LUNG AND LIVER CHANGES DUE TO THE INDUCTION OF CIRRHOSIS IN TWO EXPERIMENTAL MODELS

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ABSTRACT - Context - To evaluate lung and liver changes in two experimental models using intraperitoneal carbon tetrachloride (CCl4) and bile duct ligation (BDL). Methods - Twenty-four male Wistar rats were divided into a control group (CO) and an experimental group (EX). We evaluated the liver transaminases (AST, ALT, AP), arterial blood gases (PaO2, PCO2, and SpO2) and lipid peroxidation by TBARS (substances that react to thiobarbituric acid) and chemiluminescence. We also evaluated the antioxidant enzyme superoxide dismutase (SOD) and histology of lung tissue and liver. Results - There were significant differences in AST, ALT, ALP and PaO2 between CO group and EX group (P<0.05). The levels of TBARS, chemiluminescence and activity of enzyme superoxide dismutase were increased to different degrees in the CCl4 groups: CO and in the BDL -EX (P<0.05, respectively). In the lung histology, an increase in the wall thickness of the pulmonary artery and a diameter reduction in the CCl4 animal model were observed: comparing CO group with EX group, we observed a reduction in thickness and an increase in the diameter of the artery wall lung. Conclusion - Both experimental models have caused liver damage and alterations in the artery wall that are associated with major changes in pulmonary gas exchange.


INTRODUCTION

Fibrosis is an important process that results from hepatic injury and chronic disease process. Both of these causes can have common etiologies that can contribute to the onset of cirrhosis(2). Cirrhosis is considered to be the most advanced stage of tissue fibrosis and is characterized by the disruption of hepatic parenchyma, the appearance of the septa and fibrotic nodules, changes in hepatic blood flow and a risk of liver failure10. Cirrhosis can be caused by altered vascular tissue, which causes a shift in the supply of blood and portal blood flow directly to the liver (central vein). The altered vascular tissue compromises the hepatic sinusoids and hepatocytes. Some of the circulatory changes found in cirrhosis are splanchnic vasodilatation, vasoconstriction and renal hypoperfusion, salt and water retention and increased cardiac output, which are all closely linked to vascular disease of the liver that may cause portal hypertension11, 25. Portopulmonary hypertension (PPH) is a relatively rare complication occurring in cirrhosis, and its prevalence is high in patients with refractory ascites. PPH is mostly likely caused by excessive pulmonary vasoconstriction and factors such as transforming growth factor beta1 (TGF-β1)41. However, we do not have a perfect experimental model for the study of PPH, making the discovery of effective substances that can reduce and even prevent the progression of the disease impossible. Developing an experimental model is very useful, as such a model could lead to substantial cost savings in treatment. Carbon tetrachloride (CCl4) can be applied both intraperitoneally and by inhalation. CCl4 induces fibrosis liver cirrhosis and may be used for the study of PPH. The signals observed in the CCl4 liver injury model are similar to those found in cirrhotic patients19. The CCl4 trichloromethyl radical is converted to (°CCL3) and trichloromethyl peroxide (°OOCCl3). The CCl4 trichloromethyl radical has been described as causing hepatotoxic effects such as fibrosis, steatosis, nerosis and hepatocellular carcinoma19, 22. Bile duct ligation is another experimental model related to secondary biliary cirrhosis because it causes cell proliferation, hepatocellular necrosis, apoptosis, activation of stellate cells, and finally the formation of liver fibrosis and cirrhosis. This model has been described by our group as causing Experimental Hepatopulmonary Syndrome28, 29. Given the complications of cirrhosis, the aim of this study was to evaluate two types of pulmonary and...
hepatic injury that cause liver cirrhosis, a CCl4 model and a BDL model, to evaluate which model can best reproduce characteristics found in PPH.

METHODS

Animals and experimental groups

We used 24 male Wistar rats (average weight 250 g), purchased from the Center for Reproduction and Laboratory Animal Experimentation (CREAL), Federal University of Rio Grande do Sul (UFRGS). The animals were housed in plastic cages (47×34×18 cm) and maintained in light/dark cycles of 12/12 hs with a controlled temperature (20-25 °C) and ad libitum access to food and water. All of the procedures followed the parameters established by the Ethics and Research of Hospital Clinics of Porto Alegre, and all of the animals received care according to the “Principles of Laboratory Animal Care” formulated by the National Institutes for Medical Research and the “Guide for the Care and Use of Laboratory Animals” published by the National Institutes of Health (NIH publication 86-23, revised 1985).

The animals were randomly divided into four groups, an experimental and a control group (CO), for each of the two models used in the study, intraperitoneal Carbon tetrachloride (CCl4) and bile duct ligation (BDL):

- **CCl4-CO**: animals that received only phenobarbital in drinking water;
- **CCl4-EX**: animals that received phenobarbital in drinking water one week before starting the injections of CCl4 inductions + ip;
- **BDL-CO**: animals that underwent laparotomy and only manipulation of the duct biliar.
- **BDL-EX**: animals subjected to BDL.

Experimental procedures

To develop the CCl4 model, we used the standard recommended by Pavanato et al. (21). Phenobarbital was added to the drinking water of animals (0.3 mL/dL), serving as an enzyme inducer to potentiate the effect of CCl4 (60).

To develop the BDL model, we followed the standard described by Kontouras et al. (14) where, after the experimental procedures, the animals were caged individually for a period of 28 days.

When sacrificing animals from either group, the animals were anesthetized with xylazine (50 mg/kg body weight) and ketamine (100 mg/kg body weight), both administered intraperitoneally. Post mortem, blood samples were collected via retro-orbital plexus for liver function analysis (12).

After the abdominal region was shaved, a laparotomy was performed to collect blood from the abdominal aorta for arterial blood gas analysis. After collection of blood samples, the animals were sacrificed with an “overdose” of anesthetics.

Histology

For histological analysis of the lung, a fragment of the right lower lobe was collected for histological analysis, and a fragment of the liver was removed from the right hepatic lobe. The tissues were cut and fixed by immersion in 10% buffered formalin for 24 hours. The remains of lung and liver were immediately frozen in liquid nitrogen and stored at -80°C for later analysis.

Intracellular liver enzymes

Blood samples taken from the retro-orbital plexus were used to assess the levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase (AP) and were expressed in IU/L and measured with routine laboratory methods in the Hospital Clinics of Porto Alegre.

**Determination of oxidative stress**

The liver tissue and lung of each animal were homogenized in phosphate buffer solution (140 mM KCl, 20 mM phosphate, pH 7.4) and centrifuged at 3,000 rpm for 10 minutes. Lipid peroxidation was determined by levels of thiobarbituric acid reactive substances (thiobarbituric acid reactive substances - TBARS) mg/protein read at 535 nm. For the determination of chemiluminescence (CL), 0.5 mL of homogenate was added to 120 mM KCl, 30 mM phosphate buffer, pH 7.4, and 3 mM tert-butyl hydroperoxide at 30°C and analyzed for chemiluminescence in a counter liquid scintillation expressed as cps/mg/protein (12, 18). **Superoxide dismutase enzyme activity**

The activity of the antioxidant enzyme superoxide dismutase (SOD) was determined by quantifying the oxidation of adrenaline to adrenochrome according to Misra and Fridovich and was expressed in U/mg/protein (53).

Statistical analysis

The results were expressed as mean ± standard error. Data were compared by analysis of variance (ANOVA), and if the analysis indicated a statistically significant difference, we used a post hoc Student Newman-Keuls test. The statistical significance level was set at P<0.05. The Statistical Package for Social Sciences, version 13.0 (SPSS Inc., Chicago, IL, USA), was used.

RESULTS

Activity of transaminases

Table 1 indicates a significant difference between the CCl4 and CCl4-EX groups and between BDL and BDL-CO-EX in serum AST, ALT and AP, which were higher in the experimental groups compared to the controls (P<0.05).

Arterial blood gas analysis

To verify changes in gas exchange, we performed arterial blood gas analysis (Table 1) in which the variables PaO2 (mmHg), PCO2 (mmHg) and SO2 (%) showed changes in both experimental models. We can see that the animals belonging to the experimental groups (CCl4-EX and BDL-EX) showed significant differences when compared to their
TABLE 1. Activity of transaminases and arterial blood gas analysis

<table>
<thead>
<tr>
<th>Variable</th>
<th>CCl₄ - CO</th>
<th>CCl₄ - EX</th>
<th>BDL - CO</th>
<th>BDL - EX</th>
</tr>
</thead>
<tbody>
<tr>
<td>AST (IU/L)</td>
<td>119±37</td>
<td>5407±2592*</td>
<td>169±107</td>
<td>348±148*</td>
</tr>
<tr>
<td>ALT (IU/L)</td>
<td>39±7</td>
<td>3306±1308</td>
<td>53±11</td>
<td>213±324*</td>
</tr>
<tr>
<td>FA (IU/L)</td>
<td>67±20</td>
<td>307±88*</td>
<td>116±45</td>
<td>213±324*</td>
</tr>
<tr>
<td>PaO₂ (mmHg)</td>
<td>106,91±22,01</td>
<td>68,98±17,46*</td>
<td>95,10±17,00</td>
<td>5,97±14,55*</td>
</tr>
<tr>
<td>PaCO₂ (mmHg)</td>
<td>49,85±10,30</td>
<td>55,06±6,04</td>
<td>46,07±7,03</td>
<td>41,40±12,38</td>
</tr>
<tr>
<td>SaO₂ (%)</td>
<td>96,56±3,53</td>
<td>86,65±18,71*</td>
<td>96,52±1,67</td>
<td>93,17±9,72</td>
</tr>
</tbody>
</table>

CCl₄: Carbon tetrachloride; BDL: bile duct ligation; CO: control; EX: experimental; AST: aspartate aminotransferase; ALT: alanine aminotransferase; AP: alkaline phosphatase; PaO₂: pressure oxygen; PaCO₂: carbon dioxide partial pressure; SaO₂: oxygen saturation. Results expressed as mean ± standard deviation. * P<0.05 corresponding control group.

TABLE 2. Evaluation of lipid peroxidation in the lung tissue (TBARS e QL) and antioxidant enzyme SOD

<table>
<thead>
<tr>
<th>Model</th>
<th>TBARS (nmol/mg prot)</th>
<th>QL (cps/mg prot)</th>
<th>SOD (IU/mg prot)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CCl₄</td>
<td>CO 0,56±0,22</td>
<td>16764,20±515,48</td>
<td>11,24±4,99</td>
</tr>
<tr>
<td></td>
<td>EX 1,48±0,52*</td>
<td>17797,20±531,65</td>
<td>27,27±1,23*</td>
</tr>
<tr>
<td>BDL</td>
<td>CO 0,835±0,16</td>
<td>16008,41±1171,45</td>
<td>6,66±1,34</td>
</tr>
<tr>
<td></td>
<td>EX 1,91±0,91*</td>
<td>20250,36±827,82*</td>
<td>16,06±2,67*</td>
</tr>
</tbody>
</table>

TBARS: thiobarbituric acid reactive substances; QL: chemiluminescence; SOD: superoxide dismutase; CO: control; EX: experimental. Results expressed as mean ± standard deviation. * P<0.05 corresponding control group.

respective controls (P<0.05). Only the variable PCO₂ showed no significant difference between the CO and EX animals in the BDL model.

Markers of Oxidative Stress and Antioxidant Activity
Analysis of lipid peroxidation in lung tissue (Table 2) it appears that TBARS as well as high CL in two models are used in the study (P<0.05). Regarding the antioxidant enzyme SOD, a significant increase (P<0.05) in both experimental models was observed compared to control animals.

Histology
On the photomicrograph of the lung tissue (Figure 1), we observed an increase of 47.87 mM in wall thickness in

Figure 1. Histology of the pulmonary tissue
Photomicrograph of the lung tissue in the models experimental. a) CCl₄-CO; b) CCl₄-EX; c) BDL-CO; d) BDL-EX
the pulmonary artery model CCl4 as compared to the CO. However, when analyzing the histology of the pulmonary tissue of the animal model BDL, we observed an increase of 69.67 mM in vascular diameter when compared to controls. The photomicrograph of the liver (Figure 2) revealed the presence of necrotic foci, fibrotic nodules, lymphocytic infiltration, steatosis and cellular changes, suggesting that cirrhosis was present in the two models.

DISCUSSION

In an attempt to develop a specific model for the study of PPH, we used two experimental models developed in our laboratory that cause cirrhosis.

The CCl4 has been used to reproduce liver cirrhosis due to its potent hepatotoxic effect in which a single dose of CCl4 is associated with steatosis and necrosis but a prolonged administration leads to liver fibrosis, cirrhosis and hepatocellular carcinoma. CCl4 acts directly on hepatocytes by changing the permeability of the plasma and mitochondrial membranes. This model has been widely used to elucidate the pathogenesis of cirrhosis(11).

Secondary biliary cirrhosis is a chronic and diffuse liver disease that alters intrahepatic or extrahepatic biliary function. The BDL model is used as an animal model for secondary biliary cirrhosis and cirrhosis caused by progressive and fatal damage to the liver. This model simulates the effects of the disease present in humans, causing changes in the inflammatory reaction by leaking bile and the subsequent disorganization of parenchymal inflammation, collagen deposition and formation of fibrosis(7, 13, 20).

Studies by Kontouras et al.(14) showed that 15 days after bile duct ligation (BDL), the metabolic and biochemical changes were not fully established. Thus, in our experiment, the animals were sacrificed only after 28 days of BDL, when studies confirmed liver cirrhosis(17, 29).

In this study, both the BDL and the CCl4 caused liver damage and elevated serum levels of AST, ALT and ALP, which are commonly known to be markers of liver injury. These changes in liver enzyme levels indicate a loss of tissue integrity with consequent apoptosis and necrosis of hepatocytes. These enzymes are eventually released into circulation after cellular damage, and this finding has been reported in several studies using models of BDL and CCl4(3, 27, 31).

Patients who have secondary biliary cirrhosis exhibit hyperbilirubinemia due to biliary obstruction resulting from liver damage and present results of liver function tests in higher ranges for FA and aminotransferases. High levels of these enzymes are largely caused by the necrosis of tissues that are rich in aminotransferases(1, 7).

In the presence of liver cirrhosis, there is an increase in lipid peroxidation due to the formation of reactive oxygen species and lipid peroxidation. Some authors report that these phenomena determine the changes in the structures and cell membranes by two mechanisms, covalent binding to macromolecules and cell action on lipids, and that these two mechanisms are the result of the metabolism of CCl4 by cytochrome P-450, which produces two radicals which are highly toxic and CCl4, CCl3O2••(28) Additionally, reactive oxygen species (ROS) are involved in the BDL model.

Cholestasis in the BDL model reduces the excretion of bile salts, causing the retention of hydrophobic bile salts within hepatocytes and apoptosis and necrosis, the destruction of liver parenchyma, which contributes to a redox imbalance, and the formation of ROS. In addition, ROS cause oxidation of cellular proteins and extensive damage to mitochondrial DNA and mitochondrial synthesis by liver damage(8, 24, 34).

When we evaluated lipid peroxidation in the two models of cirrhosis, animals in the EX group had significantly higher values compared to those in the CO group, and the level of TBARS and QL were elevated in the lung and liver in both groups. This overproduction of ROS might occur in several pathophysiological situations and may explain the finding that in our study, the levels of TBARS and QL were high(22, 23).

Physiological defense mechanisms are effective in preventing or counteracting the damage caused by free radicals, which can include a set of endogenous antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx). The two models used in this study showed an increase in lipid peroxidation in lung tissue and are likely to increase in response to an increase of ROS in the SOD, which plays an important role in the balance of the redox cell, as the catalyst free radicals are generated in an attempt to protect the tissue against lesions(1, 2, 7).
Arterial blood gas analysis is an indicator used to facilitate the evaluation of trade gasosas\(^2\)\(^{20}\). In our study, there was a significant difference between the experimental and control groups with respect to arterial blood gases. We found that \(\text{PaO}_2\) and \(\text{SpO}_2\) were significantly lower in the \(\text{CCl}_4\) group. In the BDL model, the values of \(\text{SpO}_2\) were also reduced, although this did not reach statistical significance.

Approximately 85% of PPH patients show an increase in pulmonary vascular resistance in the course of the disease, of which, 15% experience increases in their pulmonary pressures that are caused by just a state of high flux\(^{15}\).

The relationship between arterial hypoxemia and PPH has received little attention because PPH is a clinical problem that involves an altered hemodynamic status. PPH patients exhibit arterial hypoxemia and arterial oxygenation that is significantly worse when comparing liver transplantation candidates with normal vascular resistance as determined by echocardiography Doppler\(^{16}\).

In the BDL model, hypoxemia may be associated with bacterial translocation occurring in 45%-75% of cirrhotic animals whereby the induced hepatocellular damage prevents adequate blood filtration and promotes the development of portosystemic shunts; this phenomenon dramatically decreases the phagocytic ability of the liver, and it also allows the entry of bacteria and endotoxins into the lung circulation\(^{23}\).

Furthermore, studies have reported arterial hypoxemia in BDL animals, which contributes to a reduction in the values of \(\text{PaO}_2\) due to increased volume of lung shunt\(^{32}\).

The differences found in this study between the BDL and \(\text{CCl}_4\) gas analyses showed that regardless of etiology, animals with cirrhosis are hypoxic because the values of \(\text{PaO}_2\) and \(\text{SpO}_2\) were lower in both experimental models. This hypoxemia might be triggered by the activation of macrophages as well as the presence of cytokines and nitric oxide in the extracellular medium. Nitric oxide is a potent vasodilator that promotes dilation of intrapulmonary vessels, leading to hypoxemia. Nitric oxide may also be involved in the mechanisms that cause an imbalance between the antioxidants and oxidants\(^{29}\).

Regarding photomicrographs of the lung tissue, the BDL model caused an increase in diameter of arterioles of the lung tissue. This finding corroborates studies reporting the increase of animals subjected to BDL\(^{29}\). This result may be due to the increased intrapulmonary shunt caused by liver injury leading to an accumulation of blood in the body, which in turn dilates the arterioles of the lung tissue and releases nitric oxide. In the \(\text{CCl}_4\) model, the observed reduction of these structures can be explained by the fact that cirrhotic \(\text{CCl}_4\) animals had increased levels of ET-1 and collagen deposition, promoting the reduction in thickness of lung tissue.

The histology of liver tissue revealed damage to liver cells and confirmed the changes observed in biochemical analyses. The presence of necrotic foci, fibrotic nodules, the infiltration of lymphocytes, and the fatty changes in liver cells are typical characteristic after the induction of cirrhosis\(^{45, 33}\).

The two experimental models used in this study reinforce and confirm the onset of liver damage and cirrhosis of the liver with consequent changes in gas exchange. The BDL model is most often used to evaluate the Hepatopulmonary Syndrome because it shows changes that are typical characteristic of the disease. The \(\text{CCl}_4\) model seems to be promising for the study of portopulmonary hypertension (PPH) as well as cirrhosis and changes the thickness and diameter of the pulmonary artery wall. More studies are needed to assess and investigate the specific mechanisms involved in these changes.

**CONCLUSION**

The model of hepatic cirrhosis caused by \(\text{CCl}_4\) modifies the gas exchange, increases the thickness of the pulmonary artery and reduces its diameter. The \(\text{CCl}_4\) model is a promising model for the study of PPH. The BDL model also causes alterations in the lung and liver, reinforcing that is a reliable model of HPS.

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