UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL

Faculdade de Farmácia

Disciplina de Trabalho de Conclusão de Curso de Farmácia

Lonomia obliqua venom activity upon extracellular matrix

Alessandra Selinger Magnusson

Porto Alegre, dezembro de 2013.

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# Sumário

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# Apresentação

Este trabalho de conclusão de curso será apresentado sob forma de artigo científico, sendo elaborado segundo as normas da revista científica "Toxicon" (fator de impacto segundo o *Journal Citation Reports* - 2012 = 2,924) conforme documentação apresentada no anexo 1.

# Lonomia obliqua venom activity upon extracellular matrix

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# Abstract

Lonomia obligua is a medically important caterpillar endemic in South Brazil. Contact with theis caterpillar's bristles causes an envenomation syndrome characterized by ecchymosis and hemorrhage. The Lonomia obligua bristle extract (LOBE) is mainly composed by highly active proteases known to interfere with the hemostatic system of the victims. Although the effects of the venom enzymes upon blood coagulation and platelet aggregation is well characterized, it is possible that LOBE might also affect the environment surrounding small blood vessels, favoring hemorrhage and impairing wound healing. In this work we evaluated the proteolytic activity of LOBE upon extracellular matrix (ECM) components. Degradation of extracellular matrix proteins was performed by in vitro digestion of purified proteins (collagen, laminin, fibronectin), and matrigel and fragments were analyzed by SDS-PAGE. Additionally, in vivo ECM organization of envenomed rat skin was investigated by immunohistochemistry and hystochemistry. Fibroblast cell migration and wound healing was evaluated by the cell scratch assay. LOBE was able to degrade all tested ECM substrates in a timedependent way. It was also observed impairment on fibroblast migratory behavior. Analysis of envenomed skin showed an increase on inflammatory cells in the connective tissue surrounding the local of LOBE injection, which was accompanied by changes on collagen organization and degradation of laminin on blood vessel basement membrane. In conclusion, Lonomia obliqua bristle extract has a proteolytic activity on extracellular matrix proteins, which might play a role on local and systemic hemorrhage and the appearance of ecchymosis.

Key Words: metaloprotease, caterpillar venom, laminin, collagen, fibronectin

# 1 Introduction

Venomous animals is the term that denominates animals capable of producing poison or poisonous secretions. There are several classes including thousands of species like snakes, spiders, frogs, caterpillars, scorpions and others. These animals are worldwide distributed and are cause of accidents with humans in tropical and subtropical regions, where the abundance of species is greater. World Health Organization has included poisoning by snake bite as a recognized neglected tropical disease, for the first time in 2009, showing the importance of studies in this area. (Williams, et al., 2010).

Animal poisons are unique and complexes mixtures of proteins and bioactive peptides. These physic-chemicals and biological properties resulted from evolutionary process of species providing such individuals with effective means of protection for defense and feeding on prey and predators. The animal toxins are able to interact with enzymes, receptors and ion channels causing destabilization of physiological systems that are essentials to the victims and prey survival. (Calvete, 2009).

The larvae or caterpillars are larval forms from butterflies and moths (Order Lepidoptera) and accidents involving caterpillars occur in all Brazilian territory. However, the South region is the one which presents the higher incidence of accidents, 7.3/100.000 inhabitants. The most important occurrences are with caterpillars that belong to the genus *Lonomia*. From 4,028 accidents caused by these animals occurred in 2009, 14.5% were caused by *Lonomia* (585/4.028) (SVS, 2010). The specie *Lonomia Obliqua* is the most relevant specie in the South of Brazil (Gamborgi, et al. 2012).

Caterpillar, unlike other venous animals, does not have a specialized gland for poison production, being this used only for defense, not for hunting and feeding. These animals have a secretory epithelium responsible for production of a poisonous secretion which is injected in the victims when the contact breaks the spikes. (Veigas, et al. 2001)

Between two and seventy two hours after a contact the symptoms appear. Bruising and bleeding being characterized as epistaxis, otorrhagia, rectal bleeding, menorrhagia and widespread bruising are the most common symptoms. Besides, in severe cases, the progress of the disease can lead to acute renal failure and intracerebral bleeding. (Duarte, et al. 1990, Kellen, et al. 1995; Duarte, et al. 1996). Pro-clotting manifestations can occur in addition to the bleeding symptoms. (Reis, et al. 1999; Zannin, et al. 2003).

Rats envenomed with *L. obliqua* venom show important biochemical, hematological and histopathological alterations, suggesting the occurrence of multiorgan damage and confirming that the rat is a good animal model for studying hemorrhagic disturbances, as well as organ specific injuries. (Berger, et. al. 2013).

Snake venom metalloproteinases hydrolyze key peptide bonds in basement membrane (BM) and supporting extracellular matrix (ECM) components, promoting the weakening of the mechanical stability of BM. As a consequence, the hemodynamic biophysical forces normally operating in the vasculature, such as microvessel wall tension and shear stress, provoke the distention of the weakened capillary, which leads to microvessel disruption and extravasation (Gutierrez, et al., 2005)

The extracellular matrix degradation or remodeling activities exerted by these toxins affect cell–cell and cell–extracellular matrix adhesion and survival and impair inflammatory cell migration into inflamed tissues. (Komegae, et al. 2011). These effects of peptidases on extracellular matrix (ECM) proteins interfere with the normal processes of tissue regeneration and angiogenesis, contributing to tissue damage observed in envenomations. Moreover, the degradation of ECM can induce cells to enter into the apoptotic pathway known as anoikis as consequence of loss of signal transduction between matrix and cells due to a physical disruption of integrin–matrix interactions (Frisch and Screaton, 2001; Grossmann, 2002).

In this work we evaluated the ability of *L. obliqua* bristle extract in degrade extracellular matrix (ECM) components, favoring hemorrhage and impairing wound healing.

# 2 Material and Methods

# 2.1 *Lonomia obliqua* venomous secretions

*L. obliqua* caterpillars were kindly provided by Centro de Informações Toxicológicas (CIT), Porto Alegre, Rio Grande do Sul, Brazil. *L. obliqua* bristle extract was obtained by cutting bristles at the caterpillar's tegument insertion and the excised material was kept at 4°C until preparation of the extract, which occurred immediately after dissection. Bristles were macerated in cold saline solution (150 mM NaCl) and centrifuged at 9600g for 20 min. The protein content of the supernatant, designed as crude *L obliqua* bristle extract (LOBE), was determined by the BCA assay kit (Pierce,Rockford,USA) and aliquots were stored at -80°C until use. For the experiments, four different preparations of bristle extract were used and the total number of caterpillars varied between 50 and 80 specimens per preparation.

# 2.2 Experimental animals

Male Wistar rats (weighing 300–350 g) were housed in a temperature-controlled room (21–25°C, in a 12-h light/ dark cycle), with free access to water and food. All procedures involving animals were carried out in accordance with the Guiding Principles in the Use of Animals in Toxicology (International Society of Toxicology, http://www.toxicology. org) and the Brazilian College of Animal Experimentation (COBEA). Experiments also followed the recommendations of Ethical Committee for the

Use of Animals of Federal University of Rio Grande do Sul, Brazil. All efforts were made to minimize the number of animals used and their suffering.

Time course of experimental envenomation In order to follow the evolution of platelet and blood coagulation disturbances we developed an experimental model of envenomation in rats (Berger. et al. 2010). Animals were divided into two groups: an experimental group, which was injected subcutaneously (s.c.) with 1mL of a solution containing 1 µg protein of crude LOBE per kg of body weight, and a control group that received subcutaneously 150 mM NaCl in a volume of 1mL, under the same conditions. Two hours after injection, rats were anesthetized and sacrificed. Skin and muscle from the injection site were collected for histological evaluation. A total of 8 animals were utilized in each group.

# 2.3 Histopathological analyses

All animals from the control and envenomed groups (at each sampling time) were necropsied and gross macroscopic alterations were examined. The skin, muscle and endothelium (at the site of venom injection) were then carefully removed and fixed in a 10% neutral buffered formaldehyde solution. The tissues were dehydrated in gradual alcohol, cleared in xylene and embedded in paraplast. Subsequently, the samples were sectioned and stained with hematoxylin and eosin (H&E), Masson's trichrome and Picrossirius for further analysis by light microscopy.

# 2.4 Immunohistochemistry

Paraplast embedded samples were cut ( $3\mu$ m), mounted on slides, deparaffinized in xylene and hydrated in ethanol solutions and 0.1M phosphate-saline buffer (PBS), pH 7.4, room temperature (RT). The endogenous peroxidase activity was inactivated by incubation with 0.3% H<sub>2</sub>O<sub>2</sub> in Methanol (10min, RT). For antigen recovery, slides were

heated in 0.01 M citrate buffer in a water bath at 95°C (pH 6.0) for 15 min. The sections were incubated with primary antibodies anti-laminin (Sigma, polyclonal antibody, 1:100, 4°C, overnight), washed with PBS, incubated with HRP-dual link (Dako) and visualized with DAB chromogen (3',3-diaminobenzidine,Dako).

# 2.5 Extracellular matrix (ECM) protein degradation

Fibronectin (Sigma), laminin (Sigma), matrigel (Sigma) was incubated with LOBE (10:1 ratio) in Tris-HCl buffer (20 mM, pH 7.4, 37°C). After specified time points (15min, 60min,120min, 240min and 24h), samples were boiled (95°C, 5 min), precipited by acetone precipitation, submitted to a 10% SDS-PAGE gel and stained for Comassie blue for analysis of ECM degradation.

# 2.6 Wound healing assay

NIH 3T3 cells were cultured near confluence, starved overnight and a scratch was performed with a plastic tip. Media were replaced by complete media supplemented with LOBE and the wound clousure was measured after 4 or 8h.

# 2.7 Angiogenesis assay

It was performed *in vivo* by the ECM gel/sponge assay. Foam sponge soaked in ECM gel in the presence or absence of LOBE was implanted intradermally in rats. After 12 days, the sponge was surgically recovered, macerated in water and blood vessels content was measured by hemoglobin quantification at 540nm.

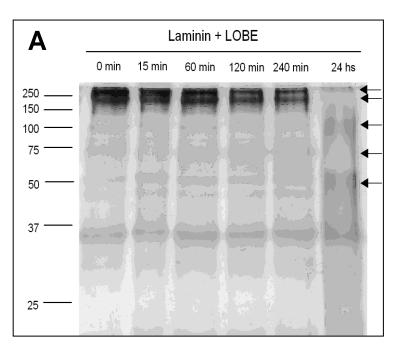
# 2.8 Statistical analysis

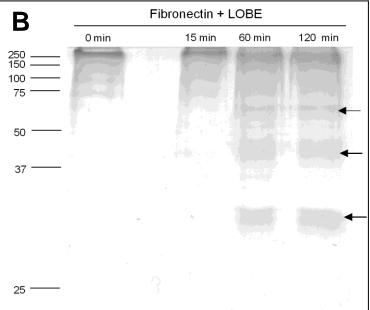
Values are expressed as mean an standard deviation e data was submitted to Student ttest.

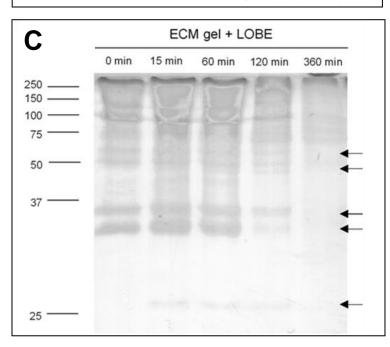
# 3. Results

# 3.1 LOBE induces in vitro degradation of ECM proteins

Laminin incubated without LOBE showed two main bands of 250 and 200kDa. Partial degradation was observed after 24h incubation with LOBE by the presence of a low-molecular-weight fragment of 100, 75 and 50 kDa. (Fig. 1A). The LOBE also hydrolyzed fibronectin as evidenced by the appearance of new protein bands. After 60 and 120 minutes of incubation the high-molecular band was completely digest by LOBE resulting in several degradation products (Fig. 1B). Matrigel or ECM gel (gelatinous protein mixture secreted by Engelbreth-Holm-Swarm (EHS) murine sarcoma cells) also was verified a decreases in concentration after reaction with ECM, after 60, 120 and 360min. (Fig.1C).



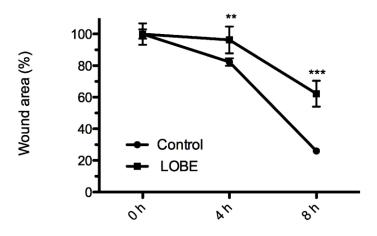




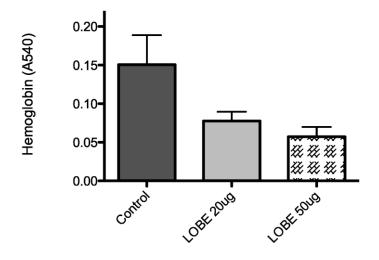
**Figure 1** Proteolytic degradation of laminin (A), fibronectin (B) and extracellular matrix derived matrix (C). Samples were incubated with LOBE (10:1 ratio) and analyzed by SDS–PAGE 10%. Arrow indicates the degradation product of reaction.

# 3.2 LOBE decreases cell migration and angiogenesis

Impairment of cell migration and wound healing was evaluated in vitro by the cell scratch assay. The LOBE induced an impairment on the cell migration process (Fig. 2). The *in vivo* analysis for angiogenesis demonstrated that LOBE induced a decrease on blood vessel formation when compared to the control group (Fig.3).



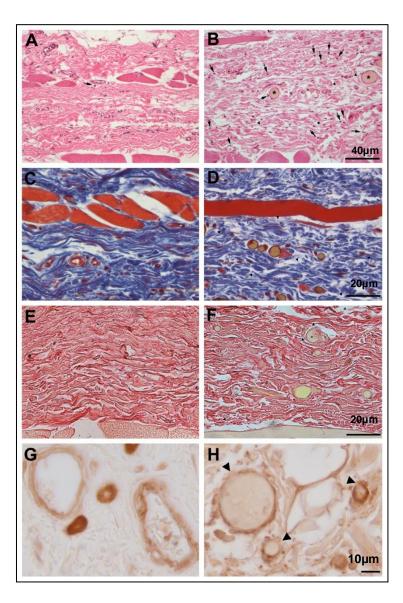
**Figure 2-** The LOBE decreases cells migration after 4h and 8h compared to the control group. NIH 3T3 fibroblasts were submitted to wound healing assay in the presence or absence of LOBE (1  $\mu$ g/ml). Student T-test (\*\* p<0.05, \*\*\* p<0.01 n=3)



**Figure 3:** *Lonomia Obliqua* venom decreases angiogenesis *in vivo.* Control animals received saline. Animals were sacrificed 12 days after implanted intradermally with LOBE 20µg and 50µg. Angiogenesis assay dermination in ECM gel/ sponge hemoglobin in the samples is quantified calorimetrically at 540 nm in spectrophotometer.

# 3.3 LOBE induces ECM remodeling in vivo

Skin morphological analysis showed that after 2h of LOBE administration there was the presence of inflammatory cells in the connective tissue surrounding envenomed region as well the presence of blood clot inside capilary and venule vessels (Fig 4B) when compared to skin exposed to PBS (Fig 4A). Masson Trichrome (Fig 4C and 4D) and Picrossirius (Fig 4E and 4F) staining showed dissociation of collagen fibers in the dermis due to marked edema after LOBE envenomation (Fig. 4D) and changes in the collagen organization and degradation (Fig. 4F). In control rats (Fig 4G), it was observed laminin staining on the basement membrane of blood vessel and, 2h after LOBE administration, there was a more punctuate distribution of this protein, specially on venous vessels (Fig 4H).



**Fig.4** Histologic alterations after L. obliqua envenomation in rats. Representative images of: **A**, **C**, **E**, **F** control animal (one that had been injected with PBS solution, s.c.), showing normal morphology. **B**, **D**, **F**, **H** the animal that had been injected with LOBE (1 mg/kg, s.c.) after 2 h of envenomation. **B**. Hematoxilin/eosin (HE). **D**. Masson Trichrome and **F**. Picrossirius **H**. IHC of Laminin.

# 4. Discussion

The *L. obliqua* venom, at low and non-hemorragic doses, exerts a direct proinflammatory effect on endothelial cells, promoting cytoskeleton reorganization, increasing focal adhesion and the expression of crucial molecules to the onset of a vascular inflammatory response. (Nascimento-Silva, et al. 2012). The poisons have been studied with the objective of better understand its structure and functionality, as well as their protein toxins individually. Consequently, understanding the mechanism and origin of the toxic effect. This is accomplished by isolation of proteins, followed by its biochemical characterization, and then determining the biological activity *in vitro* and *in vivo*. (Gallagher, et al. 2003) The venom are comprises several families of proteins, especially metalloproteases, serine proteases, phospholipases, disintegrins, lectins, and L-amino oxidase .For each family there are a number of isoforms of these proteins that act synergistically with high specificity, composing, on the whole, the biochemical basis of the processes leading to the development of the pathological. (Terra, et al. 2009).

In this study, we demonstrated that LOBE administration is able to alter cell migration and the process of angiogenesis. The venom component of bristle extract of *L. Obliqua*: prothrombin activator (Lopap) acts on the cell preservation, since it regulates the expression of molecules, such as nitric oxide (NO) and prevents cell death. (Fritzen, et al. 2005; Reis, et al. 1999). As well, the component Losac (a factor X activato) that besides its pro-coagulant activity, also functions as a growth factor and an inhibitor of cell death of endothelial cells. Possibly, TPA (tissue plasminogen activator) and NO stimulated by Losac, induces cell preservation, since the release of TPA and NO can inhibit the proliferation and induce apoptosis of endothelial cells. So that both are related to angiogenesis. (Alvarez-Flores, et al. 2006).

New blood vessel growth, or angiogenesis, occurs during the development and maturation as well as during critical physiological processes, including wound healing and reproduction. However, angiogenic processes are also usurped in many pathologies, and thereby contribute to an array of disorders including cancer, inflammation and autoimmune diseases (Folkman, 2006). Following regression of native vessels, degradation of the vessel basement membrane and surrounding ECM to allow for invasion of endothelial cells is an integral part of the ongoing angiogenic process. After regression of existing vessels and breakdown of the basement membrane, endothelial cells proliferate and begin migrating. In toward tumor cells expressing pro-angiogenic compounds. (Carmeliet & Jain, 2000, 2011). The LOBE decreases the formation of new vessels and also cause rupture of the existent.

Extracellular matrices are composed of dynamic and highly ordered structural networks of collagens, proteoglycans and noncollagenous glycoproteins. The temporal and spatial regulation of ECM formation contributes to the mechanical and biological properties of different connective tissues and has fundamental effects on cellular phenotypes (Nelson and Bissell, 2006). ECM components also regulate the local release of growth factors and play an important role on cell differentiation, migration and adhesion by providing linkages between cells and the ECM (Heinegard, et al. 2002). In this work, we demonstrated *in vitro* and *in vivo* that LOBE has a proteolitic acitivy upon collagen, laminin and fibronectin substrates. It was already reported that metalloproteinases and serine protease-like presented in *L. achelous* hemolymph also can degrade extracellular matrix proteins such as laminin, vitronectin and fibronectin. These toxins were identified as chromatographic fractions FDII, Lonomin V and Lonomin V-2. These toxins may contribute to the hemorrhagic events triggered by *L. achelous* venom because they may facilitate the spreading of the venom through the victim's body and aggravate hemorrhage by destroying the capillaries. (Lucena, et al.

2006). *In vitro* assays that use high protease to substrate ratios have shown that MMPs can cleave a wide range of ECM molecules. This has led to the widely-held concept that MMP mediated cleavage of ECM molecules is the means by which infiltrating cells, including leukocytes and transformed cells, penetrate ECM barriers and that MMPs are essential for all remodelling processes in developing or regenerating tissues. (Sorokin, 2010).

Basement membrane constitutes a complex extracellular matrix scaffold, composed mainly of collagen IV, laminin, nidogen and the proteoglycan perlecan, that provides support to endothelial cells in blood vessels and plays various roles in the communication between intracellular and extracellular environments (Martin and Timpl, 1987). Interactions between laminin, Fibronectin and collagen, and other components of basement membranes are crucial for both biological and mechanical properties of the skin (von der Mark, et al. 1992). In this work, we demonstrated evidence of laminin degradation on basement membrane of small blood vessels of envenomed skin. This data suggest that the proteolytic activity of LOBE might play an important role during the hemorrhage process. It is also possible that during the ECM remodeling induced by LOBE there is a release of small peptides that might work as signaling and chemoattractive molecules for leukocytes, as observed by HE staining. In fact, it was already showed that several molecules resulting from laminin cleavage have this property. (Sorokin, 2010, Nascimento-Silva, et al. 2012). The fibronectin analysis confirmed the degradatory activity of venom upon this glycoprotein, even when it was organized on the extracellular matrix produced by endothelial cells. (Paludo, et al., 2006)

# 5. Conclusion

Accidents with *Lonomia obliqua* are mostly characterized by bleeding as consequence of a severe hemorrhagic syndrome caused by a complex coagulopathic process that can easily evolve to death if not treated in the very beginning of the accident occurrence. Since the caterpillar venom is not synthesized by a gland, the toxic compounds are frequently derived from a mixture of a secretion produced by the specialized epithelium.

The LOBE caused morphological alterations. It was demonstrated that the LOBE caused Laminin, fibronectin and collagen degradation, observed *In vitro* and *In vivo*. It happened diapedesis and the rupture of basal membrane of small blood vessels. Beyond, a decrease on fibroblast migration and angiogenesis. Degradation of ECM by *L. obliqua* proteases may be a key factor in facilitating systemic hemorrhage and the appearance of ecchymosis.

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# Conflict of interest statement

The authors declare that there are no conflicts of interest.

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ISSN: 0041-0101

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Toxicon's "aims and scope" are laid down in the journal as:

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articles containing the results of original research on problems related to toxins derived from animals, plants and microorganisms papers on novel findings related to the **chemical**, **pharmacological**, **toxicological**, and **immunological** properties of natural **toxins molecular biological** studies of toxin and other **genes** from **poisonous** and **venomous** organisms that advance understanding of the role or function of toxins clinical observations on poisoning and envenoming where a new therapeutic principle has been proposed or a decidedly superior clinical result has been obtained material on the use of toxins as tools in studying biological processes and material on subjects related to **venom** and **antivenom** problems review articles on problems related to **toxinology**.

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Mettam, G.R., Adams, L.B., 2009. How to prepare an electronic version of your article, in: Jones, B.S., Smith , R.Z. (Eds.), Introduction to the Electronic Age. E-Publishing Inc., New York, pp. 281–304.

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