levels from serum in crack/cocaine users

Figure 6. Brain-derived neurotrophic factor (BDNF) levels from serum in

methamphetamine users

Supplemental information.

S1. Quality assessment of the eligible studies using the Newcastle-Ottawa Scale

S2. Quality assessment of the eligible studies using the Cochrane Collaboration

Table 1. Characteristics of the studies included in the meta-analysis of serum or plasma BDNF levels in Substance Use Disorder.

Study	Year	Drug	Withdrawal	Biological Matrix	Mean BDNF (casoNenhuma entrada de índice de ilustrações foi encontrada.es)	Mean BDNF (controls)	Unit	Cases (n)	Controls (n)	Gender
Cavus, S.Y.	2012	Alcohol	No Yes	Serum	Baseline: 52.9 ± 19 7º Day: 3.5 ± 8.8	65.3 ± 15.9	ng/mL	31	29	Men
Costa, M.A.	2011	Alcohol	No Yes No	Serum	Baseline: 22.98 ± 7.95 6 months (withdrawal): 31.9 ± 10.1 6 months (no withdrawal): 27.5 ± 10.4	24.2 ± 5.1	pg/mL	101	39	Mixed
D'Sa, C. (1)	2012	Alcohol	Yes Yes	Serum / Plasma	Plasma: 1.27 ± 1.12 Serum: 35.3 ± 11.1	Plasma 1.52 ± 0.68 Serum: 26.9 ± 9.5	ng/mL	16	16	Mixed
Zanardini, R.	2011	Alcohol	No No	Serum / Plasma	Plasma: 4.77 ± 3.50 Serum: 35.97 ± 10.49	Plasma 4.08 ± 4.00 Serum: 41.70 ± 11.49	ng/mL	37	37	Mixed
Geisel, O.	2016	Alcohol	Yes Yes Yes	Serum	Placebo drug group (baseline): 3446.2 ± 1285.8 Placebo drug group (T1): 3511.9 ± 1493.5 Placebo drug group (T2): 3796.7 ± 2047.4	3716.8 ± 1403.7	pg/mL	28	10	Mixed
Heberlein, A.	2015	Alcohol	No Yes Yes	Serum	Day 1: 647.67 ± 510.13 Day 7: 554.74 ± 398.25 Day 14: 653.81 ± 518.95	623.29 ± 303.03	pg/mL	99	33	Men
Huang, M-C.	2011	Alcohol	No No Yes Yes	Serum	Baseline (Without Delirium Tremens): 12.3 ± 3.3 Baseline (With Delirium Tremens): 6.2 ± 2.6 After one week (Without Delirium Tremens): 13.4 ± 3.5 After one week (With Delirium Tremens): 8.9 ± 4.4	14.8 ± 4.7	ng/mL	65	39	Mixed
Joe, K-H.	2007	Alcohol	Yes	Plasma	389.5 ± 501.7	822.5 ± 420.7	pg/mL	64	75	Men

Lee, B.C.	2009	Alcohol	No	Plasma	3502.2 ± 1726.9 a	861.75 ± 478.9	pg/mL	41	41	Men
Meng, D.	2011	Alcohol	Yes	Serum	Baseline: 12.69 ± 4.79	Baseline: 11.76 ± 5.05	ng/mL	14	10	Men
D'Souza, D.C.	2009	Cannabis	No	Serum	2552 ± 4067	2552 ± 4932	pg/mL	9	14	Mixed
Angelucci, F. (4)	2008	Cannabis	-	Serum	5984.23 ± 335.9	5683.62 ± 237.65	pg/mL	26	20	Mixed
Ke, X.	2014	Cetamine	-	Serum	9.5 ± 6.68	14.37 ± 6.07	ng/mL	93	39	Mixed
Ricci, V.	2011	Cetamine	No	Serum	12.7 ± 5.48	6159 ± 3.27	ng/mL	17	11	Mixed
Angelucci, F. (2)	2007	Cocaine	-	Serum	5182.46 ± 662.43	5433.32 ± 392.41	pg/mL	15	15	Mixed
Corominas-Roso, M.	2013	Cocaine	No	Serum	Baseline: 51.676 ± 17.505	76044 ± 32.661	ng/mL	23	46	Mixed
			Yes		After 12 days: 60.643 ± 22.607					
D'Sa, C. (2)	2011	Cocaine	Yes	Serum	36 ± 9.2	25.8 ± 8.9	ng/mL	35	34	Mixed
Pedraz, M.	2015	Cocaine	Yes	Plasma	274.9 ± 200.3	269.4 ± 242.7	pg/mL	89	85	Mixed
Narvaez, J.C.M.	2013	Crack	No described	Serum	14.21 ± 3.24	12.67 ± 3.74	ng/mL	53	50	Mixed
von Diemen, L.	2014	Crack	Yes	Serum	Baseline: 28.6 ± 11.0	39.5 ± 10.6	ng/mL	49	97	Men
			No		Discharge: 35.5 ± 12.3					
Viola, T.W.	2014	Crack	No	Plasma	4 days: 13726.88 ± 5908.50	8344.90 ± 2662.48	pg/mL	104	20	Women
			Yes		11 days: 13101.84 ± 6507.44					
			Yes		18 days: 14137.92 ± 7613.95					
Chen, S.L.	2015	Opiate (Heroin)	No	Plasma	10.1 ± 7.7	18.6 ± 9.4	ng/mL	172	102	Mixed
Heberlein, A. (1)	2011	Opiate (Heroin)	No	Serum	865.14 ± 358.8	556.62 ± 174.04	pg/mL	27	21	Men
Angelucci, F. (3)	2007	Opiate (Heroin)	No described	Serum	5091.59 ± 422.91	5433.32 ± 392.41	pg/mL	15	15	Mixed
Zhang, K.	2016	Opiate (Heroin)	No	Serum	Baseline: 987.25 ± 915.26	Baseline 3989.07 ± 2018.87 After 26 weeks 4003.05 ± 2011.83	pg/mL	53	52	Men
			Yes		After 26 weeks 2491.54 ± 1397.32					
Schuster, R.	2016	Opiate	No	Serum	48.25 ± 12.95	71.25 ± 17.43	ng/mL	30	51	Mixed

Zhang, J.	2014	Opiate (Heroin)	No No Yes	Serum	Baseline (n=72): 1680±577.37 Baseline (n=37): 1565±511.4 After one month (n=37): 1454± 555.7	1241 ± 335.52	pg/mL	72	90	Mixed
Angelucci, F. (1)	2010	MDMA (ecstasy)	- -	Serum	Non-psychotic: 11.28±0.39 Psychotic:11.36 ± 0.51	8.53±572	ng/mL	23	19	Mixed
Kim, D.J.	2005	Methamphetamine	Yes	Plasma	2536.25±2310.49	1352.61±1188.15	pg/mL	50	50	Men
Chen, P.H.	2014	Methamphetamine	No Yes Yes	Serum	Baseline: 9.84± 4.85 Acute phase: 9.32± 4.33 Subacute phase: 10.45± 5.41	16.26±4.72	ng/mL	59	59	Mixed
Ren, W.	2016	Methamphetamine	No No No	Serum	Baseline (n=179): 1460.28±490.69 Baseline (n=40): 1621.41±591.07 After one month (n=40): 1363.70 ± 580.59	1241.27± 335.52	pg/mL	179	90	Mixed

Table 2. Brain-derived neurotrophic factor (BDNF) levels from serum samples.

Analysis	N subgroups	N participants	SMD 95%CI Lower and Upper limit	P value	Trim and fill adjusted ES	I ²
Serum	38		-0.213 -0.51 0.08	0.16	0.16 (95%CI -0.16 to 0.49) [8]	93.46
User status						
Withdrawn	15		0.326 -0.64 0.21	0.35	-0.31 (95%CI -0.75 to 0.13) [1]	91.64
Active use	16		-0.456 -0.93 0.02	0.06	-0.51 (95%CI -1.12 to -0.08) [1]	94.68
Drug subtype						
Alcohol	14		-0.451 -0.89 -0.01	0.04	-0.73 (95%CI -1.22 to -0.254) [3]	90.54
Cannabis	2		0.346 -1.01 1.70	0.61	N/A	85.27
Ketamine	2		0.281 -1.80 2.36	0.79	N/A	95.08
Crack/cocaine	7		-0.233 -0.79 0.37	0.41	-0.09 (95%CI -0.65 to 0.46) [1]	90.65
Heroin	8		-0.416 -1.24 0.41	0.32	Unchanged	95.88
MDMA/Ecstasy	1		5.723 4.35 7.09	>0.001	N/A	0
Methamphetamine	4		-0.423 -1.37 0.52	0.38	-0.11 (95%CI -0.98 to 0.75) [1]	96.24
Serum and withdrawn						
Alcohol	8		-0.287 -1.01 0.43	0.43	-0.47 (95%CI -1.22 to 0.28) [1]	92.58
Crack/cocaine	3		0.076 -0.87 1.02	0.87	Unchanged	92.56
Heroin	2		-0.171 -1.52 1.17	0.80	N/A	95.69
Methamphetamine	2		-0.435 -1.86 0.99	0.55	N/A	95.40
Serum and active use						
Drug subtype						
Alcohol	6		-0.649 -1.16 -0.13	0.01	-0.97 (95%CI -1.57 to -0.32) [2]	86.84
Crack/cocaine	2		-0.962 -1.26 -0.66	>0.001	N/A	0
Heroin	4		-0.340 -1.91 1.23	0.67	Unchanged	97.87
Methamphetamine	2		-0.418 -2.21 1.37	0.64	N/A	98.22

Table 3. Brain-derived neurotrophic factor (BDNF) levels from plasma samples.

Analysis	N subgroups	N participants	SMD 95%CI Lower and Upper limit	P value	Trim and fill adjusted ES	I ²
Plasma	9		0.269 -0.36 0.90	0.40	Unchanged	95.55
User status						
Withdrawn	5		0.057 -0.58 0.70	0.86	Unchanged	91.78
Active use	4		0.545 -0.83 1.92	0.44	Unchanged	97.70
Drug subtype						
Alcohol	4		0.263 -1.04 1.57	0.69	0.67 (95%CI -0.74 to 2.09) [1]	96.52
Crack/cocaine	3		0.583 -0.06 1.23	0.07	Unchanged	85.83
Heroin	1		-1.011 -1.27 -0.75	>0.001	N/A	0
Methamphetamine	1		0.644 0.24 1.04	0.002	N/A	0
Plasma and withdrawn						
Drug subtype						
Alcohol	2		-0.668 -1.32 -0.00	0.04	N/A	66.34
Crack/cocaine	2		0.398 -0.38 1.17	0.31	N/A	86.46
Methamphetamine	1		0.644 0.24 1.04	0.002	N/A	0
Plasma and active use						
Drug subtype						
Alcohol	2		1.128 -0.73 2.99	0.23	N/A	96.41
Crack/cocaine	1		0.977 0.48 1.47	>0.001	N/A	0
Heroin	1		-1.015 -1.27 -0.75	>0.001	N/A	0

Table 4. Meta-regression of potential moderators brain-derived neurotrophic factor (BDNF) in substance use disorders

Moderator	N subgroups	β	95%CI	P value	R ²
User status					
Active use					
% of males	20	-0.013	-0.03	0.00	0.19
Mean age	20	-0.004	-0.06	0.05	0.87
Years of use	9	-0.021	-0.13	0.08	0.63
Age of the first use	9	0.134	-0.17	0.44	0.38
Length of abstinence	N/A				
Withdraw					
% of males	20	-0.153	-0.02	-0.00	0.01
Mean age	20	-0.033	-0.09	0.02	0.28
Years of use	13	-0.025	-0.09	0.04	0.47
Age of the first use	10	0.058	-0.12	0.23	0.52
Length of abstinence	10	-0.441	-0.64	0.75	0.46
Sample					
Serum					
% of males	38	-0.029	-0.04	-0.01	0.0008
Mean age	38	-0.049	-0.91	-0.00	0.02
Years of use	20	-0.064	-0.12	-0.00	0.04
Age of the first use	19	0.013	-0.18	0.21	0.88
Length of abstinence	15	-0.864	-1.71	-0.02	0.04
Plasma					
% of males	9	-0.005	-0.02	0.01	0.56
Mean age	9	-0.001	-0.09	0.07	0.81
Years of use	8	0.041	-0.09	0.18	0.55
Age of the first use	6	0.021	-0.23	0.27	0.86
Length of abstinence	4	0.916	-0.81	2.65	0.30

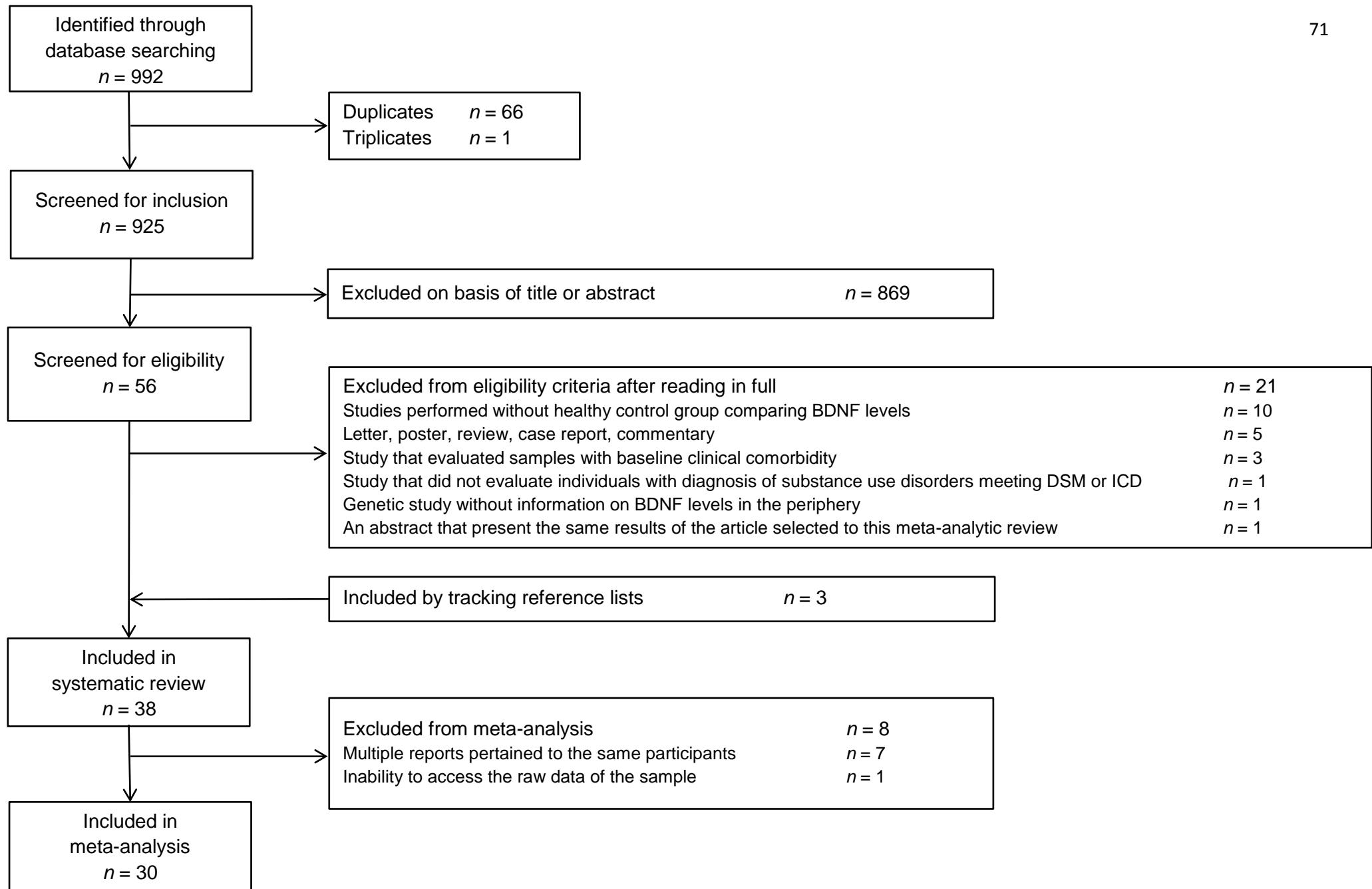


Figure 1. Flow diagram of the meta-analysis of brain-derived neurotrophic factor (BDNF) in substance use disorders.

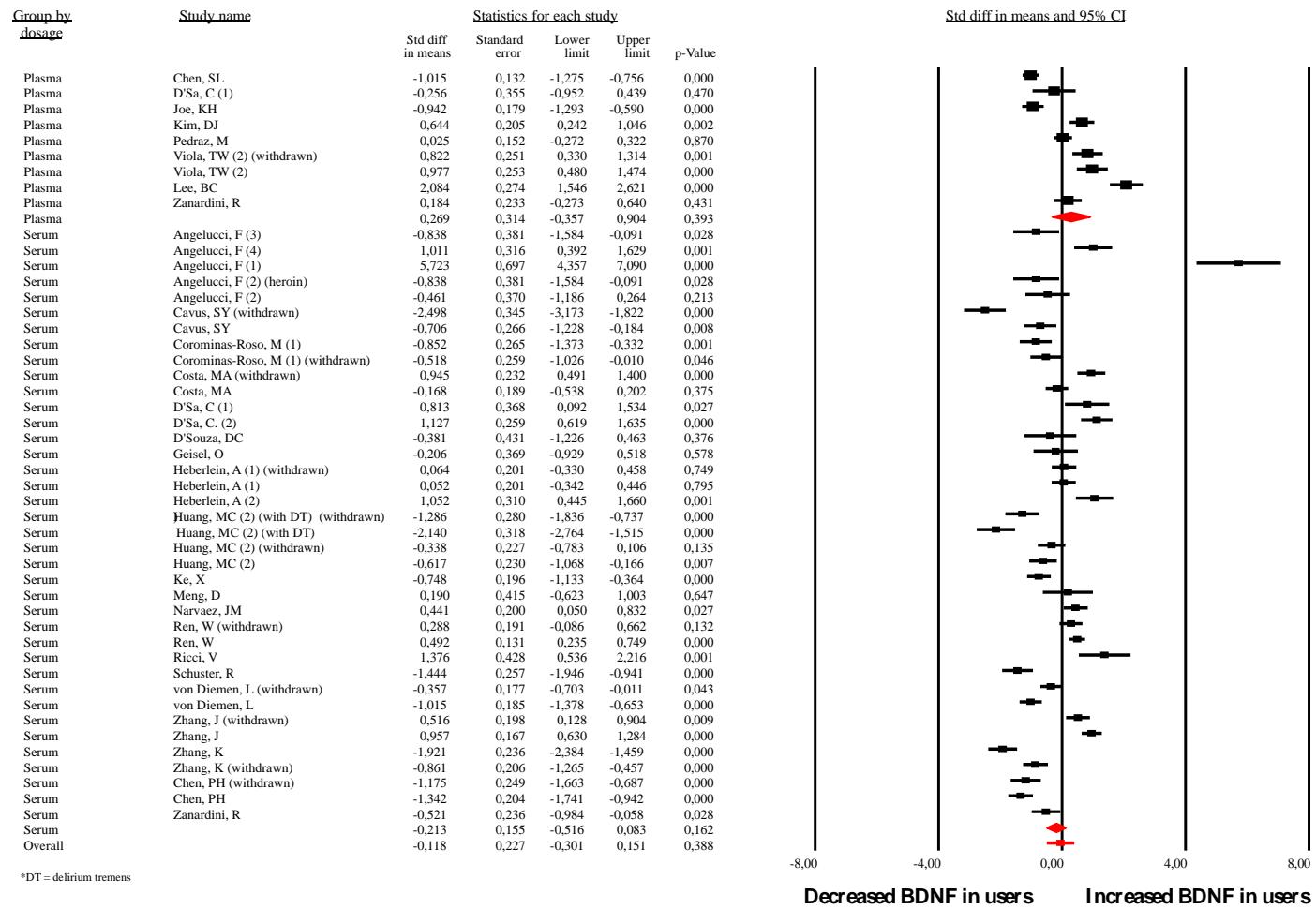
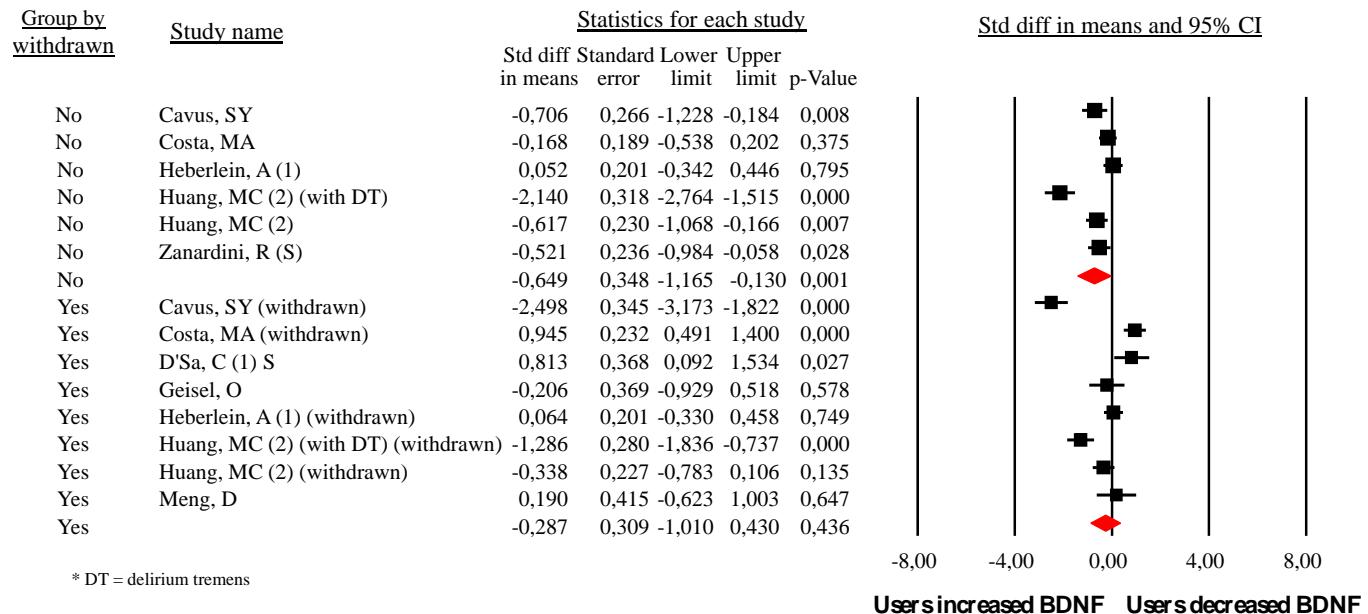


Figure 2. Overall, plasma and serum Brain-derived neurotrophic factor (BDNF) levels in substance use disorders.

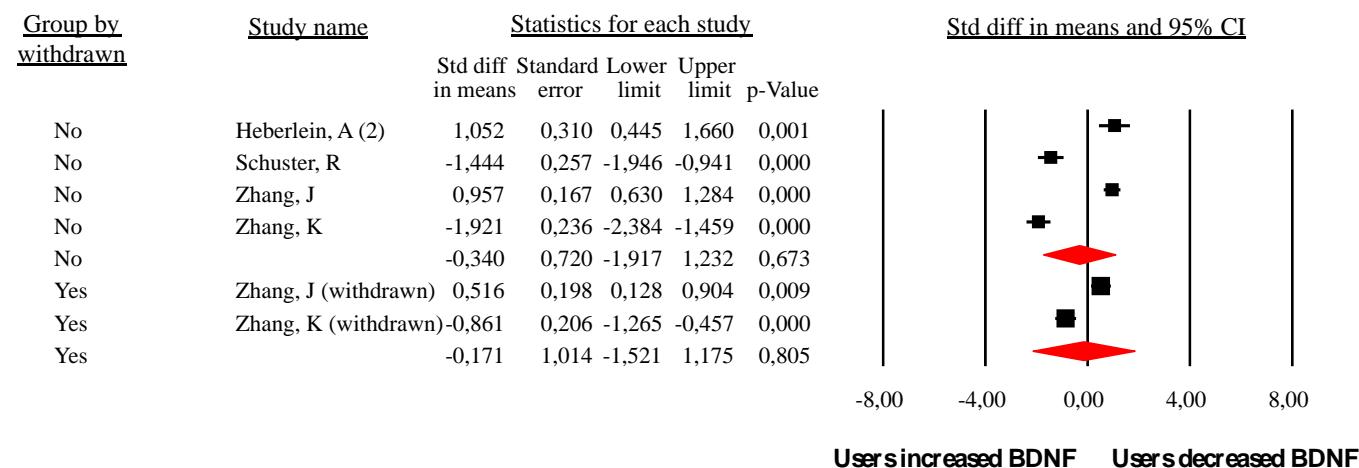


D'Sa, C (1) - Serum and plasma brain-derived neurotrophic factor (BDNF) in abstinent alcoholics and social drinkers

Heberlein, A (1) - Do changes in the BDNF promoter methylation indicate the risk of alcohol relapse?

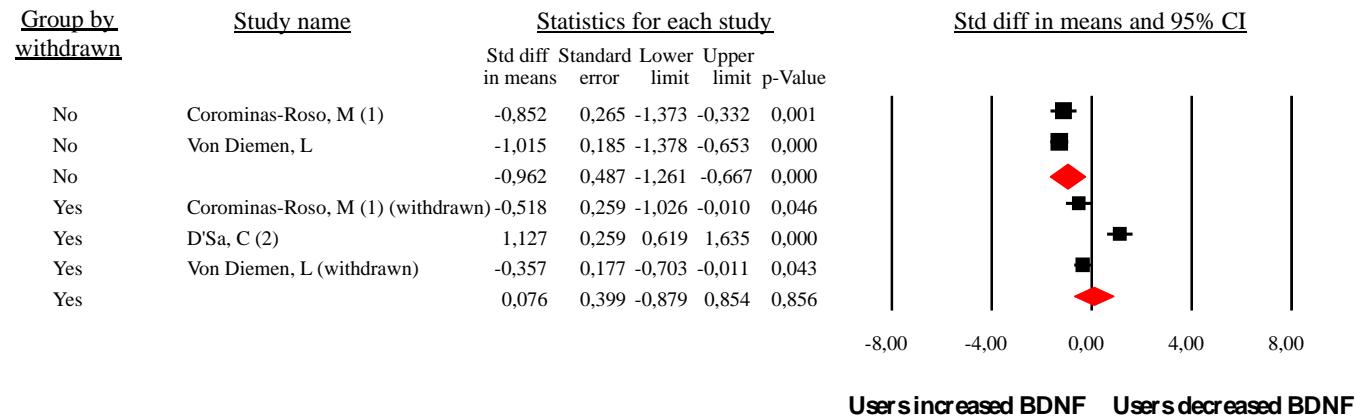
Huang, MC (2) - Differential Patterns of Serum Brain-Derived Neurotrophic Factor Levels in Alcoholic Patients With and Without Delirium Tremens During Acute Withdrawal

Figure 3. Brain-derived neurotrophic factor (BDNF) levels from serum in alcohol users



Heberlein, A (2): Serum levels of BDNF are associated with craving in opiate-dependent patients

Figure 4. Brain-derived neurotrophic factor (BDNF) levels from serum in opiate users



Corominas-Roso, M (1) - Brain-derived neurotrophic factor serum levels in cocaine-dependent patients during early abstinence

D'Sa, C (1) - Serum and plasma brain-derived neurotrophic factor (BDNF) in abstinent alcoholics and social drinkers

Figure 5. Brain-derived neurotrophic factor (BDNF) levels from serum in crack/cocaine users

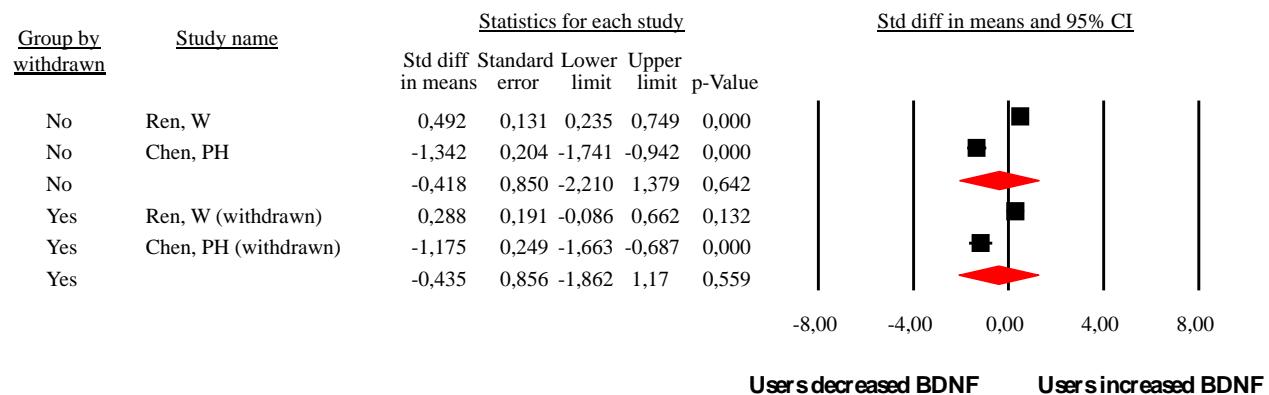


Figure 6. Brain-derived neurotrophic factor (BDNF) levels from serum in methamphetamine users.

Supplemental information 1. Quality assessment of the eligible studies using the Newcastle-Ottawa Scale.

	Selection (max. 4 stars)				Comparability (max. 2 stars)		Exposure (max. 2 stars)		Total stars (max. 8 stars)
	Adequacy of case definition	Representativeness of the cases	Selection of controls	Definition of controls	Control for possible confounding factor (major and any additional ones)	Ascertainment of exposure	Equivalence in ascertainment for cases and controls	Equivalent non-response rate between groups*	
Angelucci, F (1)	★			★	★	★	★	do not apply	5
Angelucci, F (2,3)	★		★	★	★	★	★	do not apply	6
Angelucci, F (4)	★		★	★	★	★	★	do not apply	6
Cavus, SY	★	★	★	★	★	★	★	do not apply	7
Chen, PH	★		★		★	★	★	do not apply	5
Chen, SL	★		★	★	★	★	★	do not apply	7
Corominas-Roso, M (1)	★		★	★	★	★	★	do not apply	6
Corominas-Roso, M (2)	★		★	★	★	★	★	do not apply	6
Costa, MA	★		★	★	★	★	★	do not apply	6
D'Sá, C (1)	★	★	★	★	★	★	★	do not apply	8
D'Sá, C (2)	★	★	★	★	★	★	★	do not apply	8
Han, B	★	★			★	★	★	do not apply	5
Heberlein, A (1)	★		★	★	★	★	★	do not apply	7
Heberlein, A (2)	★			★	★	★	★	do not apply	5
Heberlein, A (3)	★		★	★	★	★	★	do not apply	7
Heberlein, A (4)	★		★	★	★	★	★	do not apply	7
Huang, MC	★		★	★	★	★	★	do not apply	6
Huang, MC	★	★	★	★	★	★	★	do not apply	8
Joe, KH	★		★	★	★	★	★	do not apply	5
Ke, X	★		★	★	★	★	★	do not apply	7
Kim, DJ	★	★	★	★	★	★	★	do not apply	7
Kohler, S	★			★	★	★	★	do not apply	6
Lee, BC	★		★	★	★	★	★	do not apply	6
Meng, D	★			★	★	★	★	do not apply	5
Narvaez, JM	★		★	★	★	★	★	do not apply	7
Pedraza, M	★	★	★	★	★	★	★	do not apply	8
Ren, W	★		★	★	★	★	★	do not apply	7
Ricci, V	★			★	★	★	★	do not apply	5
Schuster, R	★		★	★	★	★	★	do not apply	6
Sordi, AO	★		★	★	★	★	★	do not apply	7
Viola, TW (1)	★		★	★	★	★	★	do not apply	6
Viola, TW (2)	★		★	★	★	★	★	do not apply	7
von Diemen, L	★		★	★	★	★	★	do not apply	7
Zanardini, R	★			★	★	★	★	do not apply	6
Zhang, J	★		★	★	★	★	★	do not apply	6
Zhang, K	★		★	★	★	★	★	do not apply	6

Supplemental information 2. Quality assessment of the eligible studies using the Cochrane Collaboration

	Selection Bias	Performance Bias	Detection Bias	Attrition Bias	Reporting Bias	Other Bias
	Random sequence generation	Allocation Concealment	Blinding of participant and personnel	Blinding of outcome assessment	Incomplete outcome data	Selective reporting
D'Souza, DC	low	uncertain	low	low	low	uncertain
Geisel, O	uncertain	uncertain	low	low	low	uncertain

4 Considerações finais

Esta é a primeira meta-análise a avaliar o BDNF periférico em indivíduos com TUS comparando com controles saudáveis. Nossos resultados indicaram que os níveis séricos de BDNF encontram-se diminuídos durante o uso ativo de substâncias psicoativas, todavia houve variações de acordo com o tipo e a classe de substância e, também, sofreu a influência do sexo, do envelhecimento, dos anos de uso de substâncias e do tempo de abstinência. Além disso, durante a abstinência não houve diferenças entre casos e controles, o que sinaliza que o BDNF, possivelmente, possa normalizar-se com interrupção do uso de SPAs, entretanto, isso pode ser afetada pela gravidade e pela cronicidade do transtorno. Também foi verificada a existência de diferenças entre os níveis séricos e plasmáticos do BDNF, talvez, devido a diferenças metodológicas de mensuração e do fato do BDNF das duas matrizes ser proveniente de locais diferentes, com meia vida distinta.

Nossos resultados corroboram a hipótese que os TUS podem ocasionar alterações neuroprogressivas que afetam, de forma persistente, a neuroplasticidade cerebral - o que parece relacionar-se ao processo de transição do uso recreativo para o a dependência. Neste sentido, o uso agudo de drogas acarretaria o aumento do BDNF; o uso continuado geraria alterações disfuncionais que, em casos mais graves, podem ser irreversíveis mesmo com a abstinência. Sabe-se que os TUS destacam-se pela baixa adesão e altos índices de recaída. Nesse sentido, a identificação de biomarcadores não invasivos, como o BDNF, pode auxiliar na compreensão fisiopatológica dos transtornos, permitindo mensurar o estado patológico e a gravidade da doença e, assim, através da proposição de intervenções mais assertivas, aumentar a adesão do tratamento e a sua efetividade.

Durante a análise dos estudos incluídos no presente trabalho, evidenciamos uma alta heterogeneidade entre os mesmos, principalmente no que se refere a diferenças amostrais e metodológicas. Logo, a realização de uma meta-análise permitiu amenizar as discrepâncias observadas, devido ao fato dessa análise combinar resultados de estudos independentes, controlar fatores de moderação e tamanhos de efeito, potencializar o poder estatístico e proporcionar a formulação de conclusões mais abrangentes.

Em isolado, nossos resultados não permitem inferir que a utilização do BDNF possa se dar na prática clínica, entretanto, funde-se com um amplo corpo de evidências que reforçam a hipótese neurodegenerativa decorrente do uso crônico de SPAS. Assim, espera-se que estes resultados, somados a estudos que investiguem a fisiopatologia dos TUS através do uso de outros potenciais biomarcadores - como o uso da neuroimagem, cortisol, NPY, análise genética, possibilitem o desenvolvimento de tratamentos mais direcionados e específicos.

Referências

1. Kessler RC, McGonagle KA, Zhao S, Nelson CB, Hughes M, Eshleman S, et al. Lifetime and 12-month prevalence of DSM-III-R psychiatric disorders in the United States. Results from the National Comorbidity Survey. *Arch Gen Psychiatry*. 1994;51(1):8-19.
2. Compton WM, Thomas YF, Conway KP, Colliver JD. Developments in the epidemiology of drug use and drug use disorders. *Am J Psychiatry*. 2005;162(8):1494-502.
3. Regier DA, Farmer ME, Rae DS, Locke BZ, Keith SJ, Judd LL, et al. Comorbidity of mental disorders with alcohol and other drug abuse. Results from the Epidemiologic Catchment Area (ECA) Study. *JAMA*. 1990;264(19):2511-8.
4. Kessler RC, Chiu WT, Demler O, Merikangas KR, Walters EE. Prevalence, severity, and comorbidity of 12-month DSM-IV disorders in the National Comorbidity Survey Replication. *Arch Gen Psychiatry*. 2005;62(6):617-27.
5. The world health report 2001: mental health: new understanding new hope. World Health Organization. Geneva; WHO; 2001. XVIII,178 p.
6. WHO report on the global tobacco epidemic, 2015: raising taxes on tobacco. World Health Organization. Genebra; WHO; 2015. 198 p.
7. World Health Organization - WHO. Global status report on alcohol and health, 2014Geneva; World Health Organization; 2014. 376 p.
8. IBGE. Instituto Brasileiro de Geografia e Estatística - IBGE. Pesquisa nacional de saúde 2013: ciclos de vida: Brasil e grandes regiões. Rio de Janeiro; IBGE; 2015. 85 p. 2015.
9. Abdalla RR, Madruga CS, Ribeiro M, Pinsky I, Caetano R, Laranjeira R. Prevalence of cocaine use in Brazil: data from the II Brazilian national alcohol and drugs survey (BNADS). *Addict Behav*. 2014;39(1):297-301.
10. van der Meer Sanchez Z, Nappo SA. From the first drug to crack: the sequence of drugs taken in a group of users in the city of São Paulo. *Subst Use Misuse*. 2007;42(1):177-88.
11. Falck RS, Wang J, Carlson RG. Crack cocaine trajectories among users in a midwestern American city. *Addiction*. 2007;102(9):1421-31.
12. van der Meer Sanchez Z, Nappo SA. [Progression on drug use and its intervening factors among crack users]. *Rev Saude Publica*. 2002;36(4):420-30.
13. Crime UNODC. World Drug Report 2016. 2016.
14. Laranjeira R, Madruga CS. II Levantamento Nacional de Álcool e Drogas (LENAD) –2012. São Paulo: Cromosete Gráfica e Editora Ltda; 2012.

15. Duailibi L, Ribeiro M, Laranjeira R. Profile of cocaine and crack users in Brazil. *Cadernos de Saúde Pública* [Internet]. 2008;[545-57 pp.].
16. Siliquini R, Morra A, Versino E, Renga G. Recreational drug consumers: who seeks treatment? *Eur J Public Health*. 2005;15(6):580-6.
17. McLellan AT, Starrels JL, Tai B, Gordon AJ, Brown R, Ghitza U, et al. Can Substance Use Disorders be Managed Using the Chronic Care Model? Review and Recommendations from a NIDA Consensus Group. *Public Health Rev*. 2014;35(2).
18. World Health Organization (WHO) Lexicon of Alcohol and Drug Terms Published by the World Health Organization. 2006. [accessed 2016]. Available at: http://www.who.int/substance_abuse/terminology/who_lexicon/en/.
19. Wikler A. Dynamics of drug dependence. Implications of a conditioning theory for research and treatment. *Arch Gen Psychiatry*. 1973;28(5):611-6.
20. APA APA-. Manual Diagnóstico e Estatístico de Transtornos Mentais (DSM-5)2014.
21. World Health Organization (WHO). International Classification of Diseases (ICD-10). [Internet] 2013 Available from: <http://www.who.int/classifications/icd/en/index.html>. 2013.
22. Diagnostic and Statistical Manual of Mental disorders - DSM - 5 . 5th.ed. Washington: American Psychiatric Association (APA), 2013. DSM -IV-TR T^M.
23. Samet S, Waxman R, Hatzenbuehler M, Hasin DS. Assessing addiction: concepts and instruments. *Addict Sci Clin Pract*. 2007;4(1):19-31.
24. Mueser KT, Noordsy DL, Drake RE, Fox L. [Integrated treatment for severe mental illness and substance abuse: Effective components of programs for persons with co-occurring disorders.]. *Sante Ment Que*. 2001;26(2):22-46.
25. Koob GF, Volkow ND. Neurocircuitry of addiction. *Neuropsychopharmacology*. 2010;35(1):217-38.
26. Robinson TE, Berridge KC. The neural basis of drug craving: an incentive-sensitization theory of addiction. *Brain Res Brain Res Rev*. 1993;18(3):247-91.
27. Robinson TE, Berridge KC. Incentive-sensitization and addiction. *Addiction*. 2001;96(1):103-14.
28. Leshner AI. Addiction is a brain disease, and it matters. *Science*. 1997;278(5335):45-7.
29. Koob GF. Neurobiology of addiction. Toward the development of new therapies. *Ann N Y Acad Sci*. 2000;909:170-85.
30. Sinha R. New findings on biological factors predicting addiction relapse vulnerability. *Curr Psychiatry Rep*. 2011;13(5):398-405.

31. Goldstein RZ, Volkow ND. Drug addiction and its underlying neurobiological basis: neuroimaging evidence for the involvement of the frontal cortex. *Am J Psychiatry*. 2002;159(10):1642-52.
32. Seger D. Cocaine, metamfetamine, and MDMA abuse: the role and clinical importance of neuroadaptation. *Clin Toxicol (Phila)*. 2010;48(7):695-708.
33. Mendelson J, Baggott MJ, Flower K, Galloway G. Developing biomarkers for methamphetamine addiction. *Curr Neuropharmacol*. 2011;9(1):100-3.
34. Büttner A. Review: The neuropathology of drug abuse. *Neuropathol Appl Neurobiol*. 2011;37(2):118-34.
35. Bough KJ, Amur S, Lao G, Hemby SE, Tannu NS, Kampman KM, et al. Biomarkers for the development of new medications for cocaine dependence. *Neuropsychopharmacology*. 2014;39(1):202-19.
36. Volkow ND, Koob G, Baler R. Biomarkers in substance use disorders. *ACS Chem Neurosci*. 2015;6(4):522-5.
37. Di Chiara G, Imperato A. Drugs abused by humans preferentially increase synaptic dopamine concentrations in the mesolimbic system of freely moving rats. *Proc Natl Acad Sci U S A*. 1988;85(14):5274-8.
38. Olive MF, Koenig HN, Nannini MA, Hodge CW. Stimulation of endorphin neurotransmission in the nucleus accumbens by ethanol, cocaine, and amphetamine. *J Neurosci*. 2001;21(23):RC184.
39. Di Chiara G, Bassareo V. Reward system and addiction: what dopamine does and doesn't do. *Curr Opin Pharmacol*. 2007;7(1):69-76.
40. Koob GF. Drugs of abuse: anatomy, pharmacology and function of reward pathways. *Trends Pharmacol Sci*. 1992;13(5):177-84.
41. Le Foll B, Gallo A, Le Strat Y, Lu L, Gorwood P. Genetics of dopamine receptors and drug addiction: a comprehensive review. *Behav Pharmacol*. 2009;20(1):1-17.
42. Di Chiara G. Drug addiction as dopamine-dependent associative learning disorder. *Eur J Pharmacol*. 1999;375(1-3):13-30.
43. Koob GF. Neurobiological substrates for the dark side of compulsion in addiction. *Neuropharmacology*. 2009;56 Suppl 1:18-31.
44. Volkow ND, Wang GJ, Fischman MW, Foltin R, Fowler JS, Franceschi D, et al. Effects of route of administration on cocaine induced dopamine transporter blockade in the human brain. *Life Sci*. 2000;67(12):1507-15.
45. Volkow ND, Fowler JS, Wang GJ. Role of dopamine in drug reinforcement and addiction in humans: results from imaging studies. *Behav Pharmacol*. 2002;13(5-6):355-66.

46. Wise RA. Dopamine, learning and motivation. *Nat Rev Neurosci.* 2004;5(6):483-94.
47. Volkow ND, Fowler JS, Wang GJ, Swanson JM. Dopamine in drug abuse and addiction: results from imaging studies and treatment implications. *Mol Psychiatry.* 2004;9(6):557-69.
48. Volkow ND, Fowler JS, Wang GJ, Swanson JM, Telang F. Dopamine in drug abuse and addiction: results of imaging studies and treatment implications. *Arch Neurol.* 2007;64(11):1575-9.
49. Volkow ND, Wang GJ, Fowler JS, Tomasi D, Telang F. Addiction: beyond dopamine reward circuitry. *Proc Natl Acad Sci U S A.* 2011;108(37):15037-42.
50. Koob GF, Bloom FE. Cellular and molecular mechanisms of drug dependence. *Science.* 1988;242(4879):715-23.
51. Kalivas PW, O'Brien C. Drug addiction as a pathology of staged neuroplasticity. *Neuropsychopharmacology.* 2008;33(1):166-80.
52. Olsen CM. Natural rewards, neuroplasticity, and non-drug addictions. *Neuropsychopharmacology.* 2011;61(7):1109-22.
53. George O, Le Moal M, Koob GF. Allostasis and addiction: role of the dopamine and corticotropin-releasing factor systems. *Physiol Behav.* 2012;106(1):58-64.
54. Robinson TE, Berridge KC. Review. The incentive sensitization theory of addiction: some current issues. *Philos Trans R Soc Lond B Biol Sci.* 2008;363(1507):3137-46.
55. Volkow ND, Wang GJ, Fowler JS, Tomasi D, Telang F, Baler R. Addiction: decreased reward sensitivity and increased expectation sensitivity conspire to overwhelm the brain's control circuit. *Bioessays.* 2010;32(9):748-55.
56. Thomas MJ, Kalivas PW, Shaham Y. Neuroplasticity in the mesolimbic dopamine system and cocaine addiction. *Br J Pharmacol.* 2008;154(2):327-42.
57. Russo SJ, Mazei-Robison MS, Ables JL, Nestler EJ. Neurotrophic factors and structural plasticity in addiction. *Neuropsychopharmacology.* 2009;56 Suppl 1:73-82.
58. Sinha R. Chronic stress, drug use, and vulnerability to addiction. *Ann N Y Acad Sci.* 2008;1141:105-30.
59. Cramer SC, Sur M, Dobkin BH, O'Brien C, Sanger TD, Trojanowski JQ, et al. Harnessing neuroplasticity for clinical applications. *Brain.* 2011;134(Pt 6):1591-609.
60. Group. BDW. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clin Pharmacol Ther.* 2001;69(3):89-95.

61. D'Sa C, Fox HC, Hong AK, Dileone RJ, Sinha R. Increased serum brain-derived neurotrophic factor is predictive of cocaine relapse outcomes: a prospective study. *Biol Psychiatry*. 2011;70(8):706-11.
62. Heberlein A, Muschler M, Wilhelm J, Frieling H, Lenz B, Gröschl M, et al. BDNF and GDNF serum levels in alcohol-dependent patients during withdrawal. *Prog Neuropsychopharmacol Biol Psychiatry*. 2010;34(6):1060-4.
63. Bartkowska K, Turlejski K, Djavadian RL. Neurotrophins and their receptors in early development of the mammalian nervous system. *Acta Neurobiol Exp (Wars)*. 2010;70(4):454-67.
64. Barde YA, Edgar D, Thoenen H. Purification of a new neurotrophic factor from mammalian brain. *EMBO J*. 1982;1(5):549-53.
65. Kuipers SD, Bramham CR. Brain-derived neurotrophic factor mechanisms and function in adult synaptic plasticity: new insights and implications for therapy. *Curr Opin Drug Discov Devel*. 2006;9(5):580-6.
66. Janak PH, Wolf FW, Heberlein U, Pandey SC, Logrip ML, Ron D. BIG news in alcohol addiction: new findings on growth factor pathways BDNF, insulin, and GDNF. *Alcohol Clin Exp Res*. 2006;30(2):214-21.
67. Ciammola A, Sassone J, Cannella M, Calza S, Poletti B, Frati L, et al. Low brain-derived neurotrophic factor (BDNF) levels in serum of Huntington's disease patients. *Am J Med Genet B Neuropsychiatr Genet*. 2007;144B(4):574-7.
68. Angelucci F, Spalletta G, di Iulio F, Ciaramella A, Salani F, Colantoni L, et al. Alzheimer's disease (AD) and Mild Cognitive Impairment (MCI) patients are characterized by increased BDNF serum levels. *Curr Alzheimer Res*. 2010;7(1):15-20.
69. Lee JG, Shin BS, You YS, Kim JE, Yoon SW, Jeon DW, et al. Decreased serum brain-derived neurotrophic factor levels in elderly korean with dementia. *Psychiatry Investig*. 2009;6(4):299-305.
70. Yasutake C, Kuroda K, Yanagawa T, Okamura T, Yoneda H. Serum BDNF, TNF-alpha and IL-1beta levels in dementia patients: comparison between Alzheimer's disease and vascular dementia. *Eur Arch Psychiatry Clin Neurosci*. 2006;256(7):402-6.
71. Autry AE, Monteggia LM. Brain-derived neurotrophic factor and neuropsychiatric disorders. *Pharmacol Rev*. 2012;64(2):238-58.
72. Hashimoto K, Shimizu E, Iyo M. Critical role of brain-derived neurotrophic factor in mood disorders. *Brain Res Brain Res Rev*. 2004;45(2):104-14.
73. Hashimoto K. Brain-derived neurotrophic factor as a biomarker for mood disorders: an historical overview and future directions. *Psychiatry Clin Neurosci*. 2010;64(4):341-57.

74. Fernandes BS, Gama CS, Ceresér KM, Yatham LN, Fries GR, Colpo G, et al. Brain-derived neurotrophic factor as a state-marker of mood episodes in bipolar disorders: a systematic review and meta-regression analysis. *J Psychiatr Res.* 2011;45(8):995-1004.
75. Carlini EA, Galduroz JCF, Noto AR, Nappo SA. I Levantamento domiciliar sobre o uso de drogas psicotrópicas no Brasil : estudo envolvendo as 107 maiores cidades do país 2002.
76. Serra-Millàs M. Are the changes in the peripheral brain-derived neurotrophic factor levels due to platelet activation? *World J Psychiatry.* 2016;6(1):84-101.
77. Fernandes BS, Berk M, Turck CW, Steiner J, Gonçalves CA. Decreased peripheral brain-derived neurotrophic factor levels are a biomarker of disease activity in major psychiatric disorders: a comparative meta-analysis. *Mol Psychiatry.* 2014;19(7):750-1.
78. Kawamoto Y, Nakamura S, Nakano S, Oka N, Akitoshi I, Kimura J. Immunohistochemical localization of brain-derived neurotrophic factor in adult rat brain. *Neuroscience.* 1996;74(4):1209-26.
79. Dugich-Djordjevic MM, Peterson C, Isono F, Ohsawa F, Widmer HR, Denton TL, et al. Immunohistochemical visualization of brain-derived neurotrophic factor in the rat brain. *Eur J Neurosci.* 1995;7(9):1831-9.
80. Bibel M, Barde YA. Neurotrophins: key regulators of cell fate and cell shape in the vertebrate nervous system. *Genes Dev.* 2000;14(23):2919-37.
81. Binder DK, Scharfman HE. Brain-derived neurotrophic factor. *Growth Factors.* 2004;22(3):123-31.
82. Cohen-Cory S, Kidane AH, Shirkey NJ, Marshak S. Brain-derived neurotrophic factor and the development of structural neuronal connectivity. *Dev Neurobiol.* 2010;70(5):271-88.
83. Karege F, Schwald M, Cisse M. Postnatal developmental profile of brain-derived neurotrophic factor in rat brain and platelets. *Neurosci Lett.* 2002;328(3):261-4.
84. Pan W, Banks WA, Fasold MB, Bluth J, Kastin AJ. Transport of brain-derived neurotrophic factor across the blood-brain barrier. *Neuropharmacology.* 1998;37(12):1553-61.
85. Karege F, Bondolfi G, Gervasoni N, Schwald M, Aubry JM, Bertschy G. Low brain-derived neurotrophic factor (BDNF) levels in serum of depressed patients probably results from lowered platelet BDNF release unrelated to platelet reactivity. *Biol Psychiatry.* 2005;57(9):1068-72.
86. Sartorius A, Hellweg R, Litzke J, Vogt M, Dormann C, Vollmayr B, et al. Correlations and discrepancies between serum and brain tissue levels of neurotrophins after electroconvulsive treatment in rats. *Pharmacopsychiatry.* 2009;42(6):270-6.

87. Klein AB, Williamson R, Santini MA, Clemmensen C, Ettrup A, Rios M, et al. Blood BDNF concentrations reflect brain-tissue BDNF levels across species. *Int J Neuropsychopharmacol.* 2010;14(3):347-53.
88. Rasmussen P, Brassard P, Adser H, Pedersen MV, Leick L, Hart E, et al. Evidence for a release of brain-derived neurotrophic factor from the brain during exercise. *Exp Physiol.* 2009;94(10):1062-9.
89. Lee BH, Kim YK. Reduced platelet BDNF level in patients with major depression. *Prog Neuropsychopharmacol Biol Psychiatry.* 2009;33(5):849-53.
90. Yamamoto H, Gurney ME. Human platelets contain brain-derived neurotrophic factor. *J Neurosci.* 1990;10(11):3469-78.
91. Donovan MJ, Miranda RC, Kraemer R, McCaffrey TA, Tessarollo L, Mahadeo D, et al. Neurotrophin and neurotrophin receptors in vascular smooth muscle cells. Regulation of expression in response to injury. *Am J Pathol.* 1995;147(2):309-24.
92. Nakahashi T, Fujimura H, Altar CA, Li J, Kambayashi J, Tandon NN, et al. Vascular endothelial cells synthesize and secrete brain-derived neurotrophic factor. *FEBS Lett.* 2000;470(2):113-7.
93. Fujimura H, Altar CA, Chen R, Nakamura T, Nakahashi T, Kambayashi J, et al. Brain-derived neurotrophic factor is stored in human platelets and released by agonist stimulation. *Thromb Haemost.* 2002;87(4):728-34.
94. Lommatsch M, Schloetke K, Klotz J, Schuhbaeck K, Zingler D, Zingler C, et al. Brain-derived neurotrophic factor in platelets and airflow limitation in asthma. *Am J Respir Crit Care Med.* 2005;171(2):115-20.
95. Corominas M, Roncero C, Ribases M, Castells X, Casas M. Brain-derived neurotrophic factor and its intracellular signaling pathways in cocaine addiction. *Neuropsychobiology.* 2007;55(1):2-13.
96. McGinty JF, Whitfield TW, Jr., Berglind WJ. Brain-derived neurotrophic factor and cocaine addiction. *Brain Res.* 2010;1314:183-93.
97. McGinty JF, Mendelson JE. Is brain-derived neurotrophic factor a selective biomarker that predicts cocaine relapse outcomes? *Biol Psychiatry.* 2011;70(8):700-1.
98. Davis MI. Ethanol-BDNF interactions: still more questions than answers. *Pharmacol Ther.* 2008;118(1):36-57.
99. Czubak A, Nowakowska E, Kus K, Burda K, Metelska J, Baer-Dubowska W, et al. Influences of chronic venlafaxine, olanzapine and nicotine on the hippocampal and cortical concentrations of brain-derived neurotrophic factor (BDNF). *Pharmacol Rep.* 2009;61(6):1017-23.
100. Angelucci F, Gruber SH, El Khoury A, Tonali PA, Mathé AA. Chronic amphetamine treatment reduces NGF and BDNF in the rat brain. *Eur Neuropsychopharmacol.* 2007;17(12):756-62.

101. Chu NN, Zuo YF, Meng L, Lee DY, Han JS, Cui CL. Peripheral electrical stimulation reversed the cell size reduction and increased BDNF level in the ventral tegmental area in chronic morphine-treated rats. *Brain Res.* 2007;1182:90-8.
102. Lu L, Dempsey J, Liu SY, Bossert JM, Shaham Y. A single infusion of brain-derived neurotrophic factor into the ventral tegmental area induces long-lasting potentiation of cocaine seeking after withdrawal. *J Neurosci.* 2004;24(7):1604-11.
103. Graham DL, Edwards S, Bachtel RK, DiLeone RJ, Rios M, Self DW. Dynamic BDNF activity in nucleus accumbens with cocaine use increases self-administration and relapse. *Nat Neurosci.* 2007;10(8):1029-37.
104. Berglind WJ, See RE, Fuchs RA, Ghee SM, Whitfield TW, Jr., Miller SW, et al. A BDNF infusion into the medial prefrontal cortex suppresses cocaine seeking in rats. *Eur J Neurosci.* 2007;26(3):757-66.
105. Fumagalli F, Di Pasquale L, Caffino L, Racagni G, Riva MA. Repeated exposure to cocaine differently modulates BDNF mRNA and protein levels in rat striatum and prefrontal cortex. *Eur J Neurosci.* 2007;26(10):2756-63.
106. Grimm JW, Lu L, Hayashi T, Hope BT, Su TP, Shaham Y. Time-dependent increases in brain-derived neurotrophic factor protein levels within the mesolimbic dopamine system after withdrawal from cocaine: implications for incubation of cocaine craving. *J Neurosci.* 2003;23(3):742-7.
107. Heberlein A, Muschler M, Wilhelm J, Frieling H, Lenz B, Groschl M, et al. BDNF and GDNF serum levels in alcohol-dependent patients during withdrawal. *Prog Neuropsychopharmacol Biol Psychiatry.* 2010;34(6):1060-4.
108. Costa MA, Girard M, Dalmay F, Malauzat D. Brain-derived neurotrophic factor serum levels in alcohol-dependent subjects 6 months after alcohol withdrawal. *Alcohol Clin Exp Res.* 2011;35(11):1966-73.
109. Huang MC, Chen CH, Liu SC, Ho CJ, Shen WW, Leu SJ. Alterations of serum brain-derived neurotrophic factor levels in early alcohol withdrawal. *Alcohol Alcohol.* 2008;43(3):241-5.
110. Shim SH, Hwangbo Y, Kwon YJ, Jeong HY, Lee BH, Lee HJ, et al. Increased levels of plasma brain-derived neurotrophic factor (BDNF) in children with attention deficit-hyperactivity disorder (ADHD). *Prog Neuropsychopharmacol Biol Psychiatry.* 2008;32(8):1824-8.
111. Huang MC, Chen CH, Liu HC, Chen CC, Ho CC, Leu SJ. Differential patterns of serum brain-derived neurotrophic factor levels in alcoholic patients with and without delirium tremens during acute withdrawal. *Alcohol Clin Exp Res.* 2010;35(1):126-31.
112. Cavus SY, Dilbaz N, Darcin AE, Eren F, Kaya H, Kaya O. Alterations in serum BDNF levels in early alcohol withdrawal and comparison with healthy controls. *Bulletin of Clinical Psychopharmacology [Internet].* 2012; 22:[210-5 pp.]. Available from:

113. Corominas-Roso M, Roncero C, Jose Eiroa-Orosa F, Gonzalvo B, Grau-Lopez L, Ribases M, et al. Brain-derived neurotrophic factor serum levels in cocaine-dependent patients during early withdrawn. *Eur Neuropsychopharmacol.* 2012.
114. von Diemen L, Kapczinski F, Sordi AO, de Magalhães Narvaez JC, Guimarães LS, Kessler FH, et al. Increase in brain-derived neurotrophic factor expression in early crack cocaine withdrawal. *Int J Neuropsychopharmacol.* 2014;17(1):33-40.
115. Heberlein A, Dursteler-MacFarland KM, Lenz B, Frieling H, Grosch M, Bonsch D, et al. Serum levels of BDNF are associated with craving in opiate-dependent patients. *J Psychopharmacol.* 2011;25(11):1480-4.
116. Kim DJ, Roh S, Kim Y, Yoon SJ, Lee HK, Han CS, et al. High concentrations of plasma brain-derived neurotrophic factor in methamphetamine users. *Neurosci Lett.* 2005;388(2):112-5.
117. Angelucci F, Ricci V, Pomponi M, Conte G, Mathe AA, Attilio Tonali P, et al. Chronic heroin and cocaine abuse is associated with decreased serum concentrations of the nerve growth factor and brain-derived neurotrophic factor. *J Psychopharmacol.* 2007;21(8):820-5.
118. Chen PH, Huang MC, Lai YC, Chen PY, Liu HC. Serum brain-derived neurotrophic factor levels were reduced during methamphetamine early withdrawal. *Addict Biol.* 2012.
119. Angelucci F, Ricci V, Spalletta G, Pomponi M, Tonioni F, Caltagirone C, et al. Reduced serum concentrations of nerve growth factor, but not brain-derived neurotrophic factor, in chronic cannabis abusers. *Eur Neuropsychopharmacol.* 2008;18(12):882-7.
120. Angelucci F, Ricci V, Martinotti G, Palladino I, Spalletta G, Caltagirone C, et al. Ecstasy (MDMA)-addicted subjects show increased serum levels of brain-derived neurotrophic factor, independently from a rise of drug-induced psychotic symptoms. *Addict Biol.* 2010;15(3):365-7.
121. Corominas-Roso M, Roncero C, Daigre C, Grau-Lopez L, Ros-Cucurull E, Rodríguez-Cintas L, et al. Changes in brain-derived neurotrophic factor (BDNF) during withdrawn could be associated with relapse in cocaine-dependent patients. *Psychiatry Res.* 2015;225(3):309-14.
122. Sordi AO, Pechansky F, Kessler FH, Kapczinski F, Pfaffenseller B, Gubert C, et al. Oxidative stress and BDNF as possible markers for the severity of crack cocaine use in early withdrawal. *Psychopharmacology (Berl).* 2014;231(20):4031-9.
123. Huang MC, Chen CH, Liu HC, Chen CC, Ho CC, Leu SJ. Differential patterns of serum brain-derived neurotrophic factor levels in alcoholic patients with and without delirium tremens during acute withdrawal. *Alcohol Clin Exp Res.* 2011;35(1):126-31.
124. Scherer JN, Schuch S, Ornell F, Sordi AO, Kessler FHP, von, et al. Higher levels of BDNF are associated with inpatient treatment adherence of crack-cocaine users. *Drug and Alcohol Dependence.* 2015.

Anexo 1 – Projetos em andamento

- Preditores Clínicos, Biológicos e Psicossociais da Recaída Precoce em usuários de crack.
- Projeto cocaínas fumáveis na Argentina, Brasil, Chile, Uruguai e Paraguai. Estudo multicêntrico sobre alterações da função cerebral em usuários de crack.
- Ensaio Clínico Randomizado, Duplo-Cego, Controlado com Placebo, para Avaliar o Efeito da N-Acetilcisteína no Tratamento dos Transtornos por Uso de Álcool e Cocaína.
- Associação da intervenção de cue exposure sobre os níveis de cortisol salivar em dependentes de crack internados no Hospital De Clínicas de Porto Alegre.
- Associação entre o estado nutricional e níveis de leptina e grelina em pacientes internados em uma unidade de adição.
- Vulnerabilidade social em usuários de crack em seis capitais brasileiras
- Fatores de personalidade e emocionais entre usuários de substâncias psicoativas

Anexo 2. Artigos em desenvolvimento:

- Fator Neurotrófico Derivado Do Cérebro (Bdnf) no Transtorno por Uso de Álcool – Uma Meta-análise (título provisório)
- Fator Neurotrófico Derivado Do Cérebro (Bdnf) no Transtorno por Uso de Cocaína – Uma Meta-análise (título provisório)
- Fatores associados a variação nos níveis séricos de BDNF em usuários de crack/cocaína
- Por que é tão difícil fazer pesquisa com usuários de crack (título provisório)
- Diferenças entre nas dosagens do BDNF após 5 anos de congelamento (título provisório)

Anexo 3 – Artigos publicados durante o período do Mestrado

1. **Ornell, Felipe**; Dotta, R. M. ; Scherer, J. ; Dal Cin, V. ; Modena, S. L. ; Halpern, S. C. . Saúde e cárcere: estruturação da atenção básica à saúde no sistema prisional do Rio Grande do Sul. *Sistema Penal & Violência* (Online), v. 8, p. 107, 2016.
2. Scherer, Juliana N. ; Schuch, Silvia ; **Ornell, Felipe** ; Sordi, Anne ; Bristot, Giovana ; Pfaffenseller, Bianca ; Kapczinski, Flávio ; Kessler, Felix H.P. ; Fumagalli, Fabio ; Pechansky, Flavio ; von Diemen, Lisia . HIGH LEVELS OF BRAIN-DERIVED NEUROTROPHIC FACTOR ARE ASSOCIATED WITH TREATMENT ADHERENCE AMONG CRACK-COCAINE USERS. *Neuroscience Letters (Print)*, p. 169-175, 2016.
3. Scherer, Juliana Nichterwitz ; Silvestrin, Roberta ; **Ornell, Felipe** ; Roglio, Vinícius ; Sousa, Tanara Rosangela Vieira ; von Diemen, Lisia ; Kessler, Felix Henrique Paim ; Pechansky, Flavio . Prevalence of driving under the influence of psychoactive substances and road traffic crashes among Brazilian crack-using drivers. *Drug and Alcohol Dependence*, p. 255-262, 2016.
4. Steckert, Amanda V. ; Comim, Clarissa M. ; Igna, Dhébora M. Dall ; Dominguini, Diogo ; Mendonça, Bruna P. ; **Ornell, Felipe** ; Colpo, Gabriela D. ; Gubert, Carolina ; Kapczinski, Flávio ; Barichello, Tatiana ; Quevedo, João ; Dal-Pizzol, Felipe . Effects of sodium butyrate on aversive memory in rats submitted to sepsis. *Neuroscience Letters (Print)*, v. 595, p. 134-138, 2015.

Anexo 4 – Artigos submetidos durante o período do Mestrado

1. Avaliação Neuropsicológica das Funções Executivas e do Controle Inibitório nos Transtornos Por Uso de Álcool e Crack

Autores: Fernanda Rasch Czermainski, **Felipe Ornell**, Luciano Santos Pinto Guimarães, Félix Kessler, Lízia von Diemen e Rosa Maria Martins de Almeida.
Revista: Psico, submetido em janeiro de 2016.

2. Histórico de situação de rua como marcador de vulnerabilidades entre usuários de crack em seis capitais brasileiras

Autores: Silvia C. Halpern, Juliana Nichterwitz Scherer, **Felipe Ornell**, Carla Dalbosco, Sibele Faller, Vinicius Roglio, Félix Kessler, Flavio Pechansky, Lízia Von Diemen .

Revista: Cadernos de Saúde Pública, submetido em março de 2017.

3. Infrational Act and the inter-relationship with psychic trauma and drug abuse

Autores: Magda Maria Rodrigues Ferreira Valadares, Laís Rodrigues Valadares, Felipe Ornell, Vinícius Serafini Roglio, Juliana Nichterwitz Scherer, Felix Henrique Paim Kessler, Silvia Chwartzmann Halpern.

Revista: Trends in Psychology/Temas em Psicologia, 2016

4. Psychiatric disorders in aesthetic medicine: The importance of recognition of signs and symptoms

Autores: Juliana Nichterwitz Scherer, Felipe Ornell, Joana C. M. Narvaez, Rafael Ceita Nunes

Revista: Revista Brasileira de Cirurgia Plástica, 2017

Anexo 5 – Capítulos de livros escritos durante o período do Mestrado

1. Comorbidities Associated With the Use and Misuse of Crack Cocaine
Autores: Narvaez, Joana; **Ornell, Felipe**; Kessler, Felix; von Diemen, Lisia; Magalhães, Pedro.
Livro: The Neuroscience of Cocaine: Mechanisms And Tretment.
2. Brain-derived neurotrophic factor in cocaine withdrawal
Autores: Giannotti, Giuseppe; Scherer , Juliana N; **Ornell, Felipe**; Caffino, Lucia; Fumagalli Fabio; von Diemen Lisia.
Livro: The Neuroscience of Cocaine: Mechanisms And Tretment.

Anexo 6 – Prêmios recebidos durante o período do Mestrado

1. Melhor trabalho da categoria Iniciação Científica – Centro de Estudos Luís Guedes – CELG, 2016 – coorientação.
2. Destaque na categoria Psiquiatria Clínica – Centro de Estudos Luís Guedes – CELG, 2016 – coautor.
3. Prêmio Jovens Cientistas, 21st International Council on Alcohol, Drugs and Traffic Safety Conference, 2016 – Coorientação.
4. Destaque XXVIII Salão de Iniciação Científica, UFRGS, 2016 – Coorientação.
5. Destaque da 35ª Semana Científica do Hospital de Clínicas de Porto Alegre, 2015 – Coorientação